

The MILE study: Expression microarray analysis for diagnosis of leukaemia

Ken Mills

CCRCB

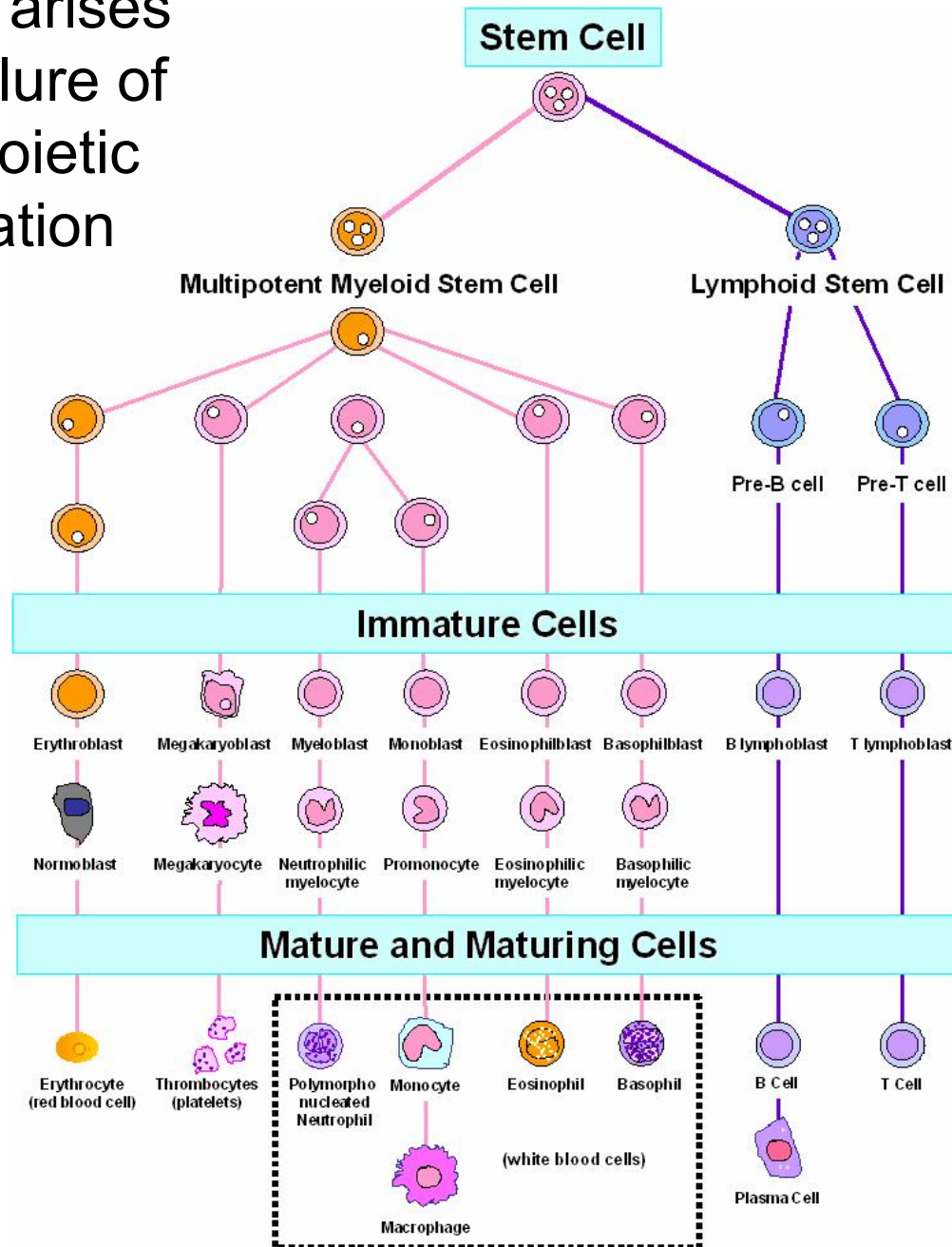
Queen's University Belfast

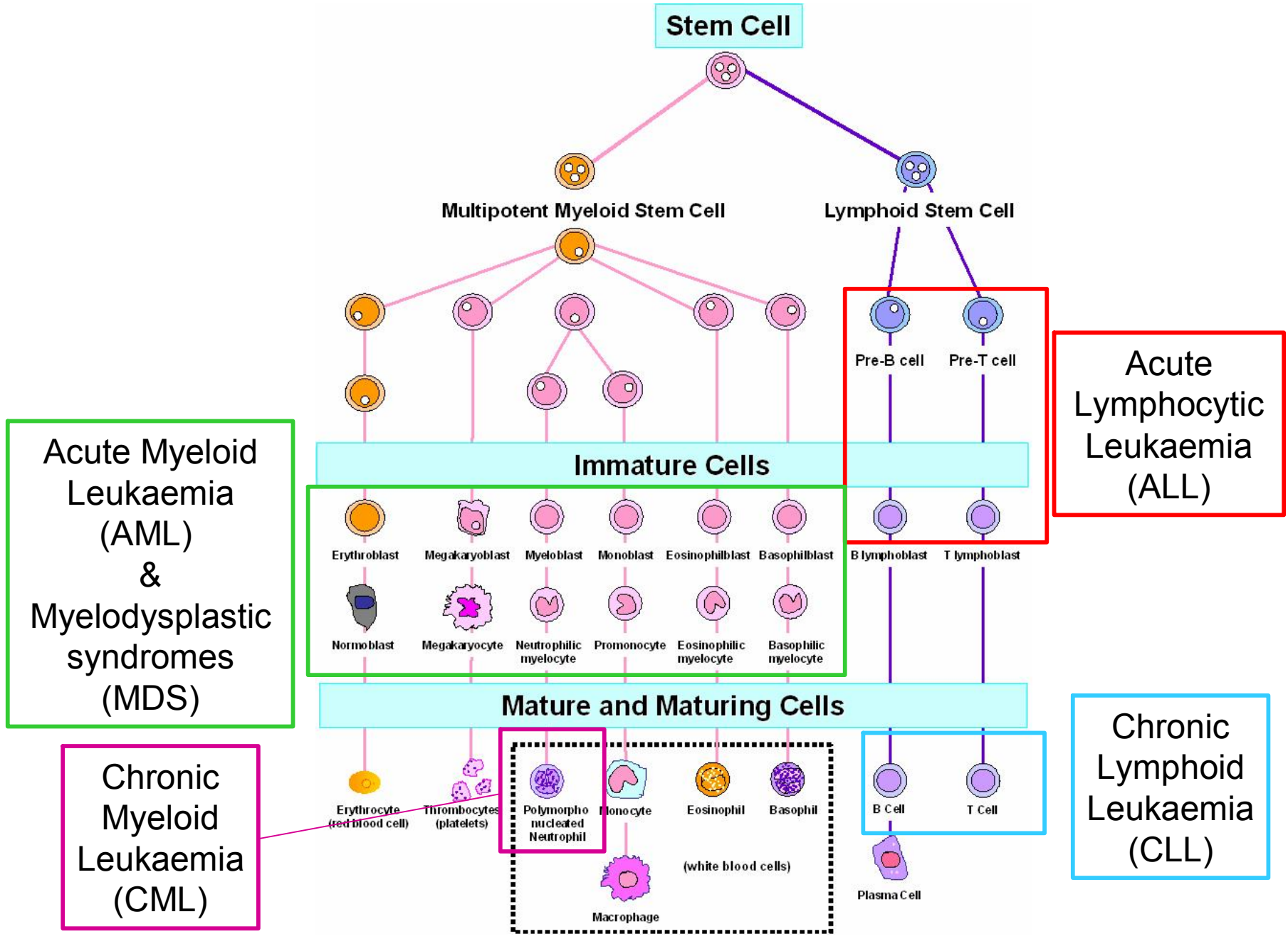


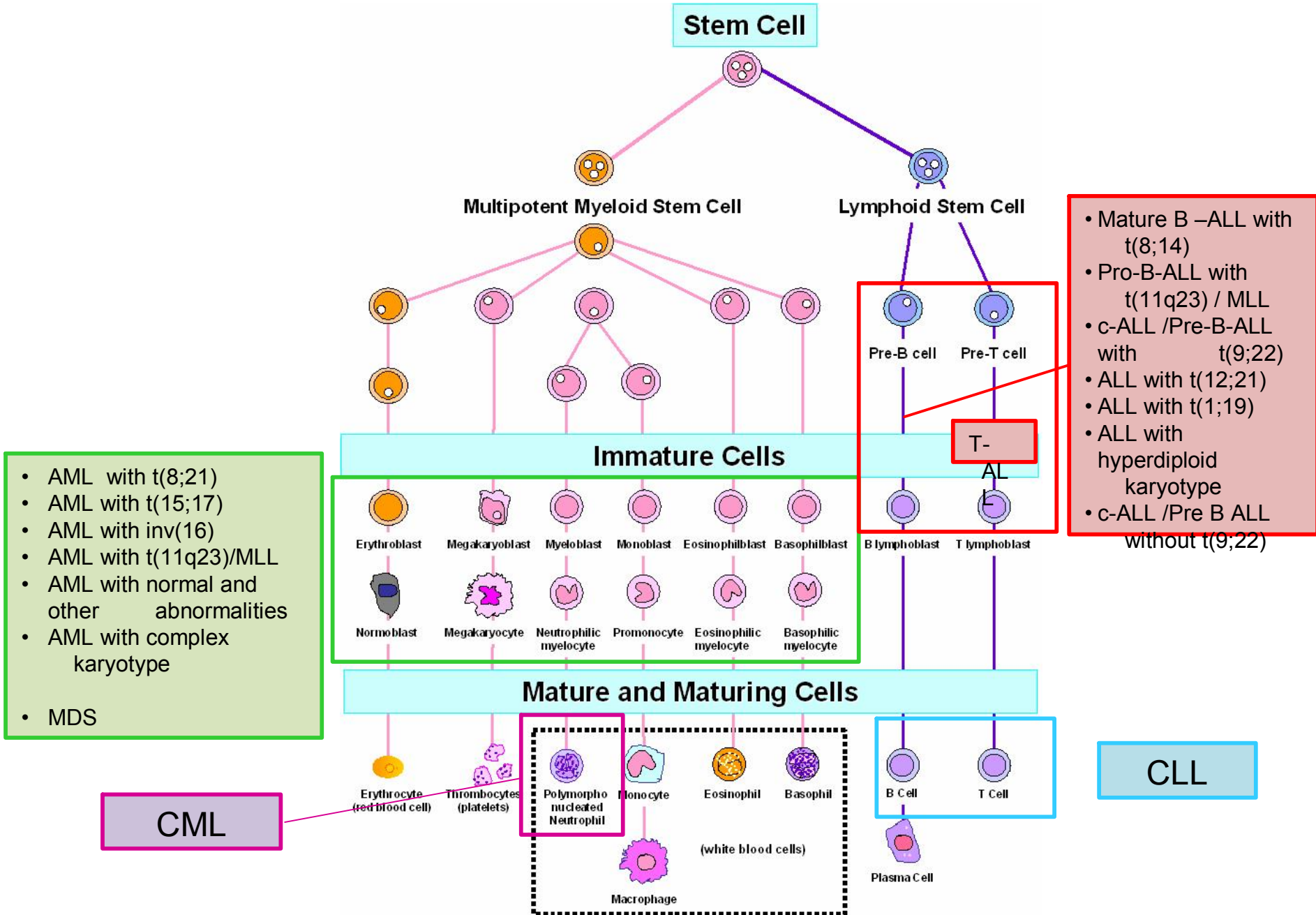
Outline

- MILE
 - Prephase
 - Stage I
 - Stage II
 - Implementation
- MDS
 - Diagnosis into Prognosis

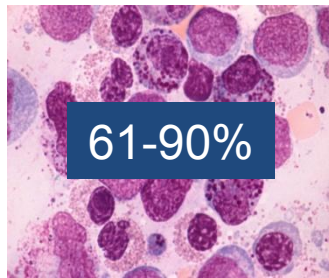
Leukaemia arises due to a failure of haematopoietic differentiation



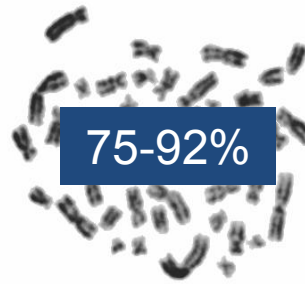




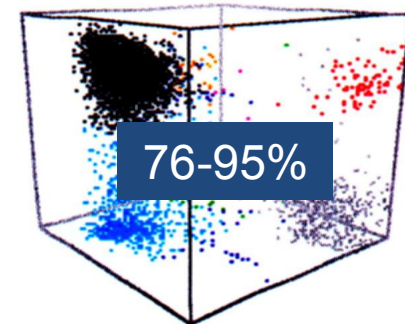
Leukaemia Diagnosis



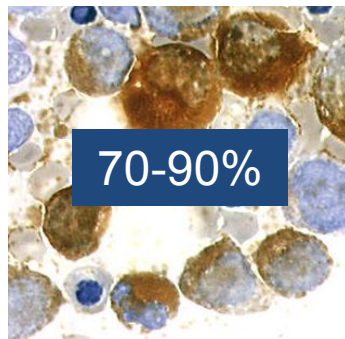
Morphology



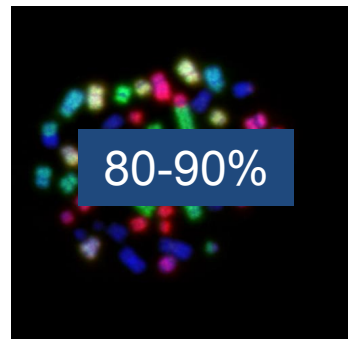
Cytogenetics



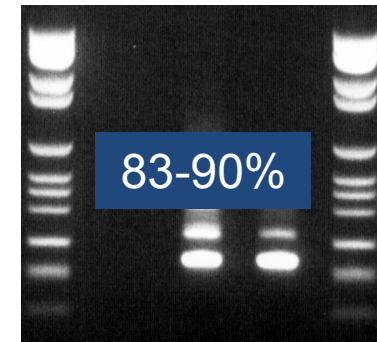
Immunophenotyping



Cytochemistry

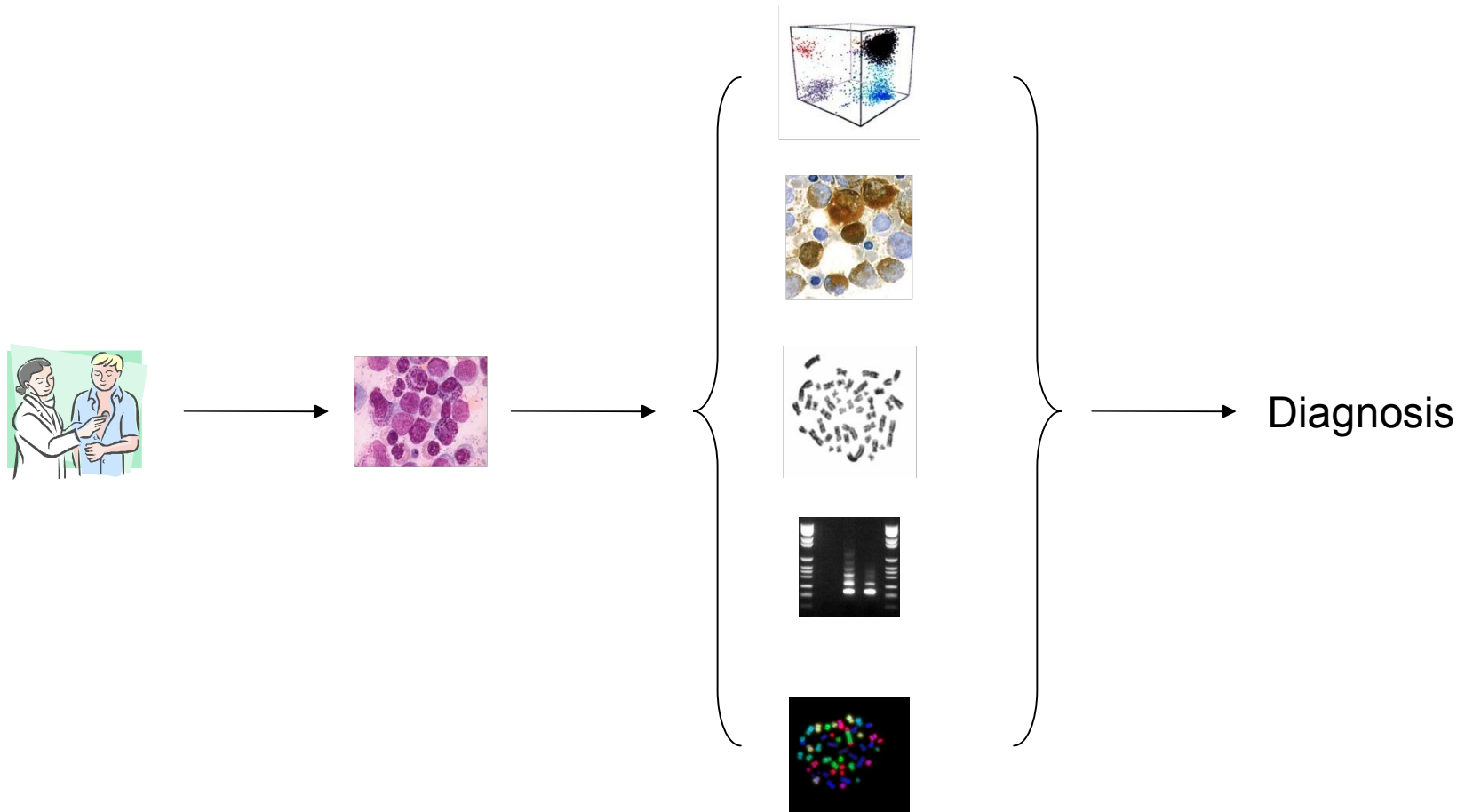


FISH

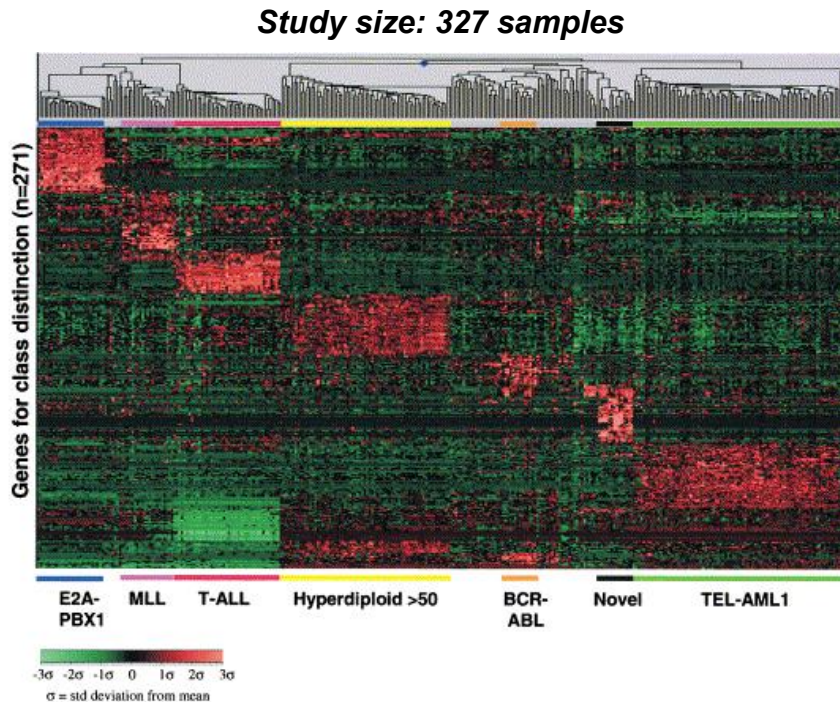


PCR

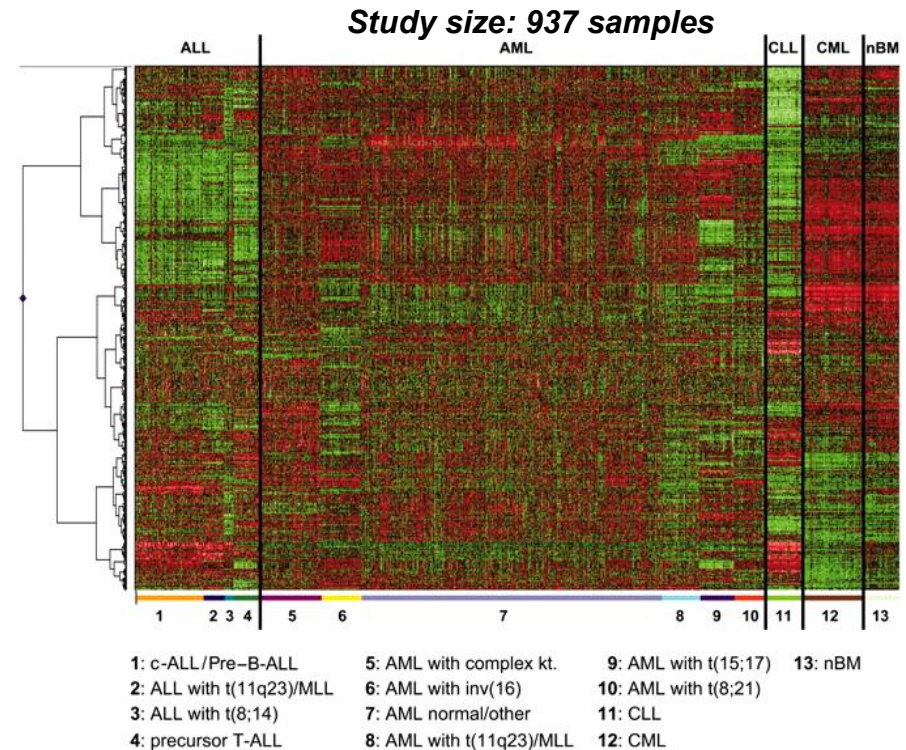
Leukaemia Diagnosis



Microarrays identify expression patterns for clinically and therapeutically relevant classes of paediatric and adult leukaemias



Yeoh et al., Cancer Cell 2002
[Clinically relevant pediatric ALL]



Haferlach et al., Blood 2005
[Clinically relevant adult leukaemias]

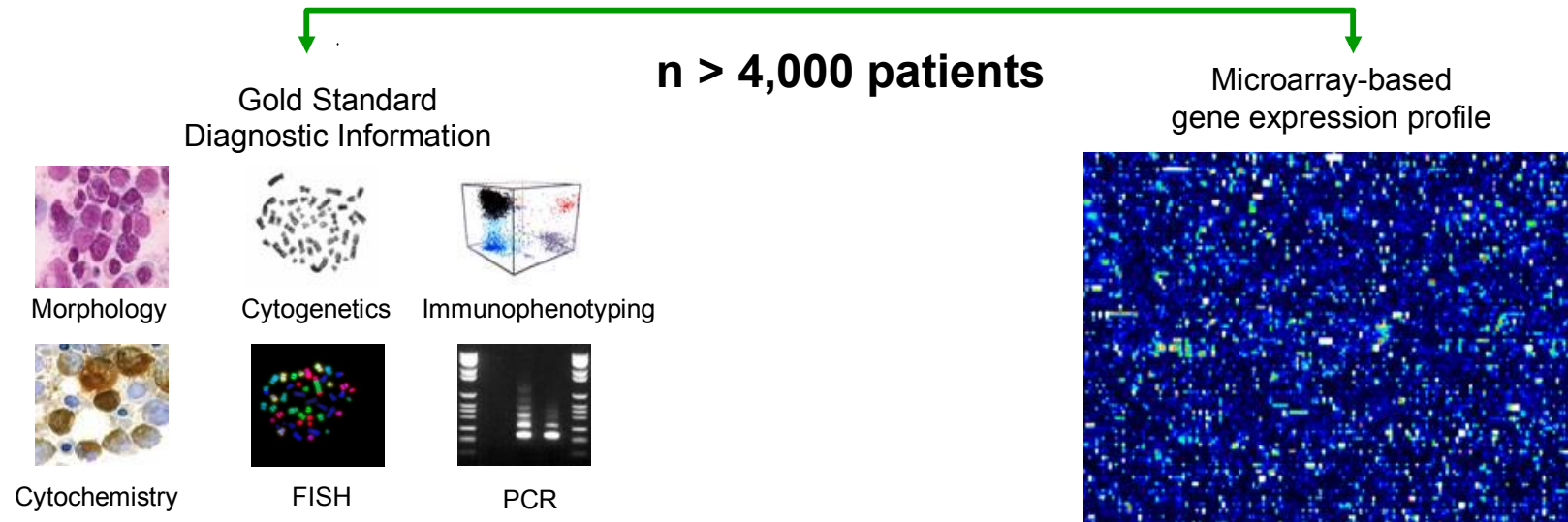


Gene expression profiling has become a powerful and robust technology. The application of microarrays for subclassification of leukaemia subtypes is possible as shown in mono-centric studies.

The MILE Study

Microarray Innovations in LEukemia

A sequential study to assess the clinical accuracy of the microarray test as compared to standard leukaemia laboratory methods (“Gold Standard”) for 16 classes of leukaemia, MDS and non-leukaemia / normal bone marrow



A global biomarker discovery and feasibility program includes 22 centers in 11 countries



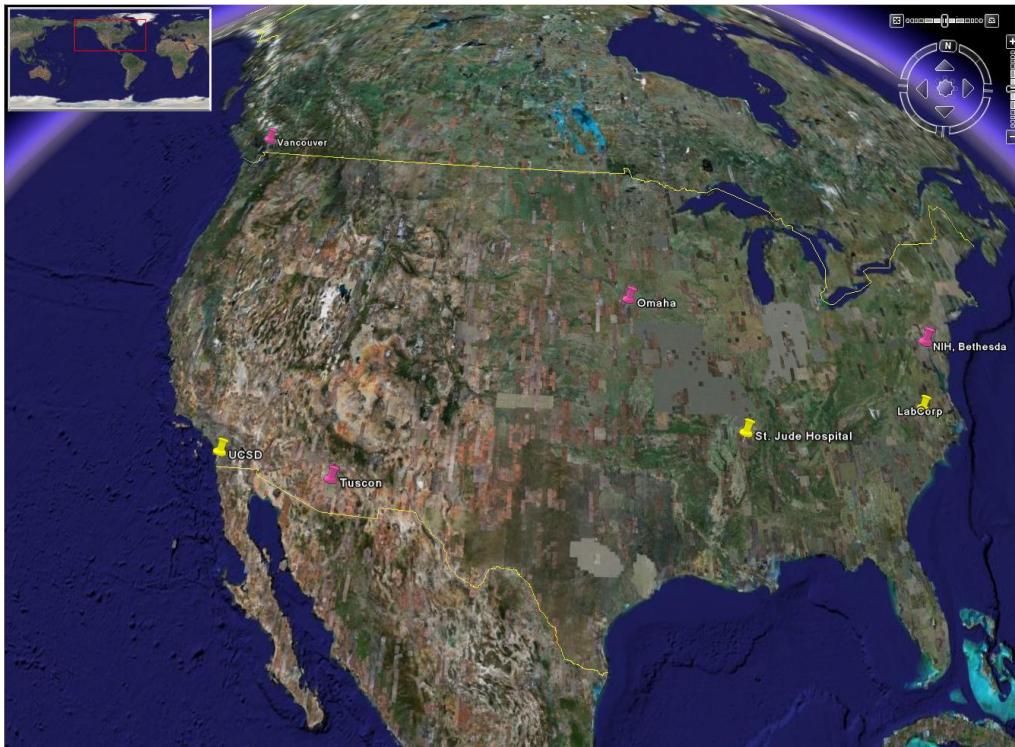
Leukaemia (MILE)

US: 3 centres, EU: 10 centres, Singapore



Lymphoma (NCI)

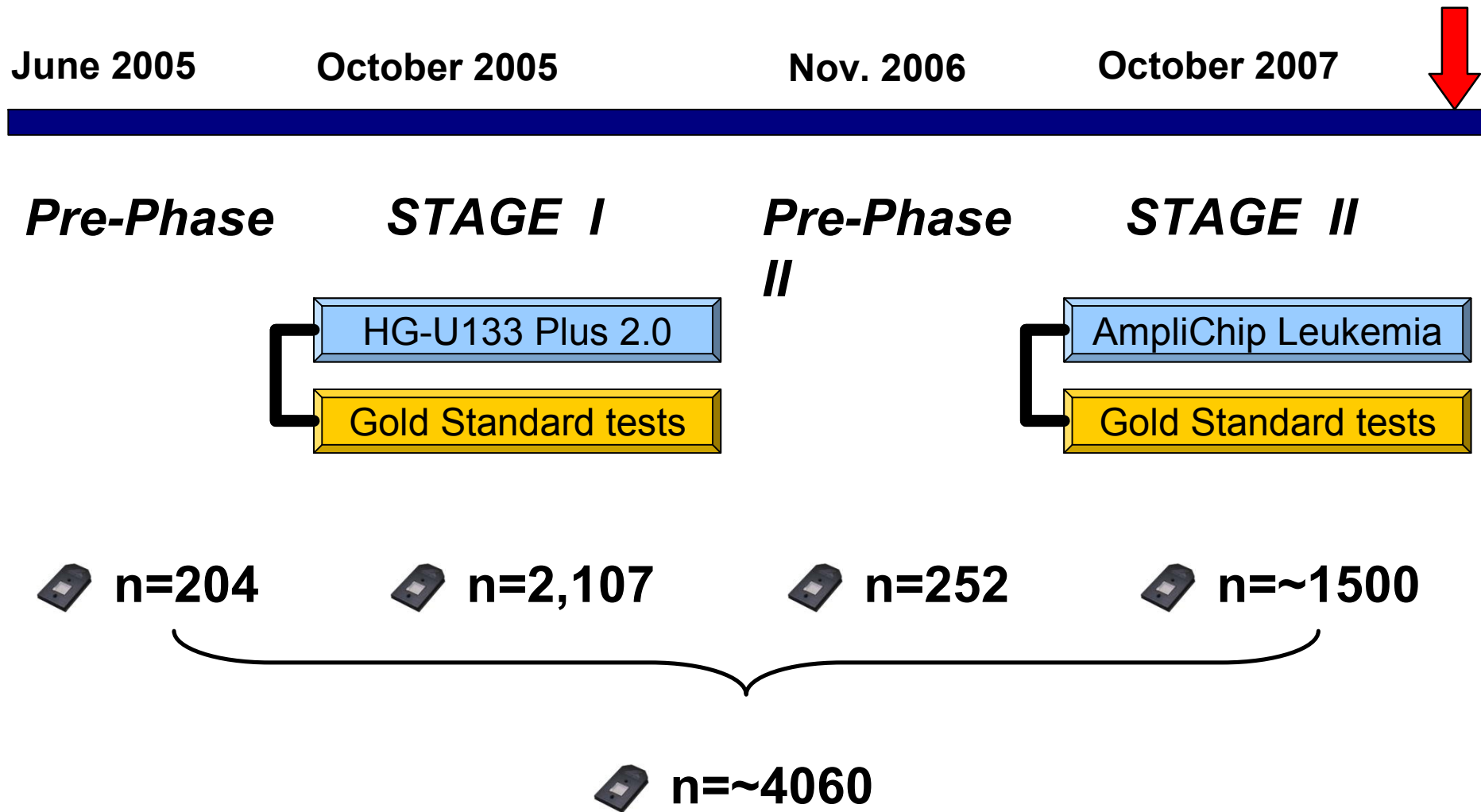
US: 3 centres, EU: 4 centres, Canada



MILE Stage I classification algorithm concept: 16 types of acute, chronic leukaemias, and MDS

No.	Class	Subclass				
1	mature B-ALL with t(8;14)					
2	Pro-B-ALL with t(11q23)/MLL					
3	c-ALL/Pre-B-ALL with t(9;22)					
4	T-ALL	<table border="1"> <tr><td><i>ALL, T-lineage, immature (Pre)</i></td></tr> <tr><td><i>ALL, T-lineage, immature (Pro)</i></td></tr> <tr><td><i>ALL, T-lineage, cortical</i></td></tr> <tr><td><i>ALL, T-lineage, mature</i></td></tr> </table>	<i>ALL, T-lineage, immature (Pre)</i>	<i>ALL, T-lineage, immature (Pro)</i>	<i>ALL, T-lineage, cortical</i>	<i>ALL, T-lineage, mature</i>
<i>ALL, T-lineage, immature (Pre)</i>						
<i>ALL, T-lineage, immature (Pro)</i>						
<i>ALL, T-lineage, cortical</i>						
<i>ALL, T-lineage, mature</i>						
5	ALL with t(12;21)					
6	ALL with t(1;19)					
7	ALL with hyperdiploid karyotype					
8	c-ALL/Pre-B-ALL without t(9;22)					
9	AML with t(8;21)					
10	AML with t(15;17)					
11	AML with inv(16)/t(16;16)					
12	AML with t(11q23)/MLL					
13	AML with normal karyotype + other abnormalities					
14	AML complex aberrant karyotype					
15	CLL	<table border="1"> <tr><td><i>mutated IgVH</i></td></tr> <tr><td><i>unmutated IgVH</i></td></tr> <tr><td><i>ZAP-70 positive</i></td></tr> <tr><td><i>ZAP-70 negative</i></td></tr> </table>	<i>mutated IgVH</i>	<i>unmutated IgVH</i>	<i>ZAP-70 positive</i>	<i>ZAP-70 negative</i>
<i>mutated IgVH</i>						
<i>unmutated IgVH</i>						
<i>ZAP-70 positive</i>						
<i>ZAP-70 negative</i>						
16	CML					
17	MDS					
18	Non-leukemia and healthy bone marrow					

MILE Study: Timelines



Pre-phase

Goals of Pre-Phase –

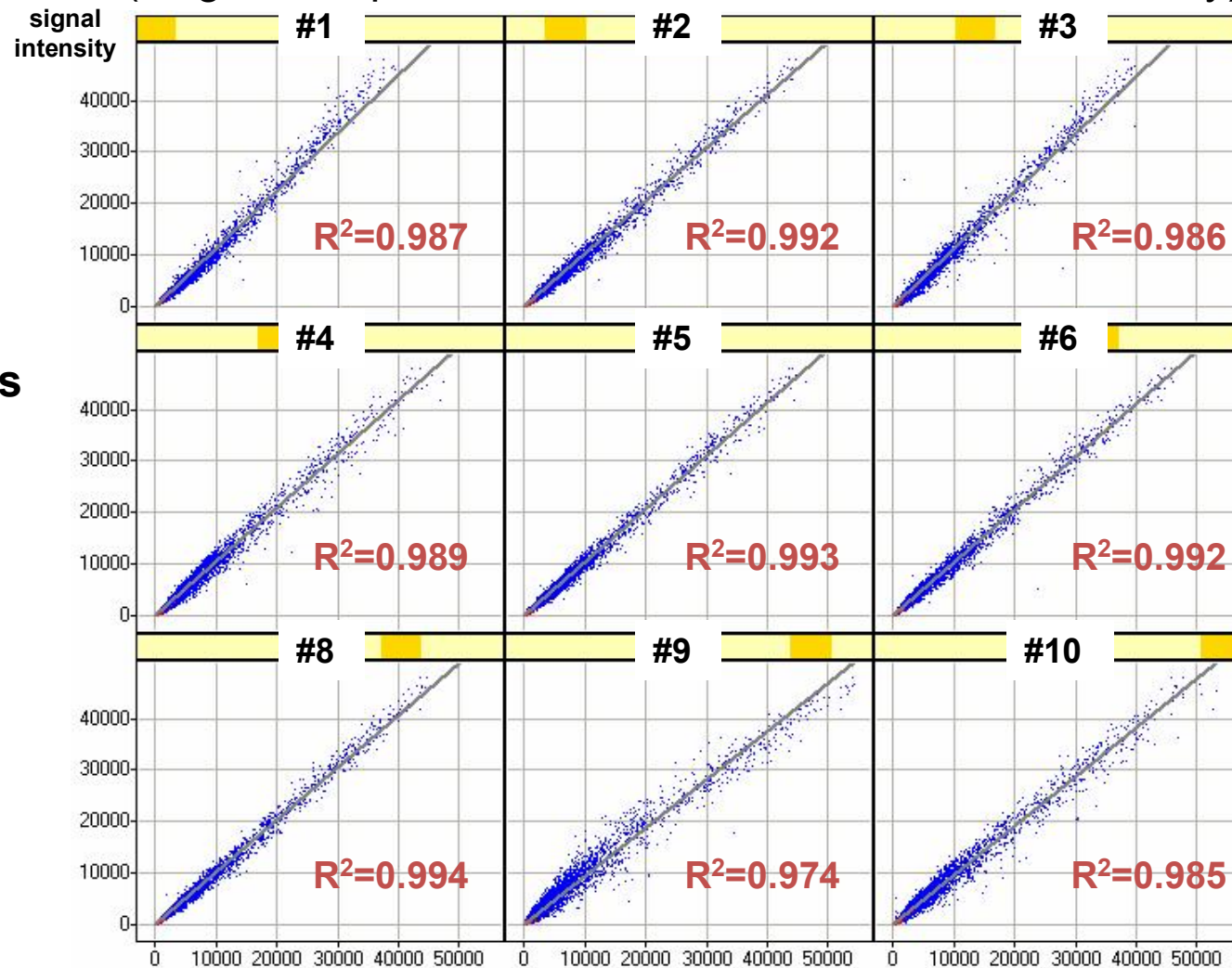
Intra-laboratory reproducibility and inter-laboratory comparability

Each **Operator** is trained on an identical sample preparation protocol

Each **Laboratory** is provided the same laboratory equipment, as well as kits & reagents from the same source for sample preparation and microarray analysis

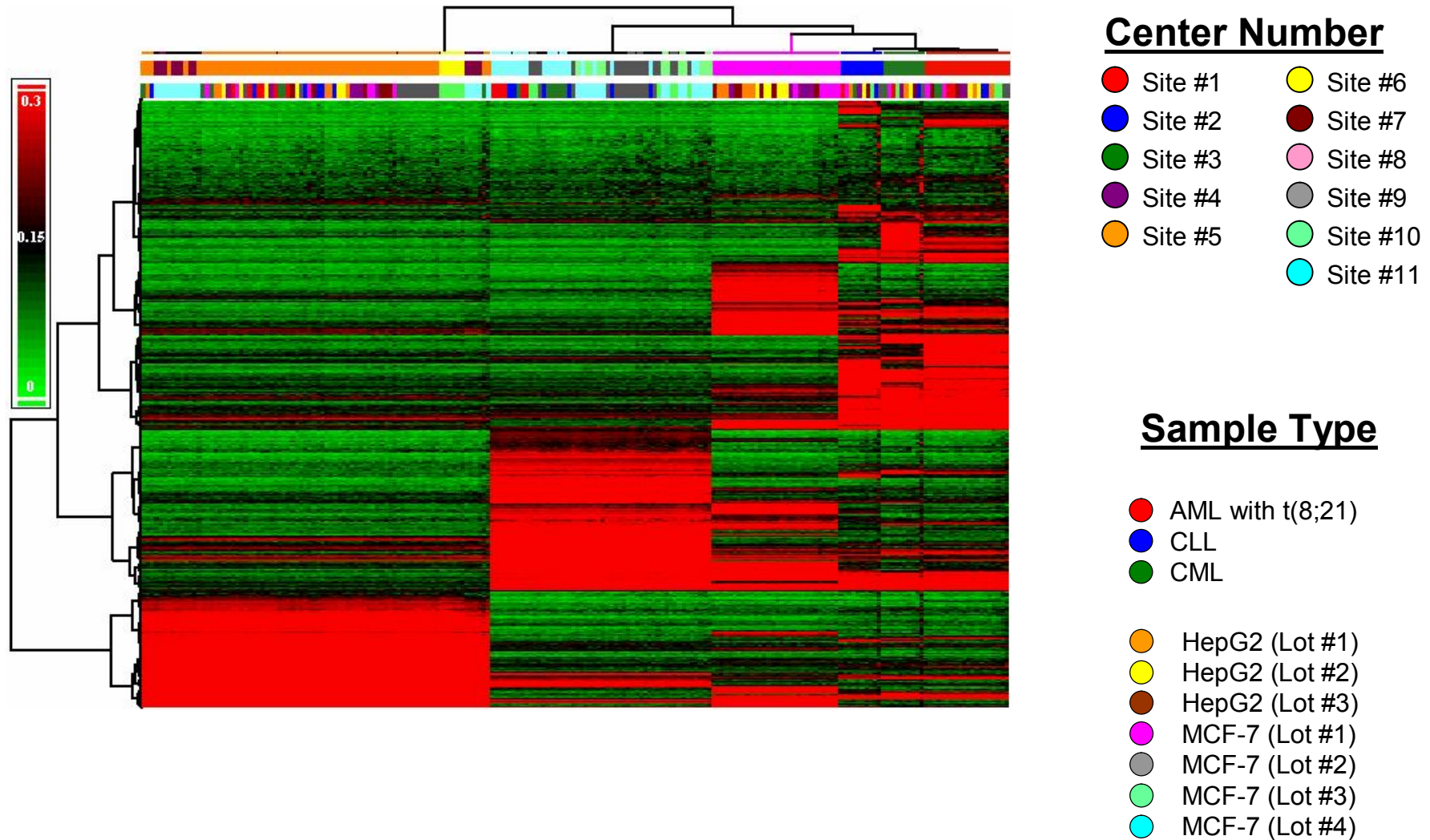
Pre-phase for inter-laboratory reproducibility

5 μ g HEPG2 sample from proficiency testing
(all genes represented on HG-U133 Plus 2.0 microarray)

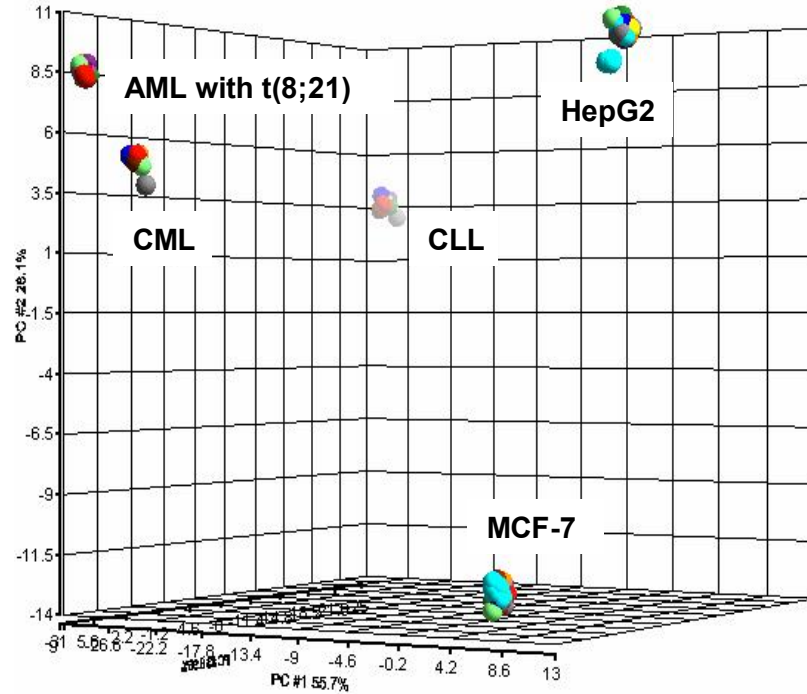
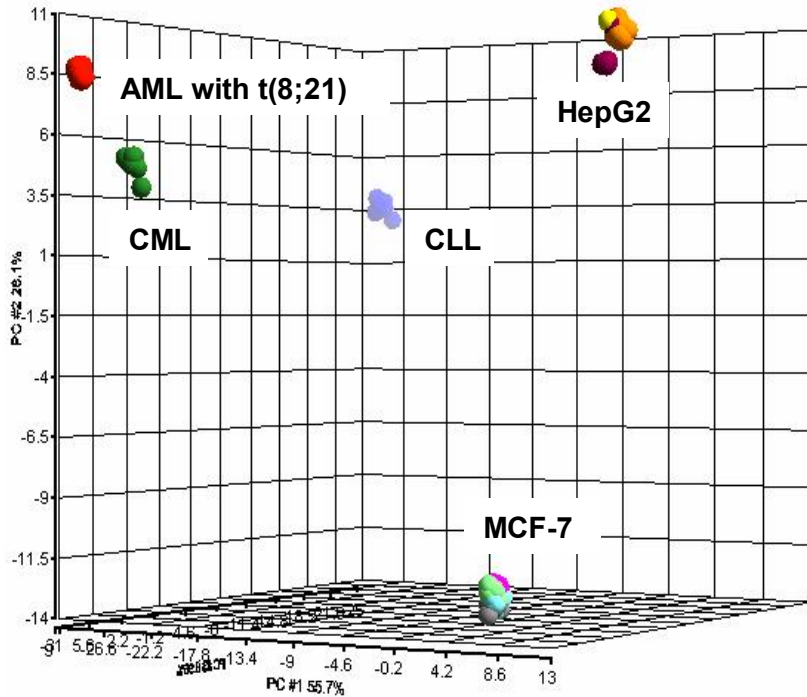


Example:
Center 7 vs.
9 other centers

Pre-phase for inter-laboratory reproducibility



Pre-phase for inter-laboratory reproducibility



Sample Type

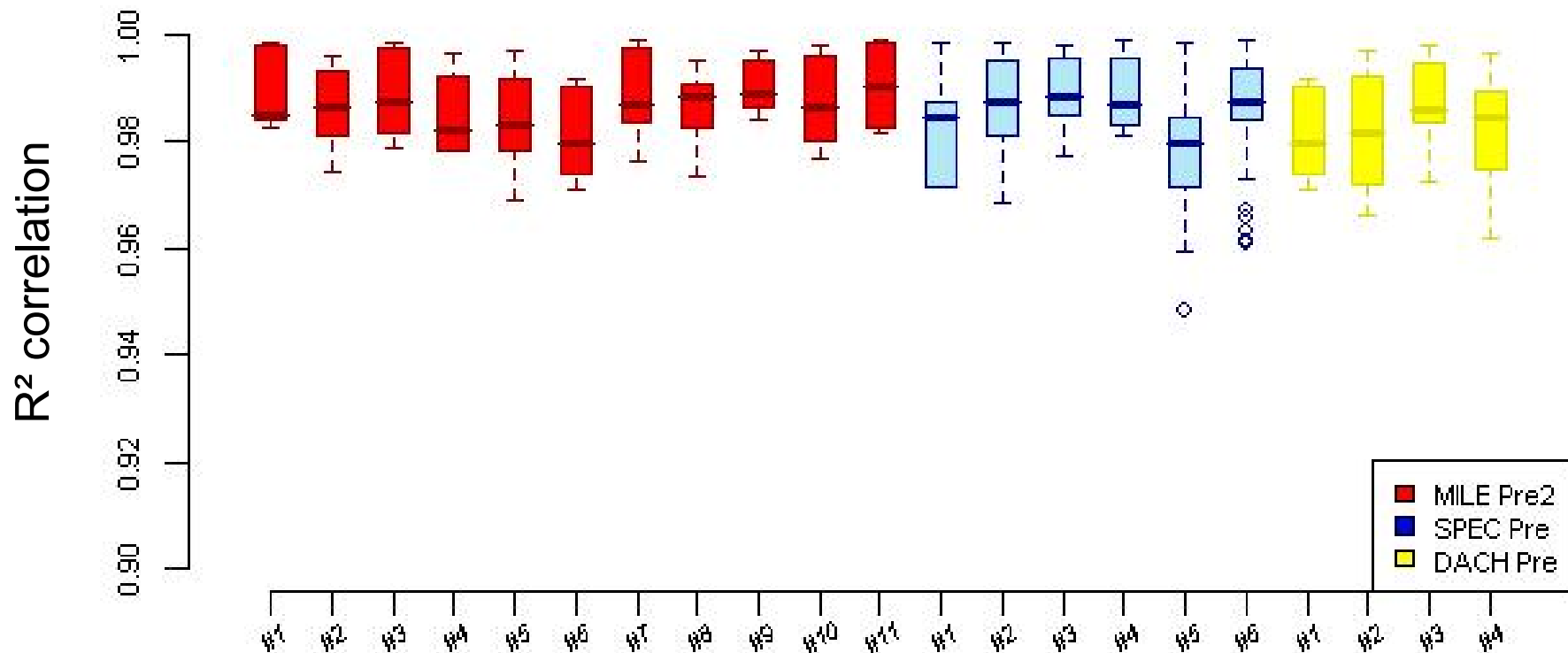
- | | |
|--------------------|------------------|
| ● AML with t(8;21) | ● HepG2 (Lot #1) |
| ● CLL | ● HepG2 (Lot #2) |
| ● CML | ● HepG2 (Lot #3) |
| | ● MCF-7 (Lot #1) |
| | ● MCF-7 (Lot #2) |
| | ● MCF-7 (Lot #3) |
| | ● MCF-7 (Lot #4) |

Center Number

- | | |
|-----------|------------|
| ● Site #1 | ● Site #6 |
| ● Site #2 | ● Site #7 |
| ● Site #3 | ● Site #8 |
| ● Site #4 | ● Site #9 |
| ● Site #5 | ● Site #10 |
| | ● Site #11 |

Leukaemia and Lymphoma Proficiency Data

Intra-site correlation analysis for 21 laboratories



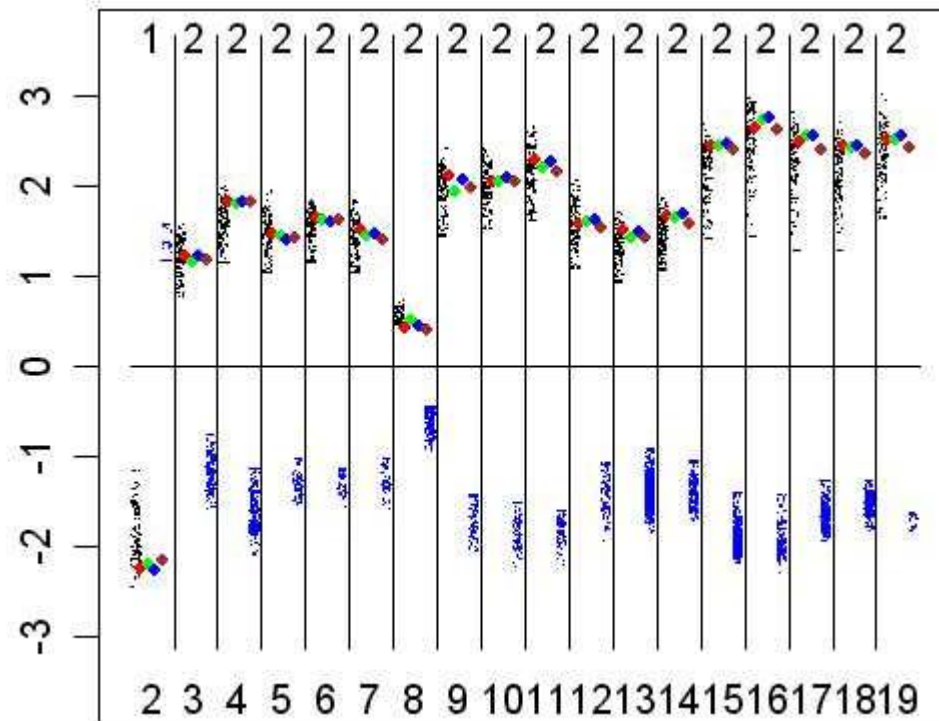
Data is shown for the HepG2 cell line (all genes represented on the Plus 2.0 array)

Intra-laboratory comparison of gene expression values from HepG2 replicates

Leukaemia Classifier – Version 7 (LCv7)

- Model 7 is an all-pair-wise linear classification model for 18 classes (16 leukaemia classes, MDS, and none-of-the-targets)
- Consists of $18 * (18 - 1) / 2 = 153$ classifiers
- Uses 534 probe sets on HG-U133 Plus 2.0 microarray

- Training Data: class 2
- Training Data: other classes



Test Data

- ◆ MUC_00715: Class 2
- ◆ BAS_00174: Class 2
- ◆ LIN_00142: Class 2
- ◆ GEN_00142: Class 2

ID: 07-008434 (c2)

Stage I

Data Set (2,916 whole genome profiles)

Class		Sensitivity	Specificity	
1	mature B-ALL with t(8;14)	0.85	1.00	
2	Pro-B-ALL with t(11q23)/MLL	1.00	1.00	
3	c-ALL/Pre-B-ALL with t(9;22)	0.90	1.00	
4	T-ALL	<p>Accuracy by cross-validation:</p> <h2>93.9%</h2> <ul style="list-style-type: none"> • based on 30-fold CV • all HG-U133 Plus 2.0 	1.00	
5	ALL with t(12;21)		1.00	
6	ALL with t(1;19)		1.00	
7	ALL with hyperdiploidy		1.00	
8	c-ALL/Pre-B-ALL with t(9;22)		0.99	
9	AML with t(8;21)		1.00	
10	AML with t(15;17)		1.00	
11	AML with inv(16)/t(16;16)		1.00	
12	AML with t(11q23)/MLL		0.92	1.00
13	AML with normal karyotype + other abnormalities		0.93	0.99
14	AML complex aberrant karyotype		0.90	1.00
15	CLL	1.00	1.00	
16	CML	0.97	1.00	
17	MDS	0.88	0.98	
18	<i>None of the above</i>	0.99	0.99	

Stage I

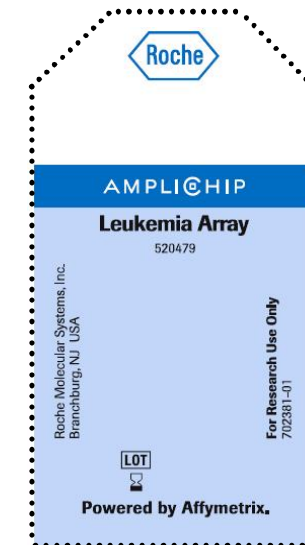
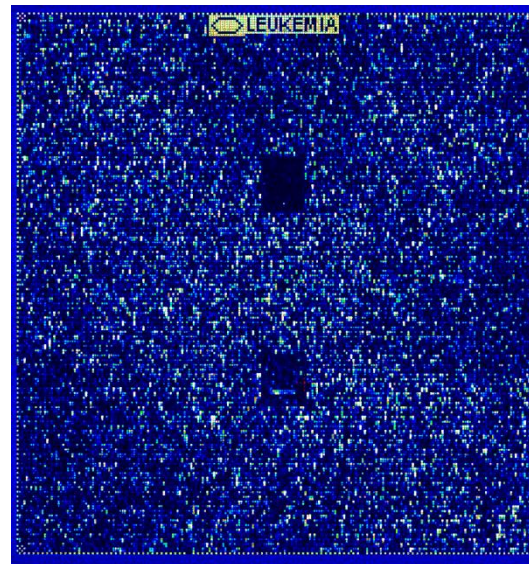
Data Set (2,662 whole genome profiles excluding MDS)

Class		Sensitivity	Specificity	
1	mature B-ALL with t(8;14)	0.85	1.00	
2	Pro-B-ALL with t(11q23)/MLL	1.00	1.00	
3	c-ALL/Pre-B-ALL with t(9;22)	0.90	1.00	
4	T-ALL	<p>Accuracy by cross-validation:</p> <p>95.3%</p> <ul style="list-style-type: none"> • based on 30-fold CV • all HG-U133 Plus 2.0 	1.00	
5	ALL with t(12;21)		1.00	
6	ALL with t(1;19)		1.00	
7	ALL with hyperdiploidy		1.00	
8	c-ALL/Pre-B-ALL with t(9;22)		0.99	
9	AML with t(8;21)		1.00	
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11	AML with inv(16)/t(16;16)		1.00	
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15	CLL	1.00	1.00	
16	CML	0.97	1.00	
18	<i>None of the above</i>	0.99	0.99	

Stage II

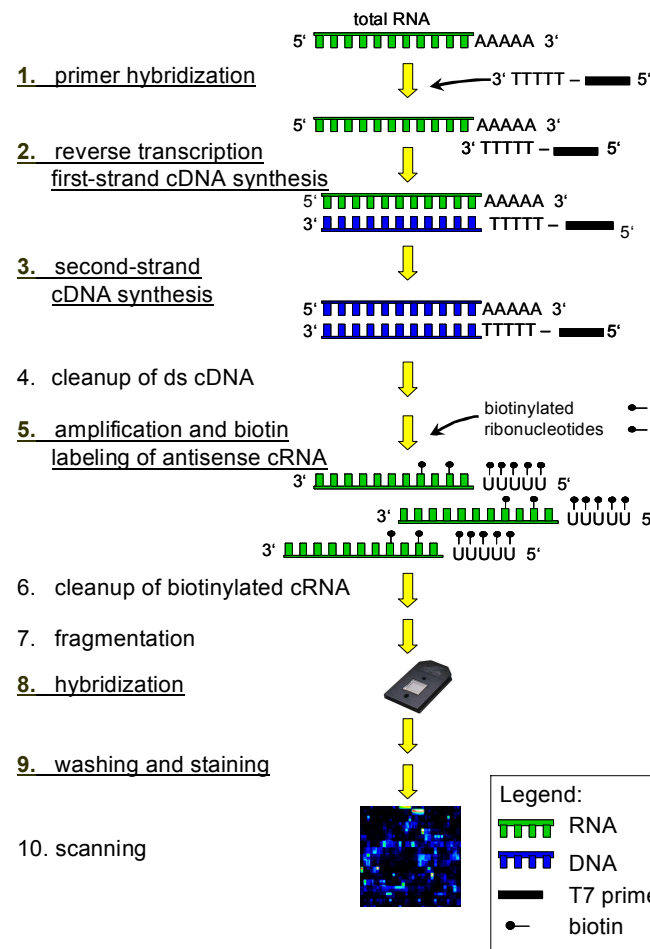
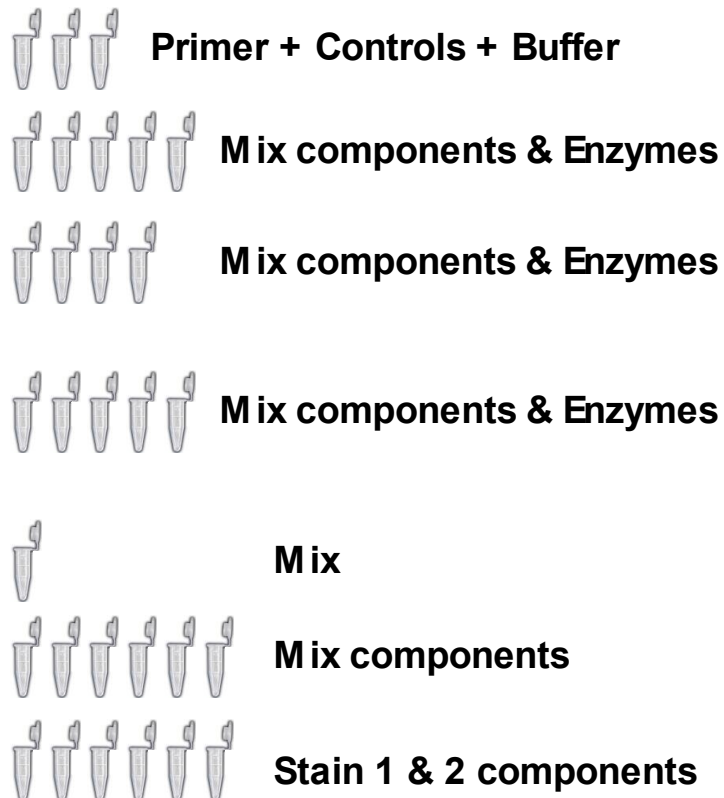
- Class 19 introduced
 - Normal PB
 - Class 18: normal BM)
- Prospective samples obtained
- Amplichip used for analysis
 - ~1100 probe sets
 - 539 probe sets for classification
 - CD markers
 - Normalisation and control probe sets
 - Repeated 4 time across chip
- Centre-blinded array result
- QC checked
 - Lower QC failure rate than Stage I
- Concordance determined
- Discordant results checked
 - Array call confirmed
 - Array call not confirmed

Customized design: AmpliChip Leukaemia

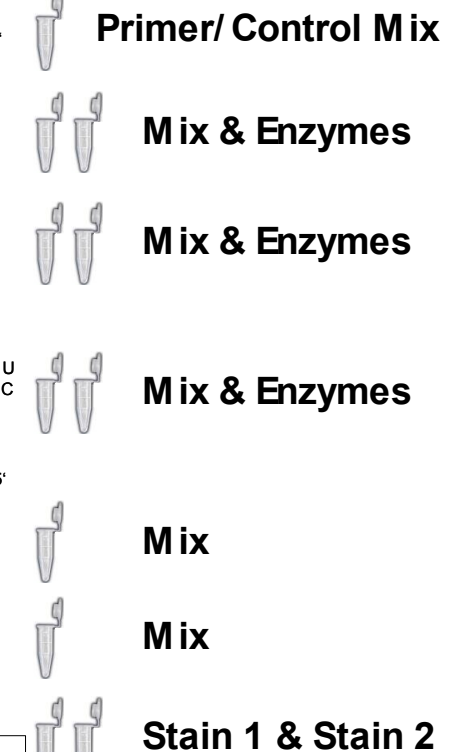


Standardized workflow for routine clinical laboratory usage (< 2.5 h hands-on time)

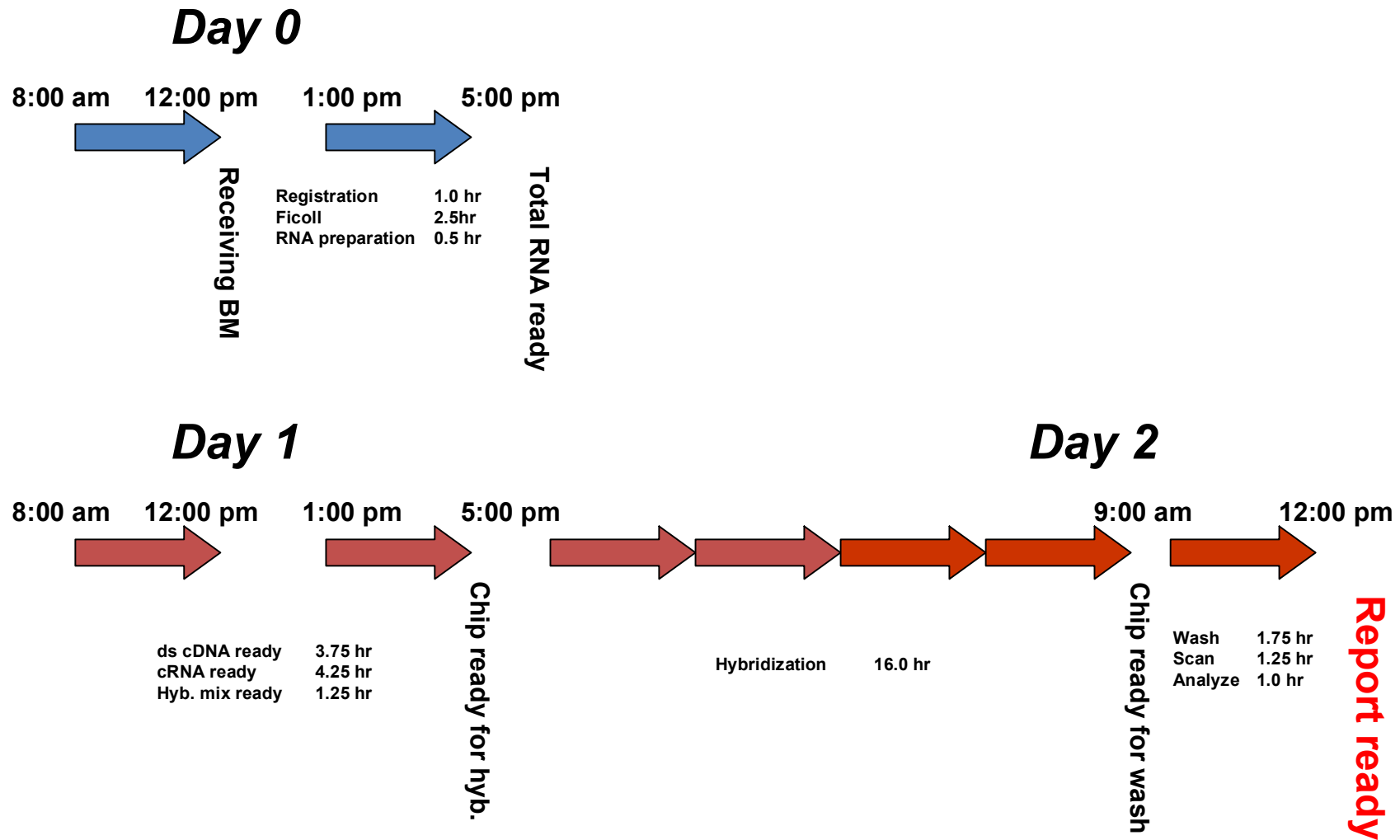
previous kit configuration



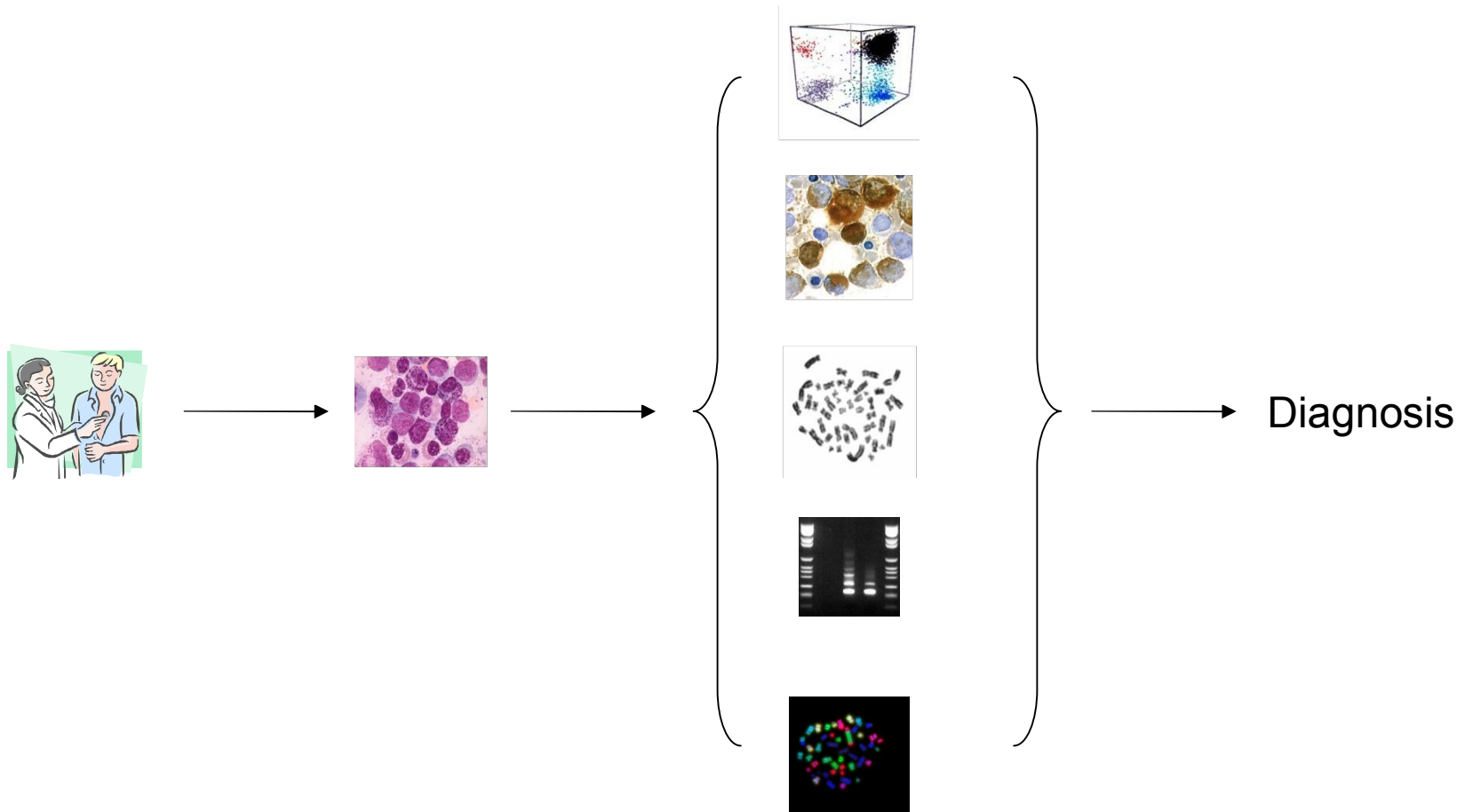
reduced complexity with final kit



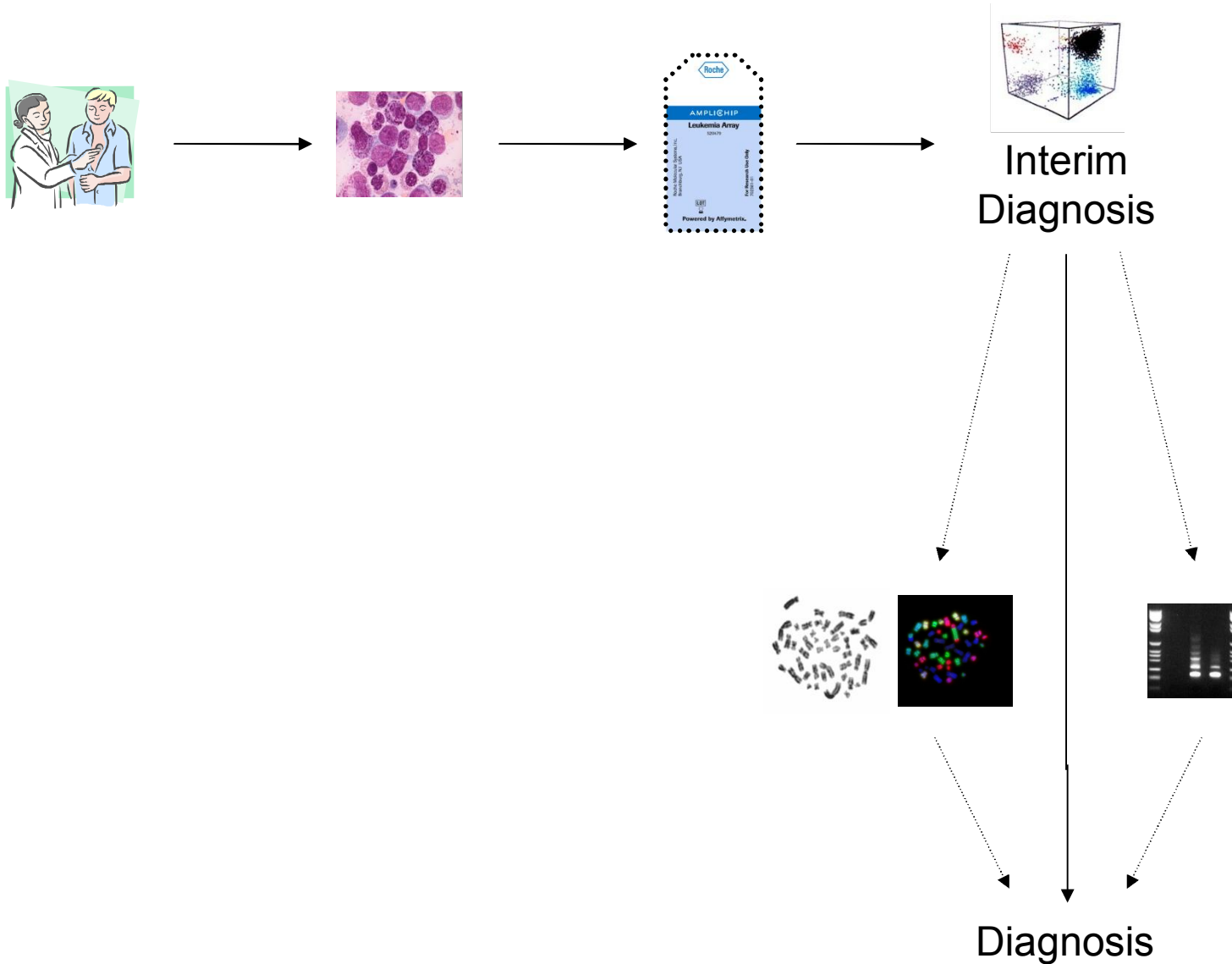
Laboratory workflow concept for prospective specimen collection: Results in < 48 hours



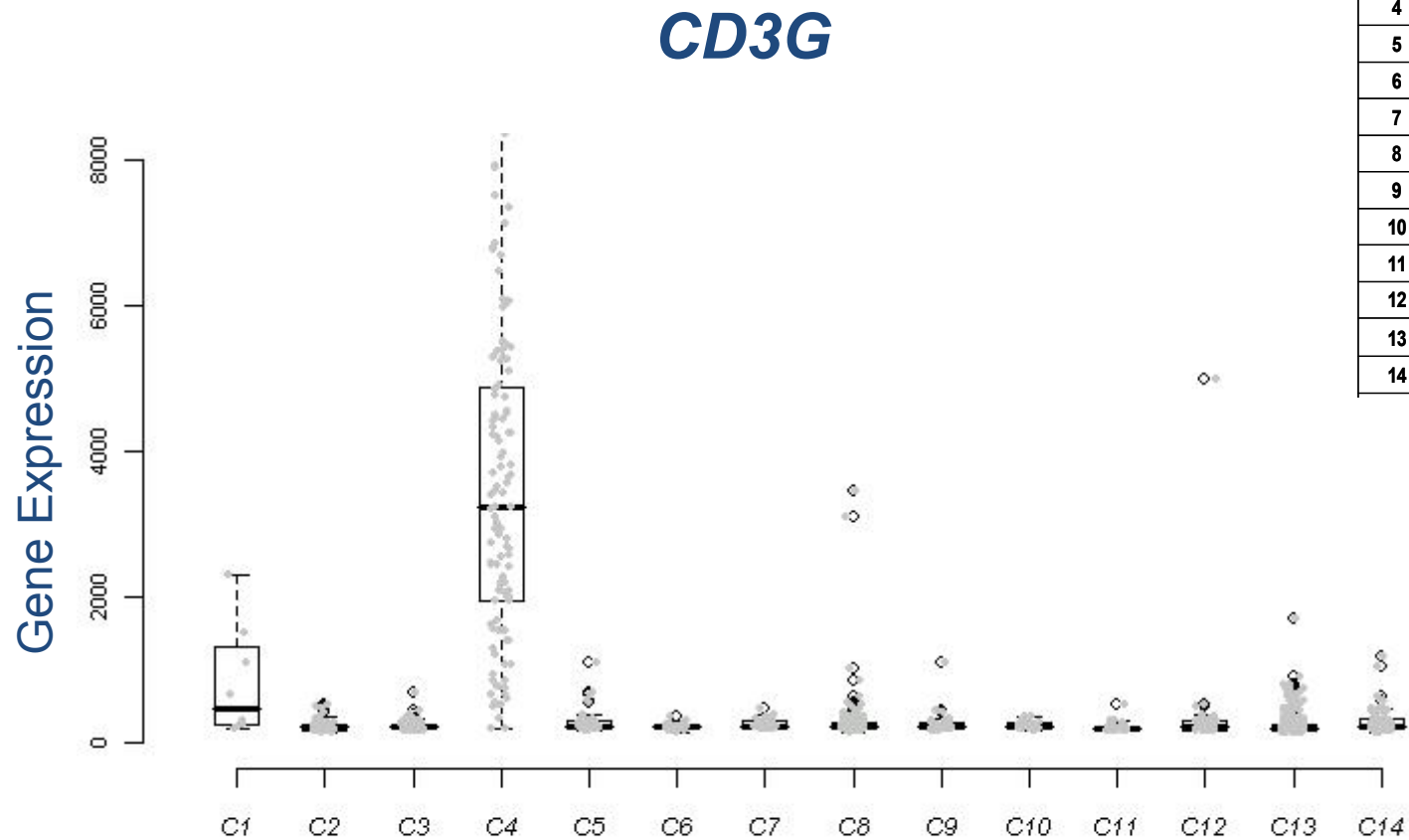
Leukaemia Diagnosis



Leukaemia Diagnosis



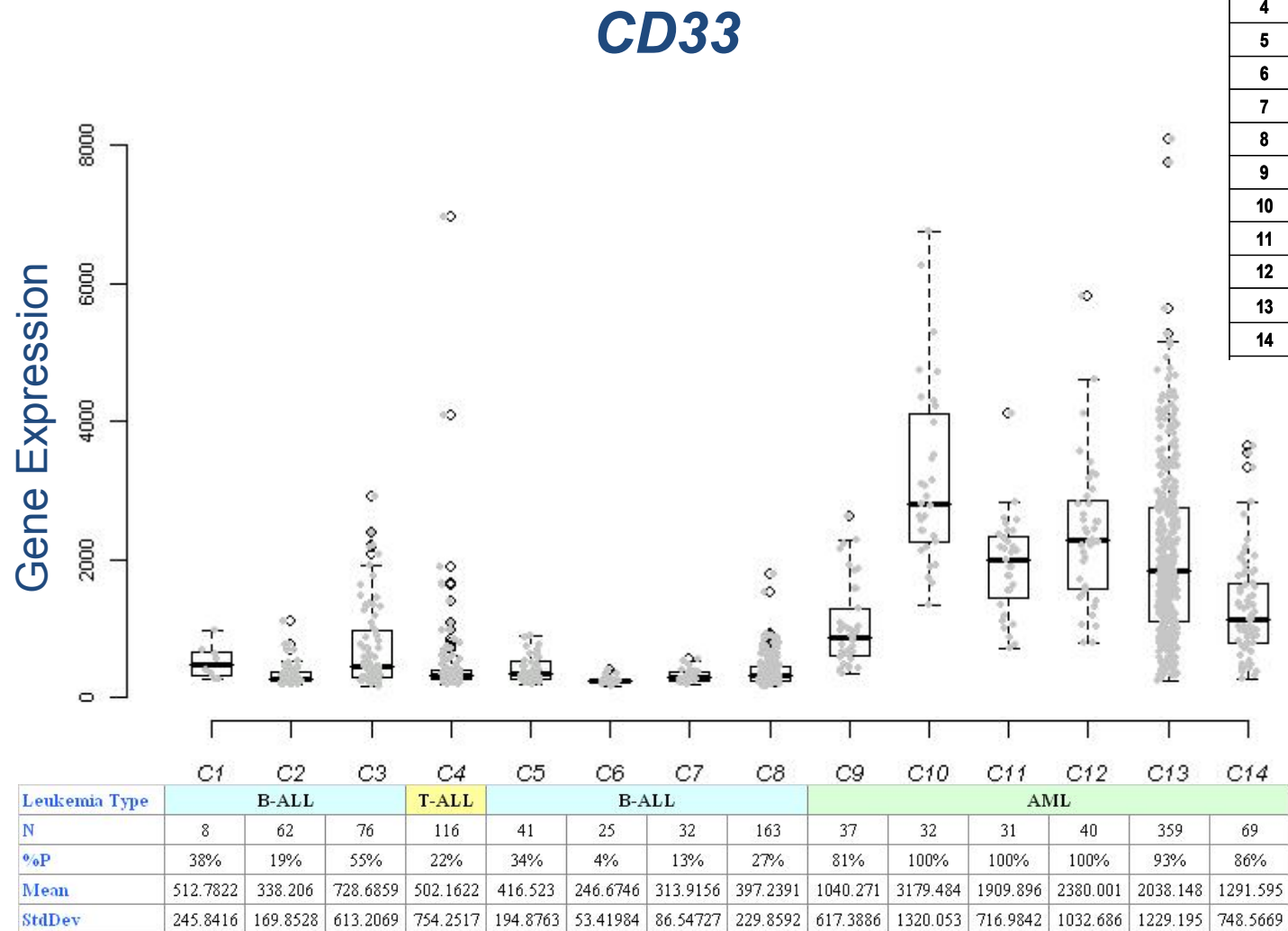
“Virtual Immunophenotyping”



1	mature B-ALL with t(8;14)
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3	c-ALL/Pre-B-ALL with t(9;22)
4	T-ALL
5	ALL with t(12;21)
6	ALL with t(1;19)
7	ALL with hyperdiploid karyotype
8	c-ALL/Pre-B-ALL without t(9;22)
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10	AML with t(15;17)
11	AML with inv(16)/t(16;16)
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13	AML with normal karyotype + other abnormalities
14	AML complex aberrant karyotype

Leukemia Type	B-ALL			T-ALL	B-ALL				AML					
N	8	62	76	116	41	25	32	163	37	32	31	40	359	69
%P	63%	18%	24%	97%	54%	4%	28%	30%	22%	16%	16%	20%	22%	39%
Mean	824.8148	237.0229	235.0884	3447.732	307.3067	224.1451	259.2421	303.223	270.3386	244.0073	224.28	367.2348	255.1604	296.3412
StdDev	768.1744	77.79264	72.51045	2063.485	177.5311	42.27795	69.52565	351.0806	155.9649	59.49919	70.50167	753.2512	145.1394	179.2288

“Virtual Immunophenotyping”



1	mature B-ALL with t(8;14)
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3	c-ALL/Pre-B-ALL with t(9;22)
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13	AML with normal karyotype + other abnormalities
14	AML complex aberrant karyotype

MDS and AML - Relationship

Are AML and MDS part of a continuous spectrum of disease and not specific disorders?

MDS is a pre-leukaemic disorder in which the neoplastic clone has been established, but which may or may not fully progress to AML

MDS

DNA damage

Accelerated apoptosis

Ineffective
haematopoiesis



AML

Further genetic evolution

Defective apoptosis

Leukaemic
transformation

MILE study Stage I

MDS:

- 174 MDS samples analysed
 - Berlin
 - Cardiff
 - Munich
 - Salamanca
- 49.1% correctly called **MDS**
 - (Class 17)
- 24.6% called “AML”
 - 20.0% AML with normal or others (Class 13)
 - 4.6% AML with complex cytogenetics (Class 14)
 - **“MDS with an AML like signature”**
- 24.0% called Non-leukaemia / Normal bone marrow
 - (Class 18)
 - **“MDS with a non-leukaemia like signature”**

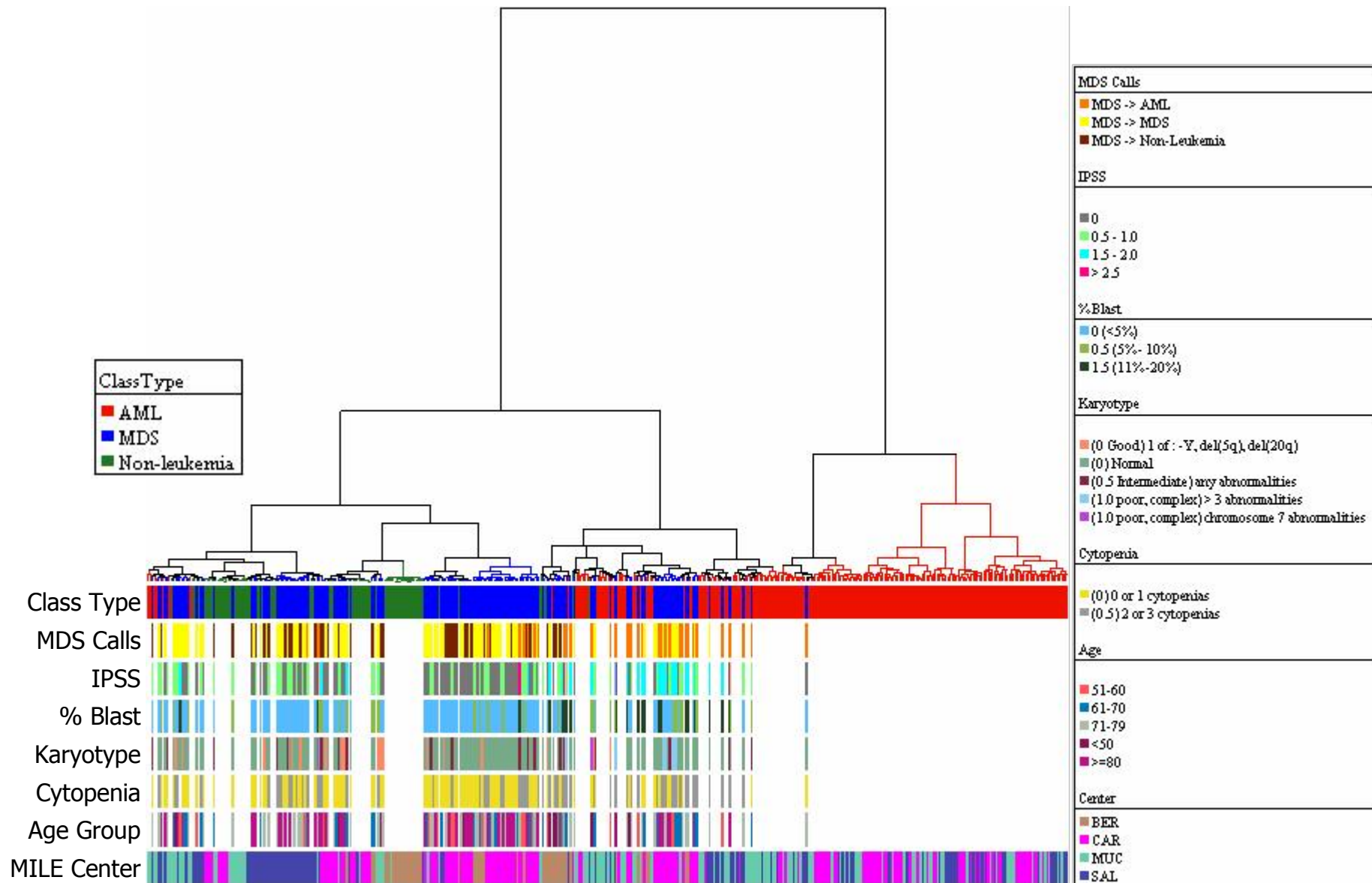
External Morphology Review

- Morphology slides for submitted MDS samples reviewed by:
 - Prof Loeffler; Germany
 - Dr Bowen; UK
- Review of slides included:
 - FAB
 - WHO
 - Blast count
 - Cytopenia

164 confirmed as MDS and assigned a WHO classification

10 discordant samples removed from MDS analysis

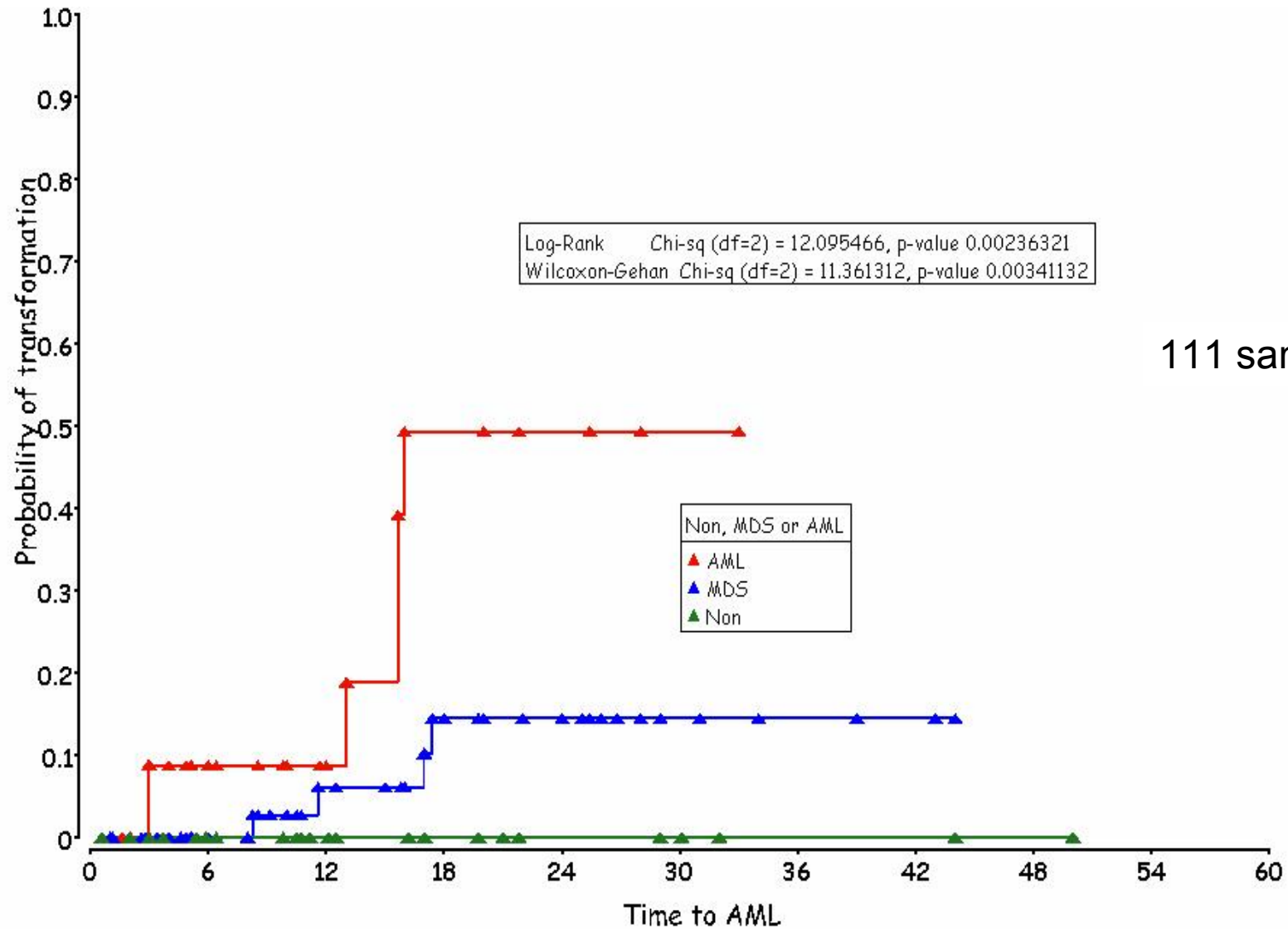
- 6 Reclassified as AML
- 4 Excluded from MDS & MILE study
 - aCML, CLL, myeloma and incomplete diagnosis



436 samples: 203 **AML**, 164 **MDS**, 69 **Non-leukaemia**

ICv7: 534 probe sets: Euclidean & Ward's

Transformation to AML



MILE Study

- High accuracy of disease classification over 16 classes of leukaemia
- High degree of inter- and intra-laboratory correlation
- Rapid result - <48 hrs
- Cost comparable to existing diagnostic tests
 - Refined workflow
- Adaptable to include new validated disease classifications
- Possible prognostic implications

Acknowledgements



- Germany
 - Munich: Torsten Haferlach, Claudia Haferlach
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- Spain
 - Salamanca: Jesus Hernandez
- France:
 - Nancy: Marie Christine Bene
 - Paris: Elizabeth McIntyre
 - Montpellier: Tom de Vos
- UK:
 - Cardiff: Alan Burnett



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NUH Singapore

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Koay



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