

# Targeting by probe-based circularisation

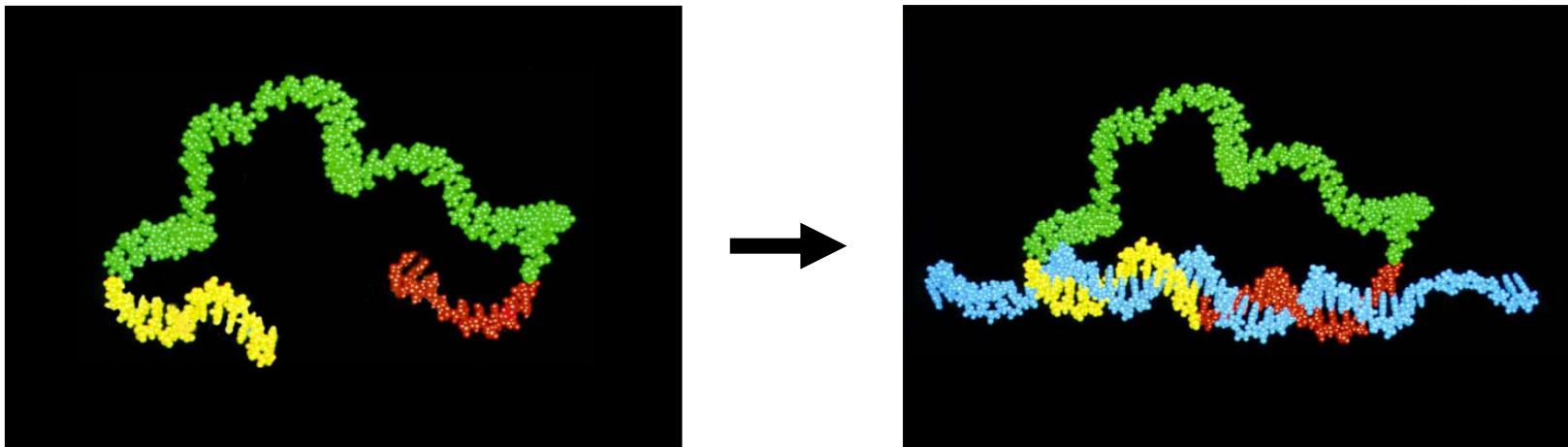
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Uppsala University

# Outline

- Padlock probes for multiplex targeted genotyping
- Selector probes for multiplex targeted genome analysis
  - sequencing (clinical sequencing)
  - CNV detection
- Amplified single-molecule detection (digital RCA)
- In situ genotyping

# Padlock probes

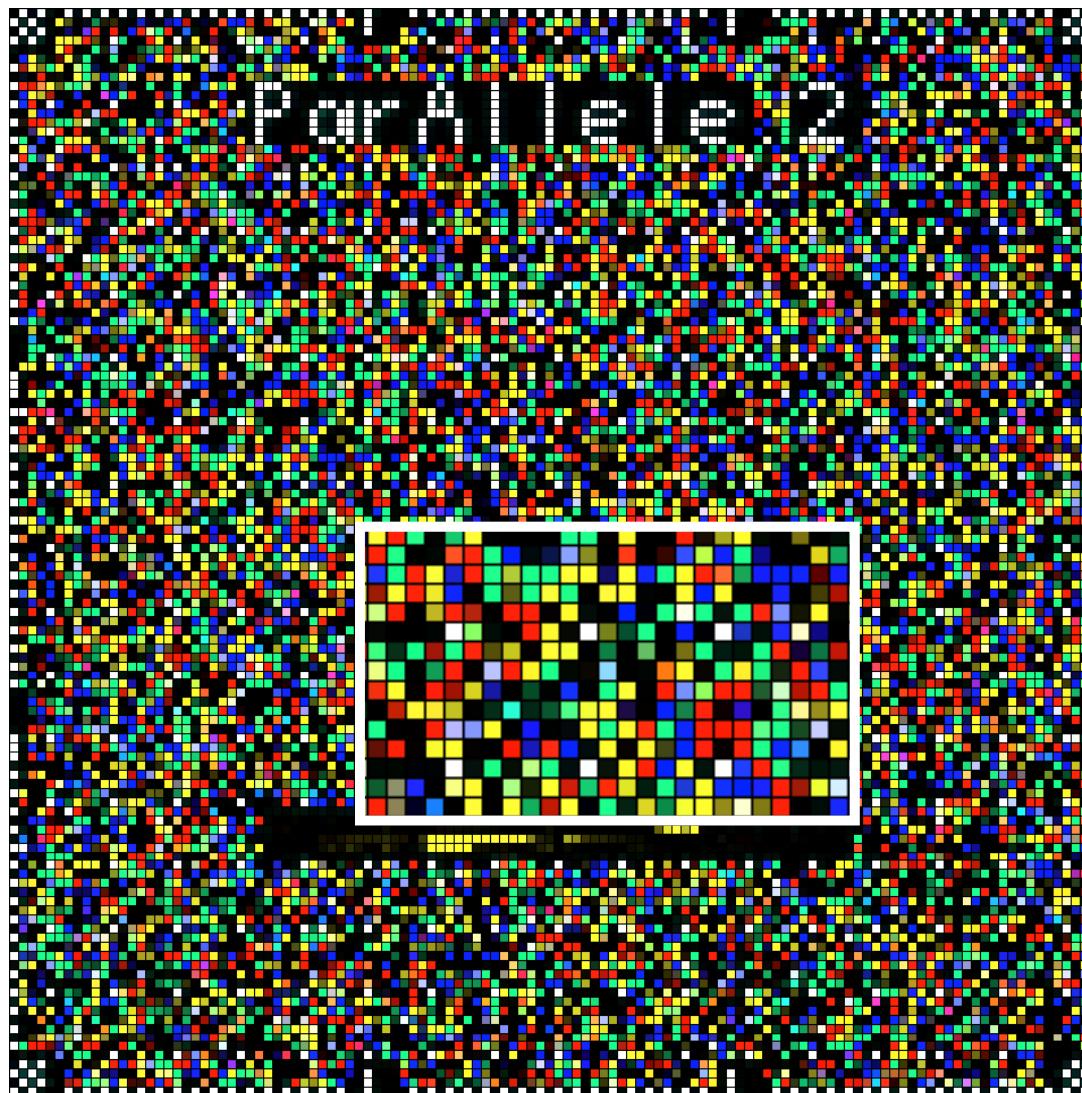


- Dual-recognition provides specificity
- Robust allele-distinction due to enzymatic ligation using standard reaction conditions
- Probes become topologically linked
- Unimolecular

Nilsson, et al. (1994) *Science*, **265**, 2085-2088.

Nilsson, et al. (1997) *Nature Genetics*, **16**, 252-255.

Nilsson, et al. (2000) *Nature Biotechnol*, **18**, 791-793.

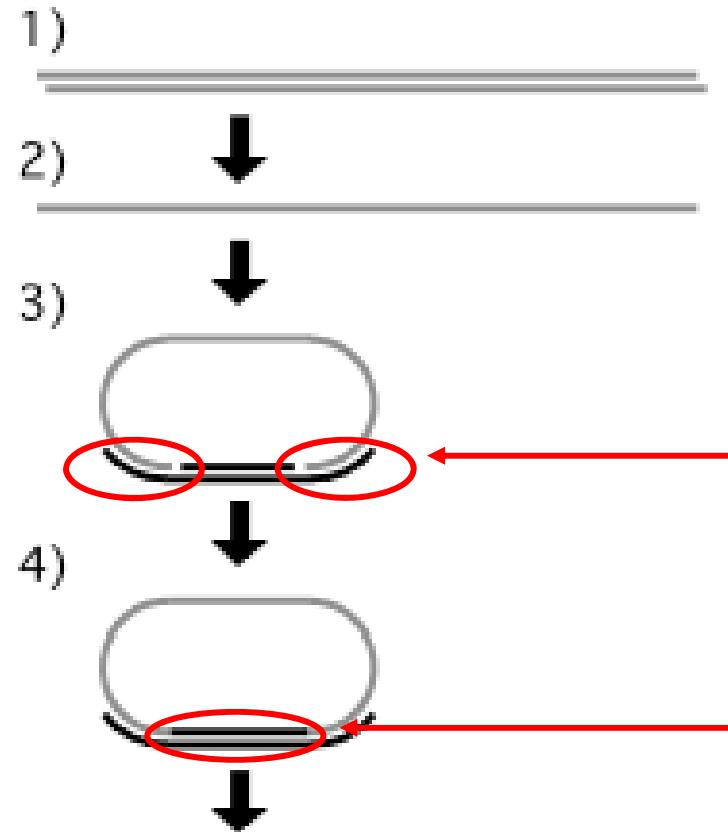
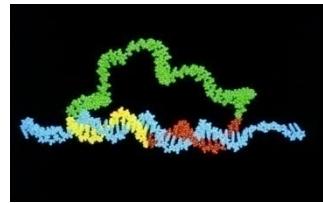


20,000-plex  
Genotyping

Affymetrix targeted  
genotyping

Hardenbol *et al*,  
*Nature Biotechnol* 2003;  
Hardenbol *et al*,  
*Genome Res* 2005

# Targeted genome analysis using selector probes

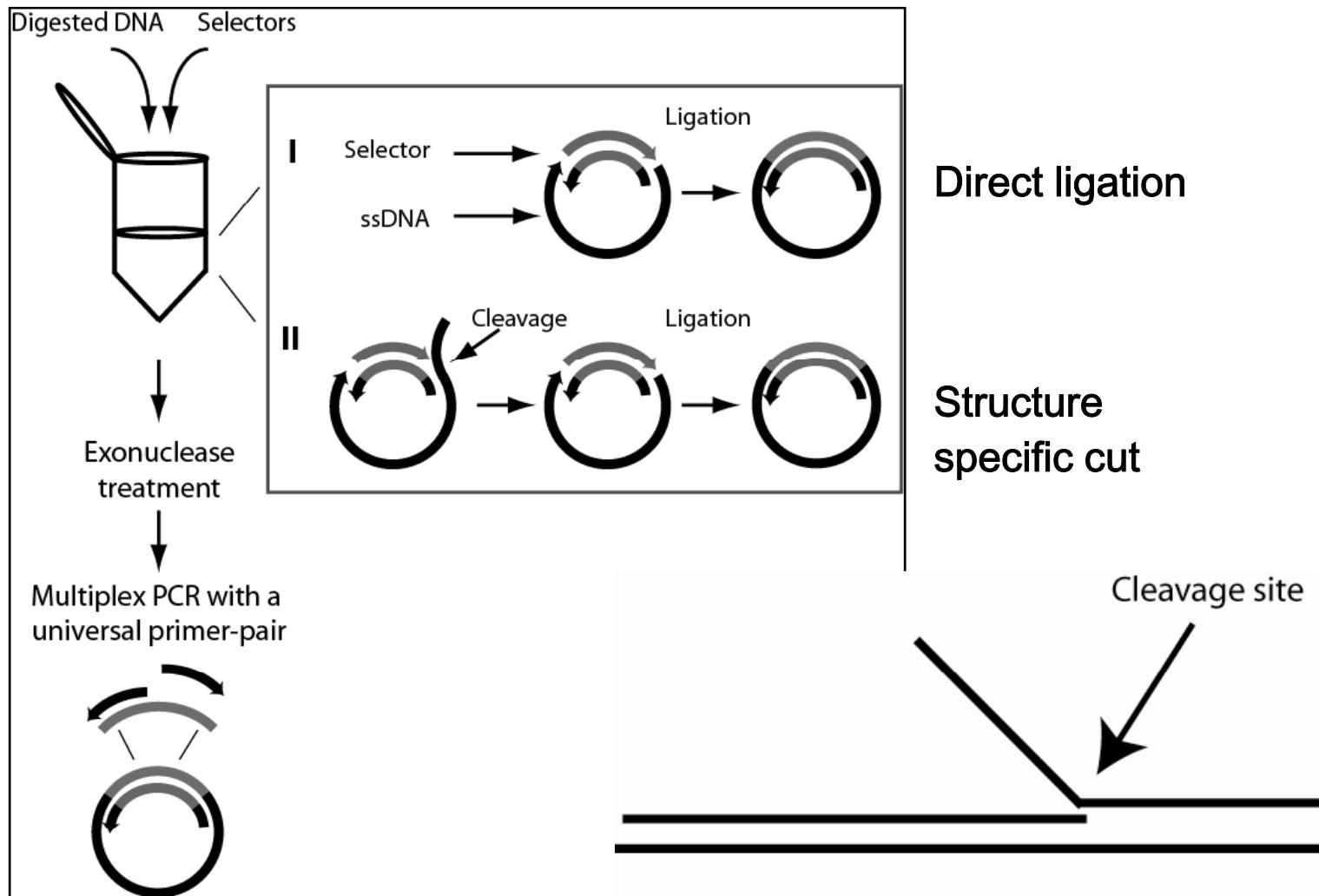


Sequence-specific  
circularization

Universal sequence  
inserted

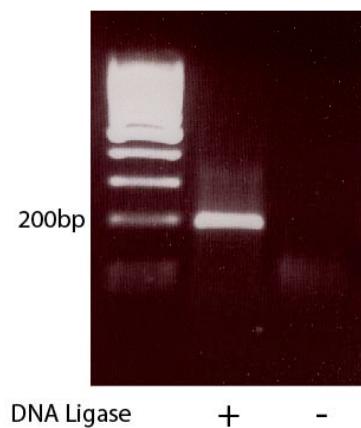
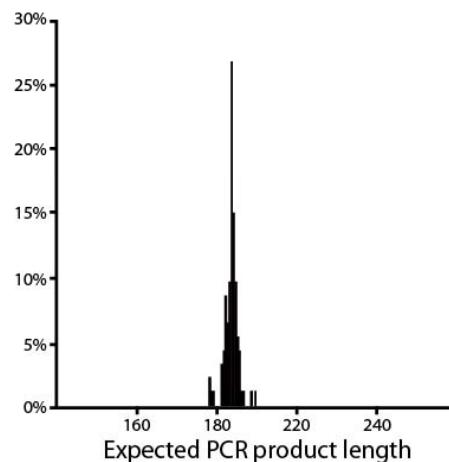
Circle selection and amplification

# The Selection Process



Lyamichev et.al (1993). Science 260, 778-83.

# 96-plex amplification of 96 selected genomic fragments

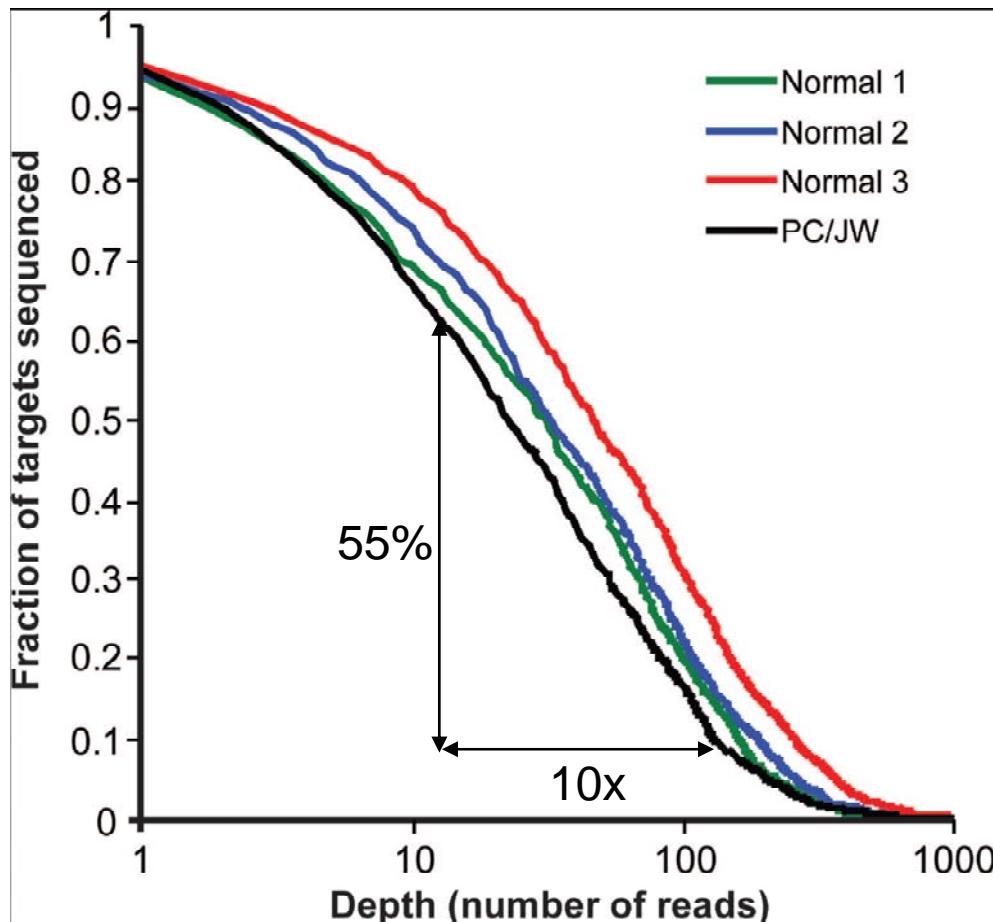


7,5k cDNA array

- 89% efficiency  
(96% <60% GC-content)

Dahl *et al*, (2005) *Nucleic Acids Res.*, 33, e71

# Parallel sequencing of 177 exons in 10 cancer genes



454 sequencing  
59.000 reads per exp

90% map to the selected sequence (49 kb) =>  
Average enrichment:  
600.000

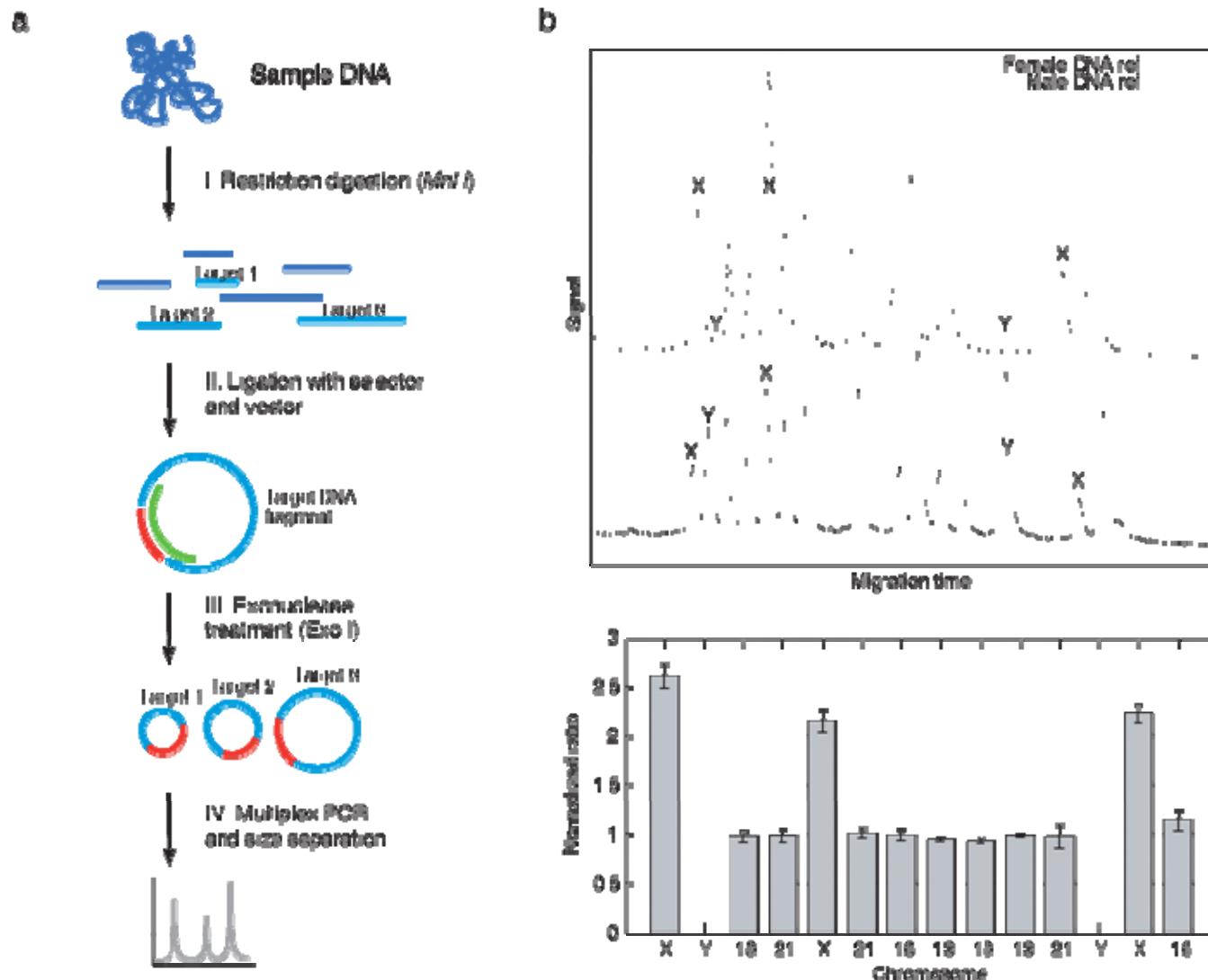
508 probes

Dahl, *et al.* (2007) PNAS, **104**, 9387-9392

# Further development

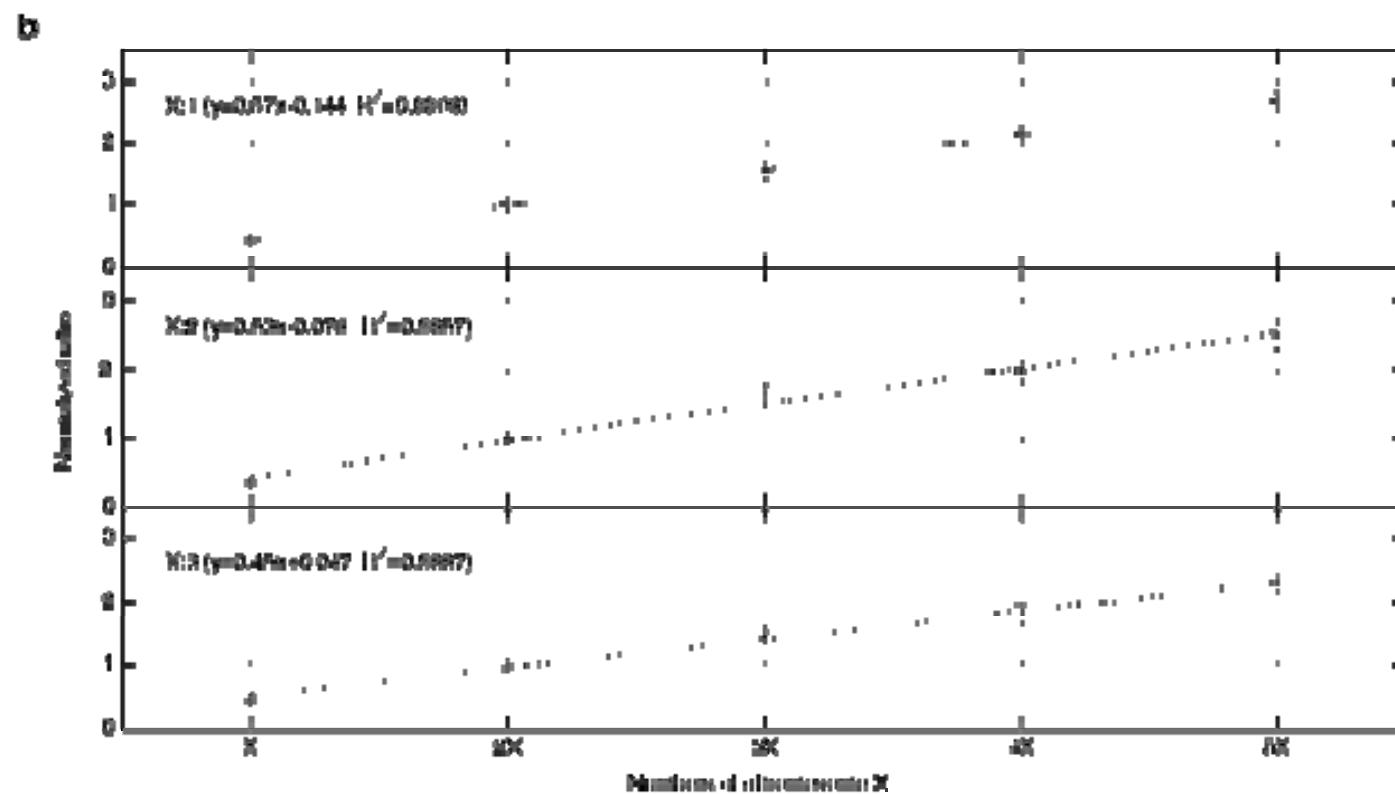
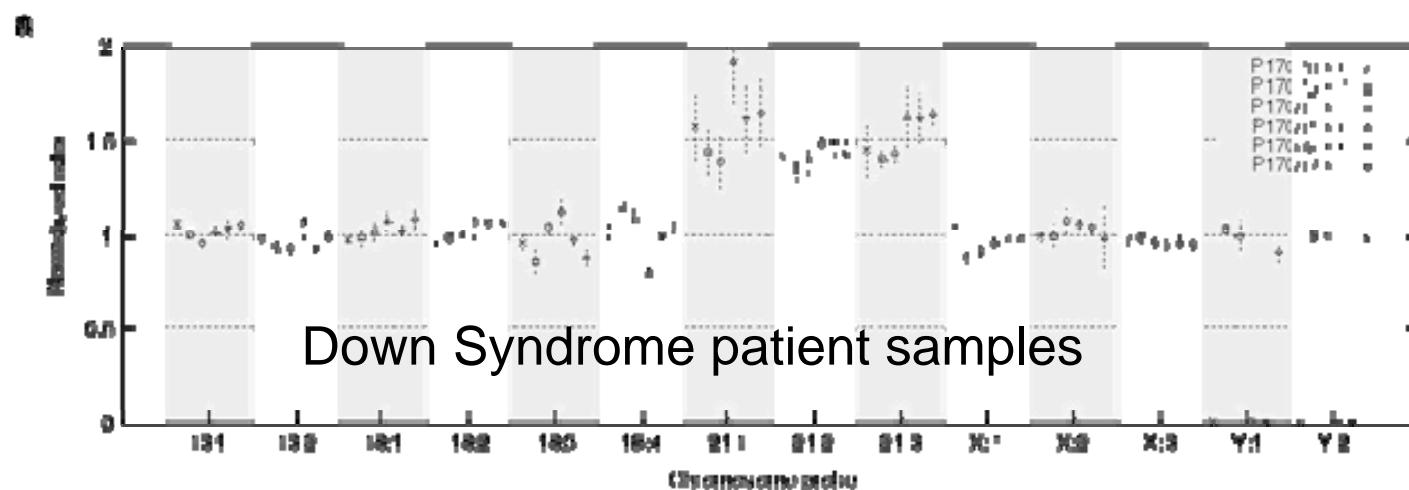
- New probe design
  - reduced cost (4 EUR/probe)
  - More even representation (5-fold range)
  - Compatible with Solexa and SOLiD seq.
- Double the size of selected fragments
- Less input DNA (100 ng)
- Applied to a cancer resequencing project with Tobias Sjöblom at UU.
- Spun out a company: **OLINK**  
GENOMICS

# Copy-number measurements

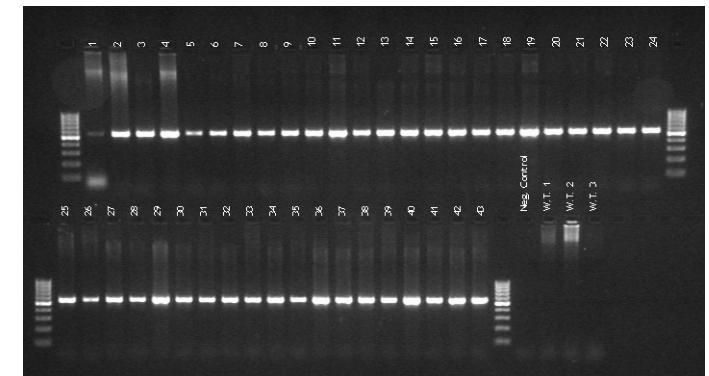
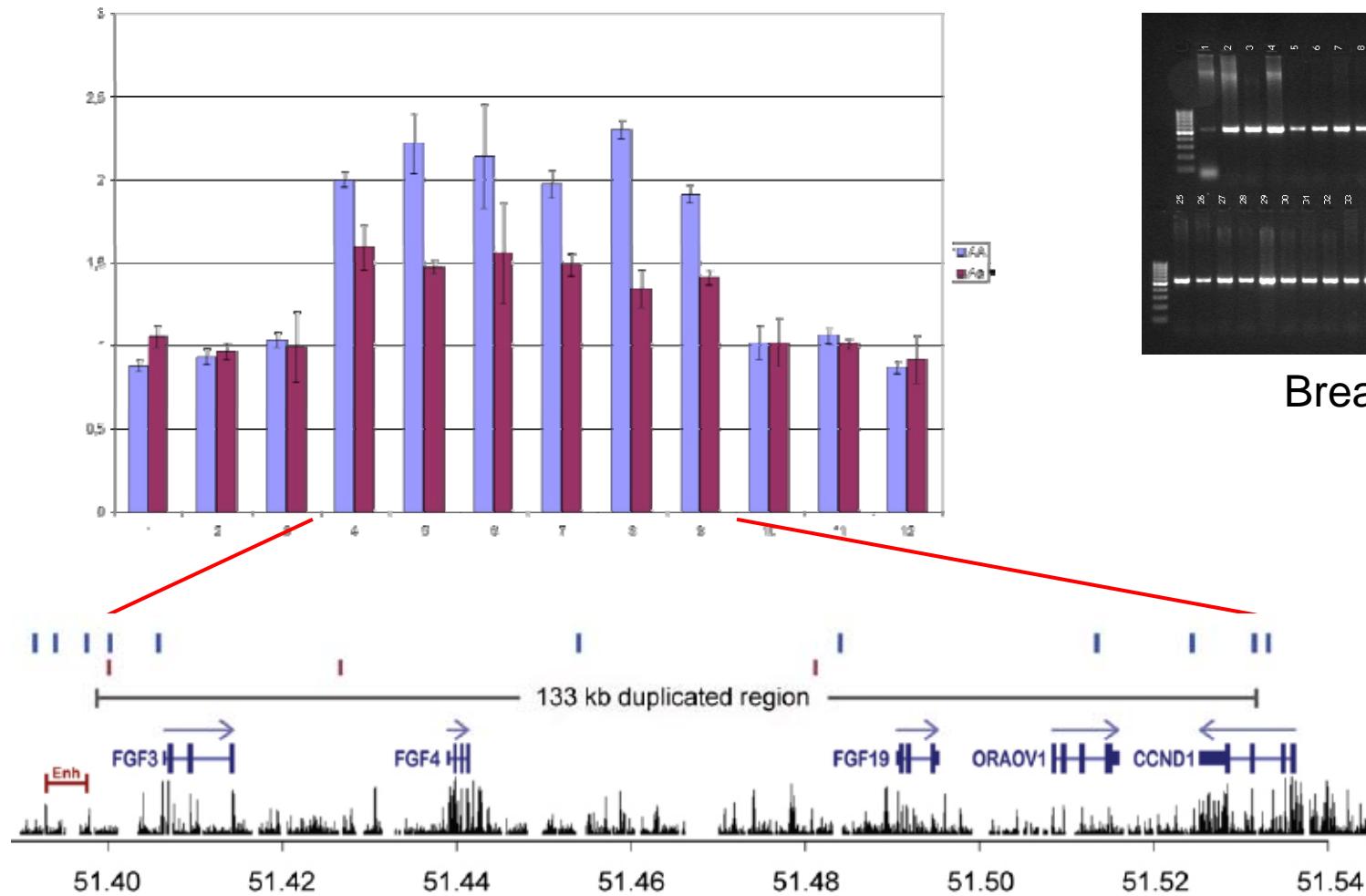


Isaksson *et al.* Nucleic Acids Res., 35, e115 (2007)

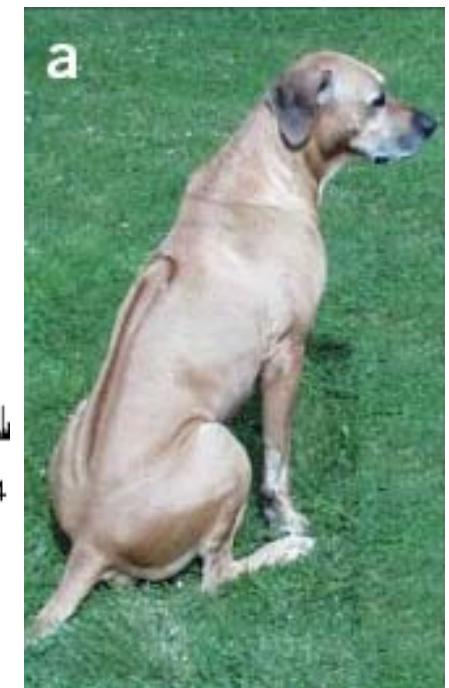
# Copy-number measurements



# Sizing a duplication in Rhodesian ridgeback dogs and mapping the break-point

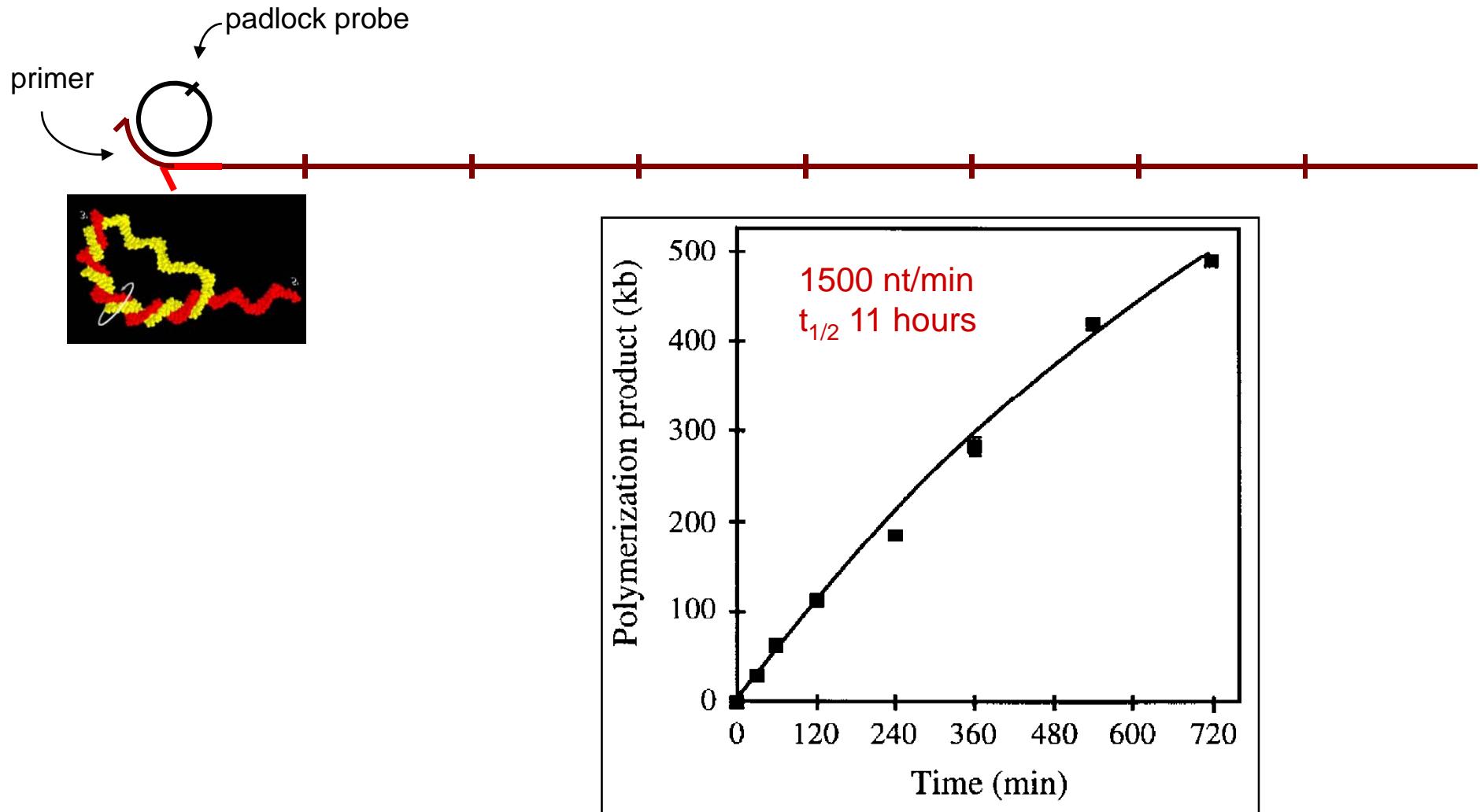


Breakpoint PCR



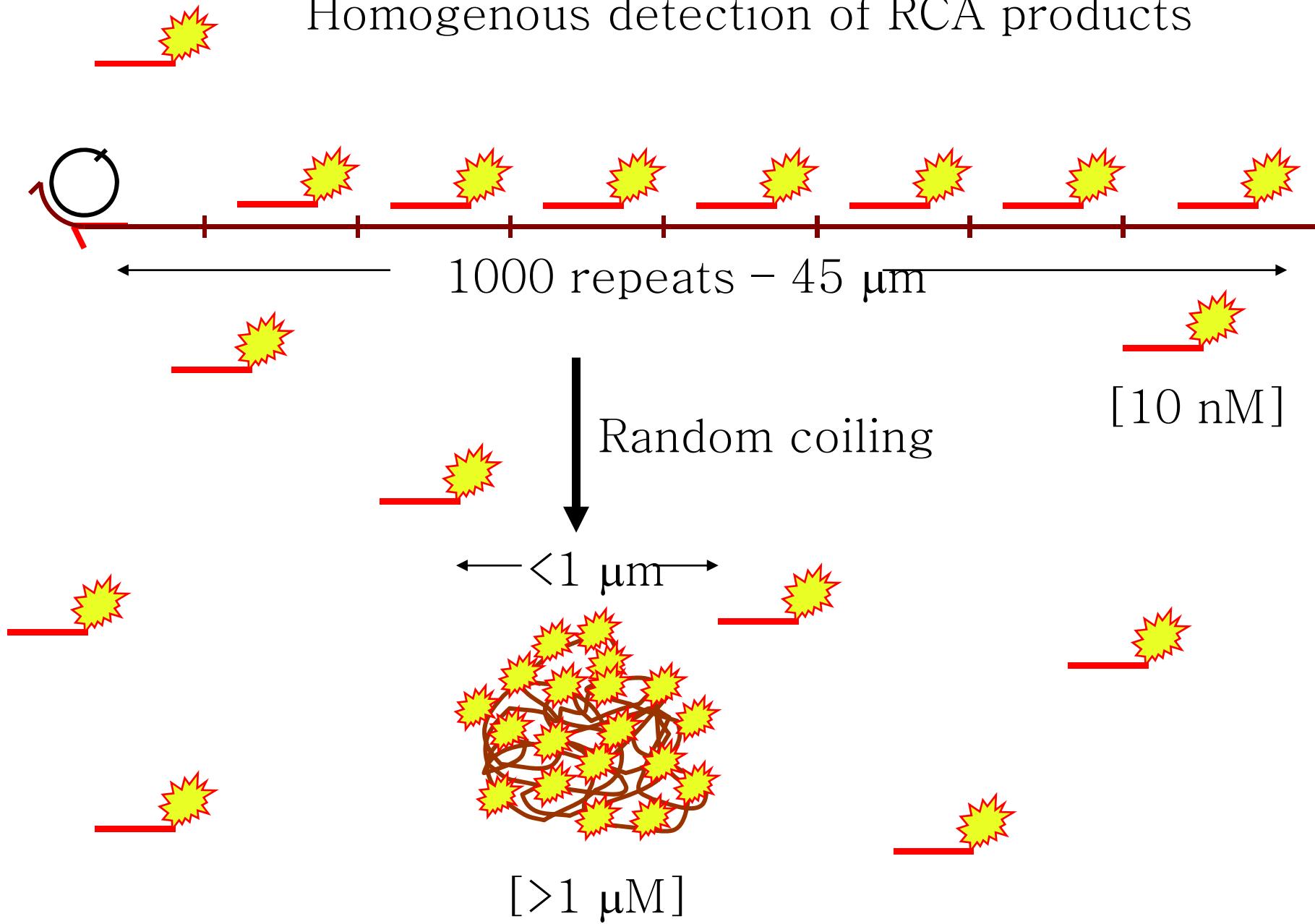
Salmon-Hillbertz, *et al.* (2007) *Nature Genet.*, 39, 1318

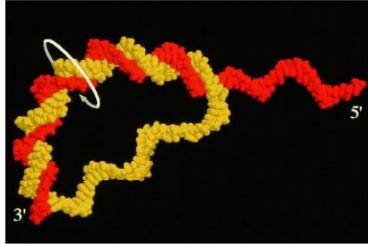
## Signal amplification through rolling circle amplification (RCA)



Banér, et al. (1998) *Nucleic Acids Res*, **22**, 5073-5078.

## Homogenous detection of RCA products

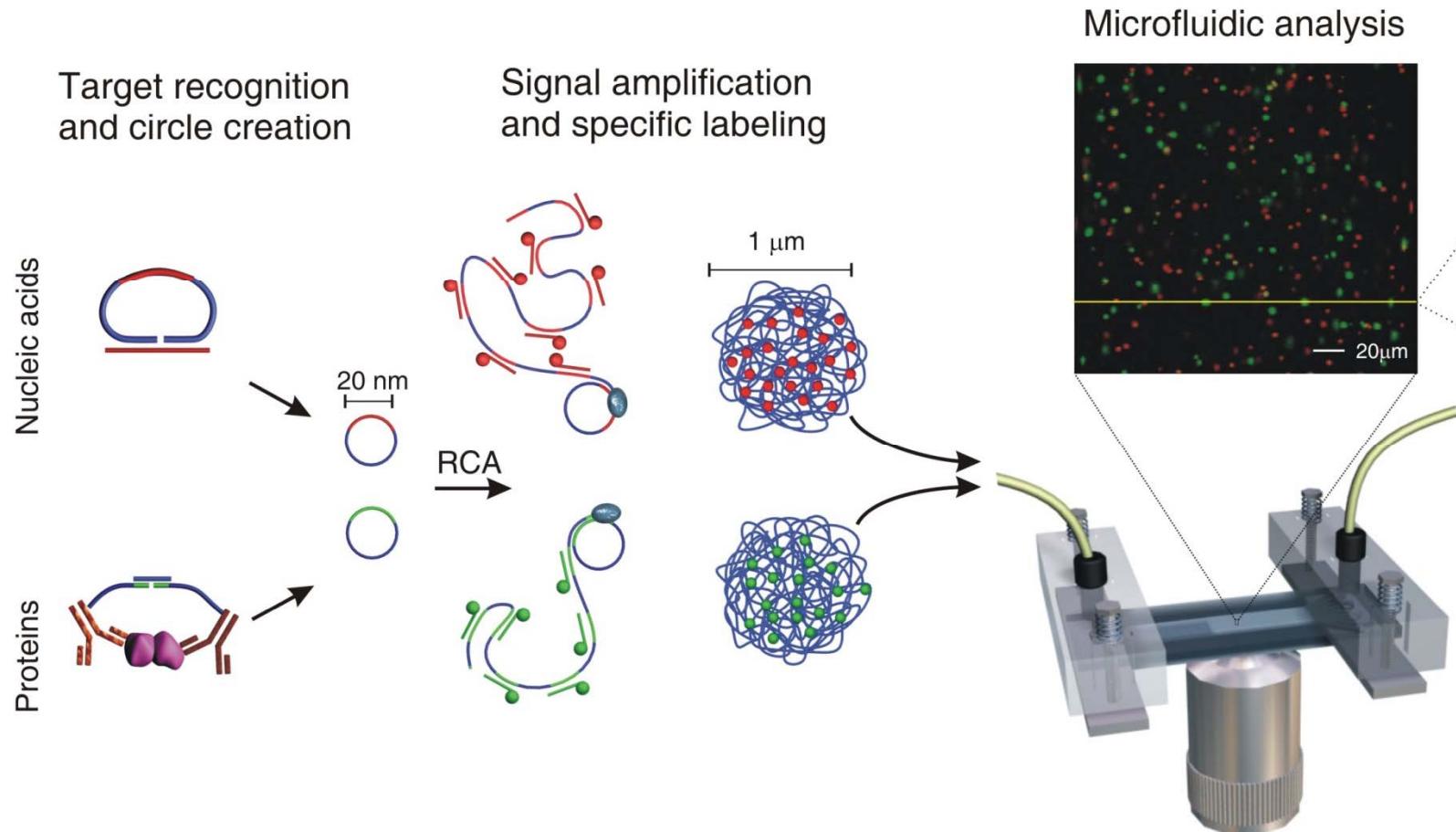




Homogenous detection of RCA products

Blab *et al.* (2004) *Anal. Chem.*, 76, 495-8

# Amplified single-molecule detection (digital RCA)

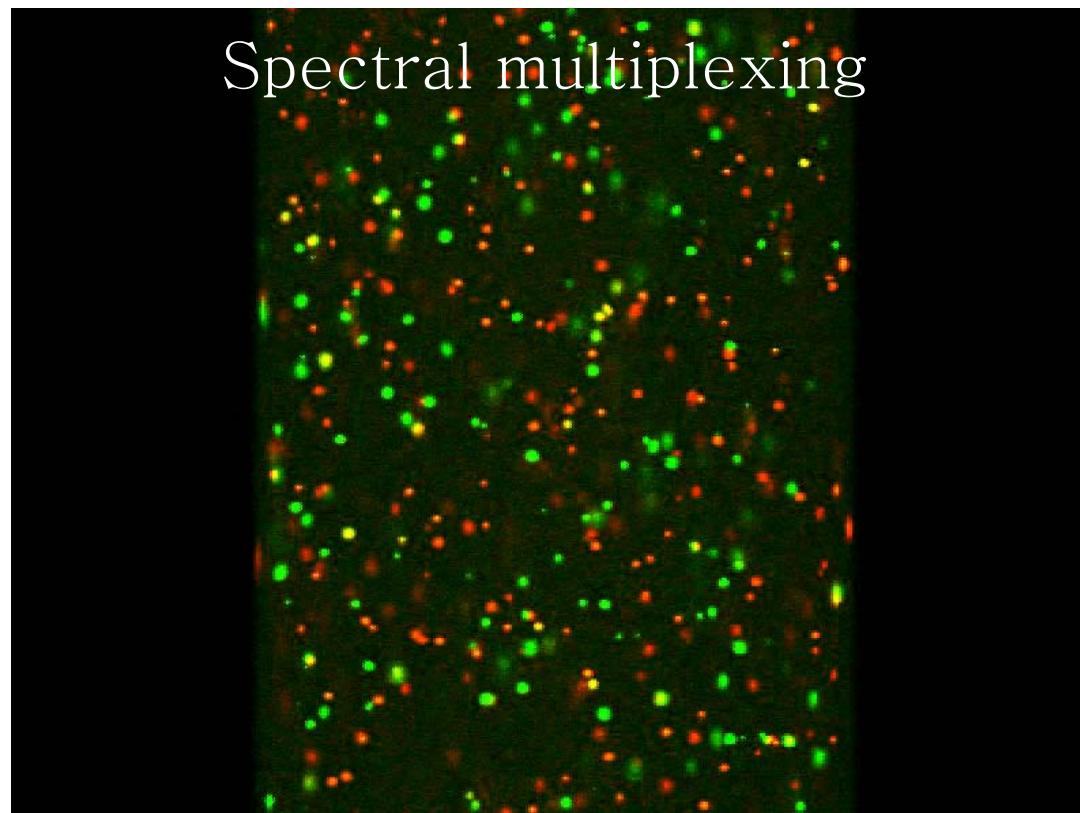


Jarvius *et al.* (2006) *Nature Methods*, 3, 725-727

# Amplified single-molecule detection

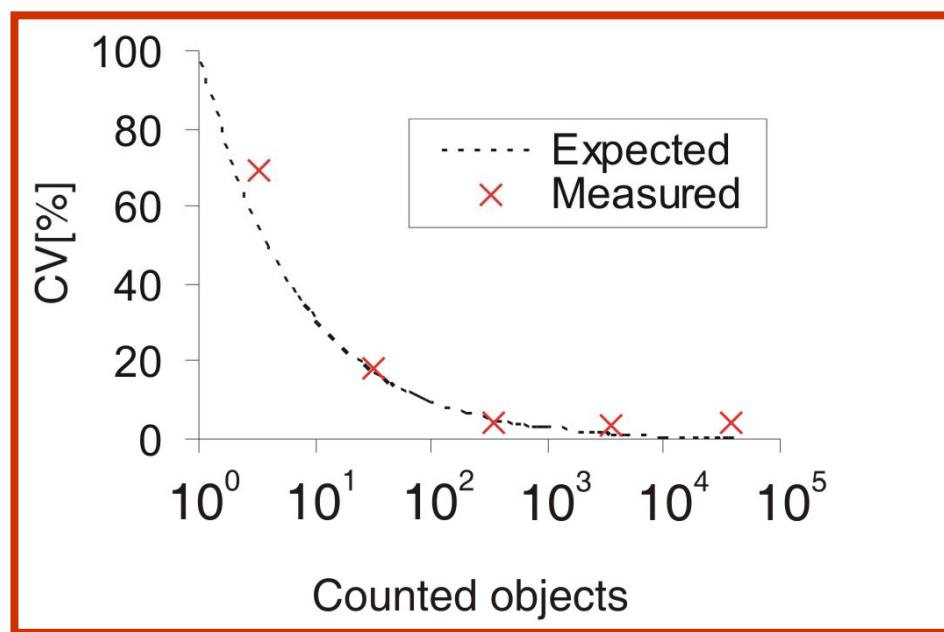
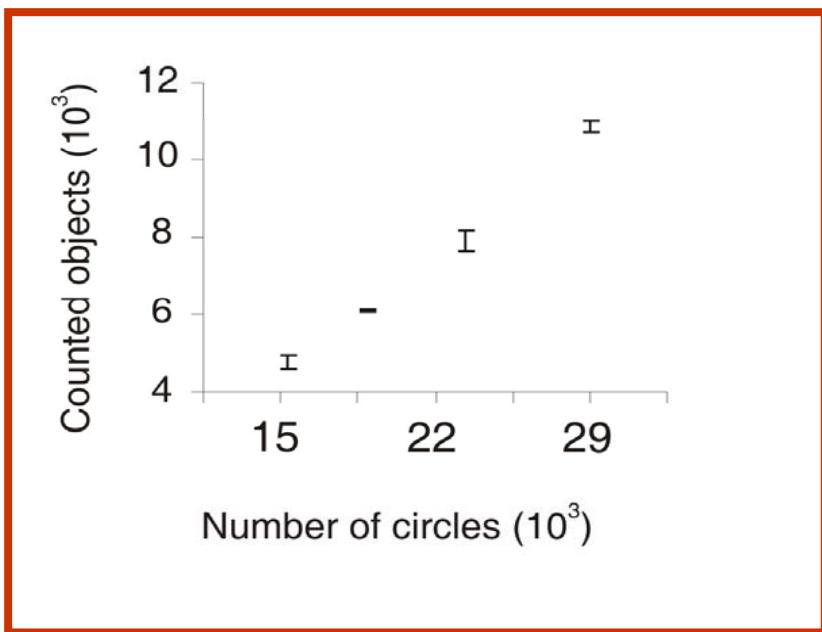
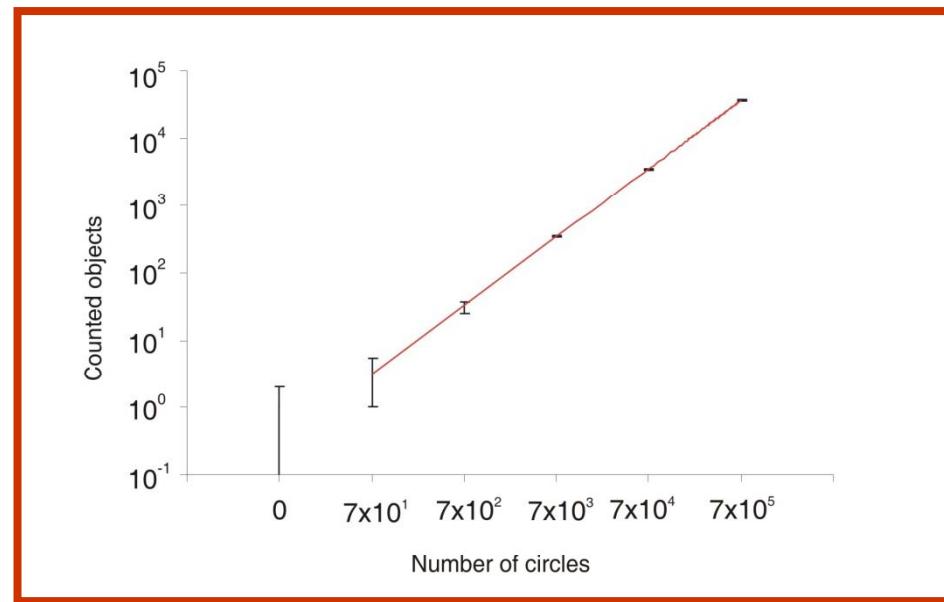
Advantages:

- Ultimate detection sensitivity
- Ultimate quantitative precision



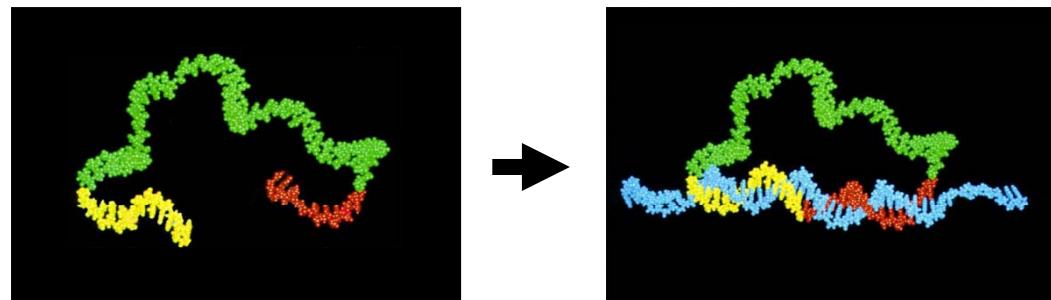
# Characterization of ASMD key properties

- Dynamic range  $10^4$ 
  - 30 s data acquisition
- High quantitative precision
  - Typically 3 percent
  - Poisson limited (<1000 counts)
- High resolving power

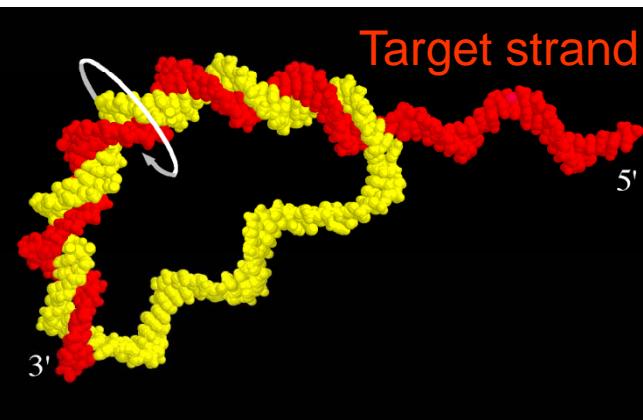


# In situ genotyping

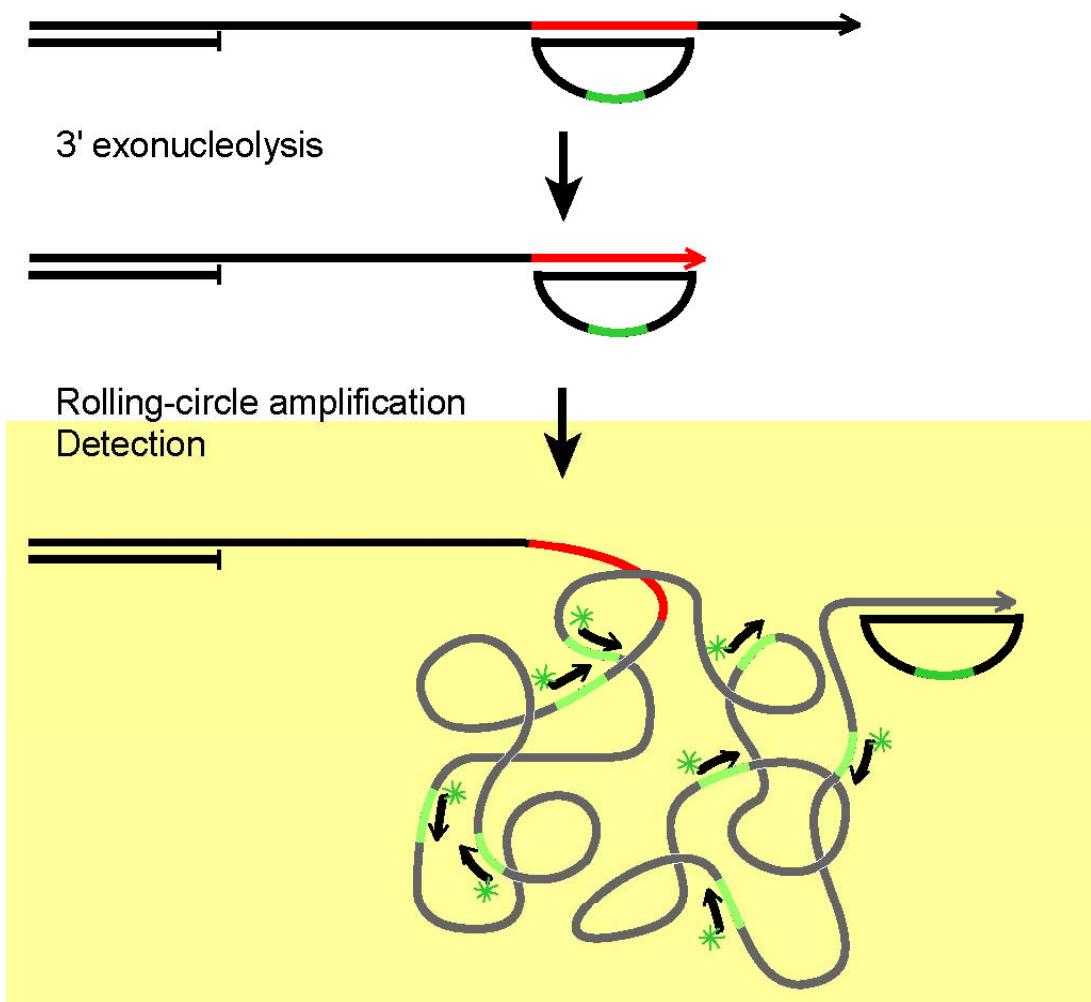
- Padlock probing



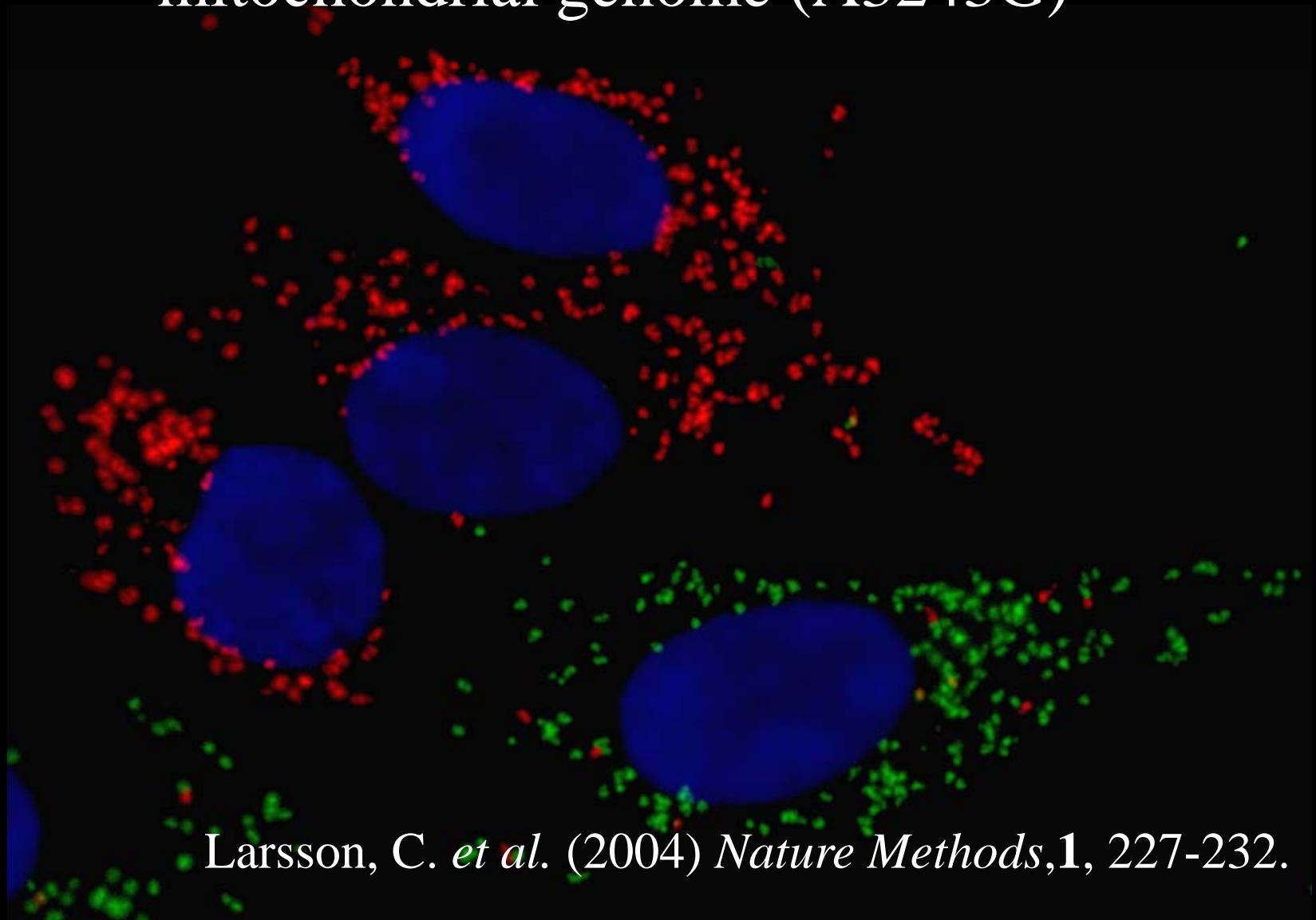
- Rolling-circle amplification



# Target-primed RCA



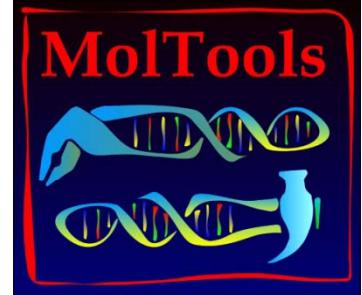
# Detection of single nucleotide variation in the mitochondrial genome (A3243G)



Larsson, C. *et al.* (2004) *Nature Methods*, **1**, 227-232.



# Acknowledgements



- Rudbeck lab, UU
    - Johan Banér
    - Mats Gullberg
    - Chatarina Larsson
    - Sara Henriksson
    - Jonas Jarvius
    - Jonas Melin
    - Henrik Johansson
    - Yuki Tanaka
    - Jenny Göransson
    - Fredrik Dahl
    - Magnus Isaksson
    - Johan Stenberg
    - Ulf Landegren
  - Collaborators at Rudbeck
    - Ola Söderberg
    - Marie-Louise Bondeson
    - Lotta Thuresson
    - Niklas Dahl
    - Fredrik Öberg
    - Fredrik Pontén
    - Tobias Sjöblom
    - Marie Allen
  - SLU/UU/Broad Inst.
    - Leif Andersson
    - Göran Andersson
    - Kerstin Lindblad-Toh
  - Ångström lab, UU
    - Fredrik Nikolajeff
    - Maria Strömme
  - Image Analysis, UU
    - Carolina Wählby
    - Ewert Bengtsson
  - Aarhus University
    - Jörn Koch
  - Leiden University
    - Ton Raap
    - Thomas Schmidt
  - University of Tokyo
    - Takehiko Kitamori
  - Nagoya University
    - Yoshinobu Baba
  - Stanford/ParAllele/Affymetrix
    - Paul Hardenbol *et al.*
  - Karolinska Institute
    - Nils-Göran Larsson
- EU FP-7 READNA
  - EU FP-6 "COMICS"
  - Swedish Research Councils (M & NT)
  - Uppsala BioX
  - The Wallenberg Foundation
  - Swedish Defence Nanotech Program
  - VINNOVA/SSF/JST