

# Establishment of the 1st World Health Organization International Genetic Reference Panel for quantitation of BCR-ABL mRNA



HE White<sup>1</sup>, P Matejtschuk<sup>2</sup>, P Rigsby<sup>2</sup>, J Gabert<sup>3</sup>, YL Wang<sup>4</sup>, S Branford<sup>5</sup>, MC Müller<sup>6</sup>, N Beaufils<sup>3</sup>, E Beillard<sup>7</sup>, D Colomer<sup>8</sup>, D Dvorakova<sup>9</sup>, H Ehrencrona<sup>10</sup>, H Goh<sup>11</sup>, H El Housni<sup>12</sup>, D Jones<sup>13</sup>, V Kairisto<sup>14</sup>, S Kamel-Reid<sup>15</sup>, D-W Kim<sup>11</sup>, S Langabeer<sup>16</sup>, ESK Ma<sup>17</sup>, R Press<sup>18</sup>, G Romeo<sup>19</sup>, L Wang<sup>20</sup>, K Zoi<sup>21</sup>, T Hughes<sup>5</sup>, G Saglio<sup>22</sup>, A Hochhaus<sup>23</sup>, J Goldman<sup>24</sup>, P Metcalfe<sup>2</sup>, NCP Cross<sup>1</sup>

<sup>1</sup>National Genetics Reference Laboratory (Wessex), Salisbury District Hospital, UK. <sup>2</sup> National Institute for Biological Standards & Control, South Mimms, UK. <sup>3</sup> IFR Jean Roche, Faculté de médecine NORD, Marseille cedex 20, France. <sup>4</sup>Molecular Hematopathology Laboratory, Dept of Pathology & Laboratory Medicine, Weili Medical College of Cornell University, New York, NY 10021-4873, USA. <sup>5</sup>Division of Molecular Pathology, Institute of Medical and Veterinary Science, Adelaide, South Australia 5000. <sup>6</sup>III. Medizinische Klinik, Medizinische Fakultät Mannheim der Universität Heidelberg, Mannheim, Germany. <sup>7</sup> Dept of Cancer & Stem Cell Biology - Duke-NUS - Singapore 169857. <sup>8</sup> Unitat D' hematopatologia, Hospital Clinic, 08036 Barcelona, Spain. <sup>9</sup> University Hospital Brno, Center of Molecular Biology and Gene Therapy, 625 00 Brno, Czech Republic. <sup>10</sup> Dept of Genetics & Pathology, Uppsala University, 5 751 85 Uppsala, Sweden. <sup>11</sup>Molecular Genetics Research Institute, The Catholic University of Korea, Seocho-gu, Seoul 137-701, Republic of Korea <sup>12</sup> Erasme hospital, Medical Genetics Dept, 1070 Brussels, Belgium. <sup>13</sup>Molecular Diagnostics Laboratory, The University Health Network, Toronto, Ontario, Canada. <sup>16</sup>Cancer Molecular Diagnostics, Central Pathology Laboratory, St. James's Hospital, Dublin 8, Republic of Ireland. <sup>17</sup>Hong Kong Sanatorium & Hospital, Happy Valley, Hong Kong. <sup>18</sup>Oregon Health & Science University, Dept of Pathology, Portland, Oregon 97201 USA. <sup>19</sup>Molecular Heamatology Laboratory, PathWest Laboratory Medicine, Royal Perth Hospital, Wellington Street, Perth, Western Australia 6000. <sup>20</sup> Dept of Haematology, Royal Liverpool Universit Haematology Research Laboratory, Foundation of Biomedical Research, Academy of Athens, Athens, Greece. <sup>22</sup> Dept of Clinical & Biological Science, University of Turin, Turin, Italy. <sup>20</sup>Dept Hematology/ Oncology, Universititskinikum Jena, Jena, Germany. <sup>24</sup>Dept of Haematology, Inperial College Academic Health Science Centre, London, UK.

## Introduction

• Estimation of *BCR-ABL* mRNA levels is an important indicator of therapeutic response for patients with chronic myeloid leukemia and Philadelphia-chromosome positive acute lymphoblastic leukemia, but there is substantial variation in the real time quantitative PCR (RQ-PCR) methodology employed, what control gene is used for normalisation and how results are reported

• An international scale (IS) has recently been established that is anchored to two key points defined in the IRIS trial: a common baseline (100% BCR-ABL<sup>IS</sup>) and major molecular response (0.1% BCR-ABL<sup>IS</sup>)

• To help propagate and improve the accessibility of the IS we sought to develop internationally accredited BCR-ABL reference reagents.

• Following an evaluation of candidate cell lines, the aim of the international collaborative study was to manufacture and evaluate a reference material panel comprising four different dilution levels of freeze dried preparations of K562 cells diluted in HL60 cells and assign fixed % BCR-ABL / control gene IS values to each material

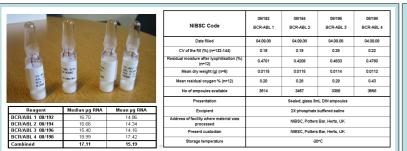
## Methods

Development of materials was co-ordinated by the UK National Genetics Reference Laboratory (Wessex) in conjunction with the National Institute for Biological Standards and Control • Four dilutions of K562 in HL60 corresponding approximately to 10%IS (*BCR/ABL* 4 08/198), 1%IS (*BCR/ABL* 3 08/196), 0.1%IS (*BCR/ABL* 2 08/194) and 0.01%IS (*BCR/ABL* 1 08/192) *BCR-ABL / ABL* were prepared.

• 3ml glass ampoules were filled with 1.5 x 10<sup>6</sup> cells (0.5ml) for freeze drying and approximately 3500 vials of each material were produced

- Field trial evaluation of the reagents was performed by 10 laboratories with stable and validated conversion factors (6 European Union, 1 Canada, 3 Asia/Australasia) RNA was extracted from each material and, following reverse transcription, cDNA was tested in lab specific qPCR assays.
- Transcript copy numbers for BCR-ABL and the three control genes were used to calculate %BCR-ABL/ABL, %BCR-ABL/BCR and %BCR-ABL/GUSB.
- The mean % values for each material were then converted to the IS by multiplying the % BCR-ABL/control gene value by the established CF for each laboratory.

## Results



**Figure 1: RNA Extraction and product summaries for the four reference materials.** The median and mean quantity of RNA extracted from each vial (1.5 x 10<sup>6</sup> cells / vial) was 17.11µg and 15.19µg respectively (n=30). The product summaries fulfilled the requirements of the WHO for establishment of the materials as an international genetics reference panel

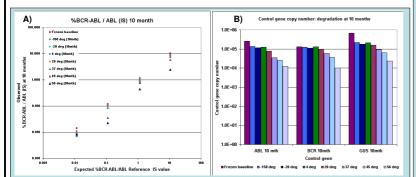


Figure 2: Accelerated degradation study. A) %*BCR-ABL*/ ABL IS at 10 months post fill. The *BCR-ABL*/ABL ratios per fixed volume of cDNA for each of the four reference materials are shown after storage at the designated temperatures for 10 months. B) Mean total copy number of *ABL*, *BCR* and *GUSB* control gene transcripts per fixed volume of cDNA for the four reference materials after storage at the designated temperatures for 10 months. These data demonstrate that the materials are acceptable for analysis if stored at or below +37°C for 10 months

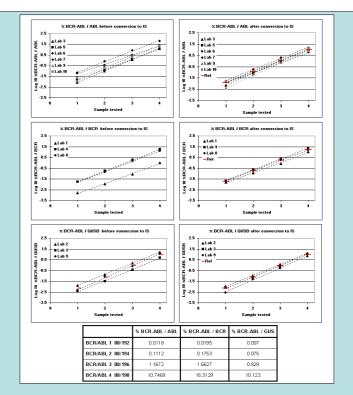


Figure 3: Linear regression plots of log transformed %*BCR-ABL/* control gene. (Sample 1; BCR/ABL 1 08/192, Sample 2; BCR/ABL 2 08/194, Sample 3; BCR/ABL 3 08/196 & Sample 4 BCR/ABL 4 08/198). Regression plots are shown for each control gene before and after conversion to the IS. The red lines indicated the IS reference values assigned for each material as designated in the table. The assigned IS values were slightly different for each control gene as the relative level of expression of these three genes in HL60 cells is not identical to that seen in peripheral blood leukocytes.

# Summary

This study indicates that the freeze dried materials are suitable for use as reference materials for the quantitation of *BCR-ABL* mRNA (b3a2 transcript) using RQ-PCR. The materials were approved as the 1st International Genetic Reference Panel quantitation of *BCR-ABL* translocation by RQ-PCR by the Expert Committee on Biological Standardization of the World Health Organization in November 2009 and are available from NIBSC (www.nibsc.ac.uk) for the calibration of secondary reference reagents.

Acknowledgements: This work was supported by funding from the UK Dept of Health. We thank all study participants; Chris Morris, Alex Hiscott (ECACC) & Feng Lin (NGRL W) for assistance with cell culture; Elaine Gray & Ross Hawkins (NIBSC) for assistance with field trial design; DSMZ, Braunschweig, Germany & Prof John Goldman, Dept of Haematology, Imperial College Academic Health Science Centre, London, UK for use of the cell lines.