

New techniques for DNA methylation analysis

Helen White

**NGRL (Wessex)
Salisbury NHS Foundation Trust**

Techniques

- ❖ MS-MLPA
- ❖ Pyrosequencing
- ❖ High resolution melt curve analysis
- ❖ Mass Spectrometry

Prader Willi and Angelman Syndromes

Two clinically distinct phenotypes that map to 15q11-q13

PWS

- Caused by loss of the paternal (unmethylated) contribution
 - Paternal deletion (~70%)
 - Maternal UPD (~30% cases)
 - Mutation in the imprinting region causing abnormal methylation (<2%)

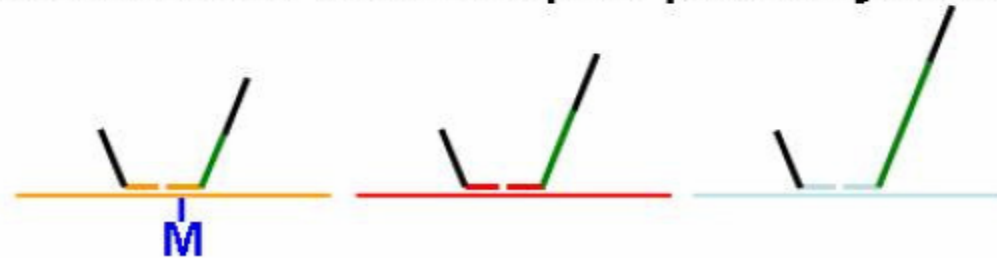
AS

- Caused by loss Maternal (methylated) contribution
 - Maternal deletion (~70%)
 - Paternal UPD (~5% cases)
 - Mutation in the imprinting region causing abnormal methylation (~5%)
- A single gene, UBE3A has been implicated as the AS gene

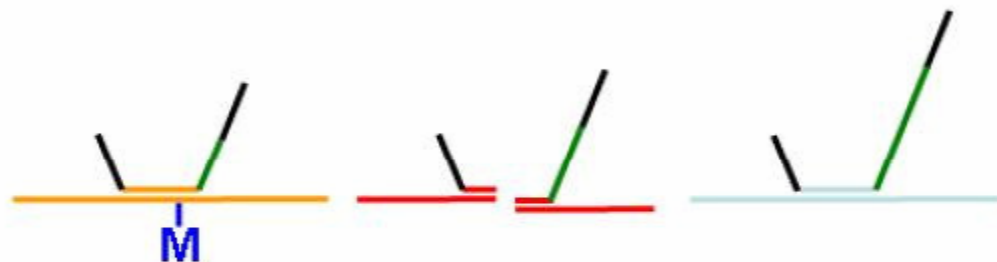
MS-MLPA



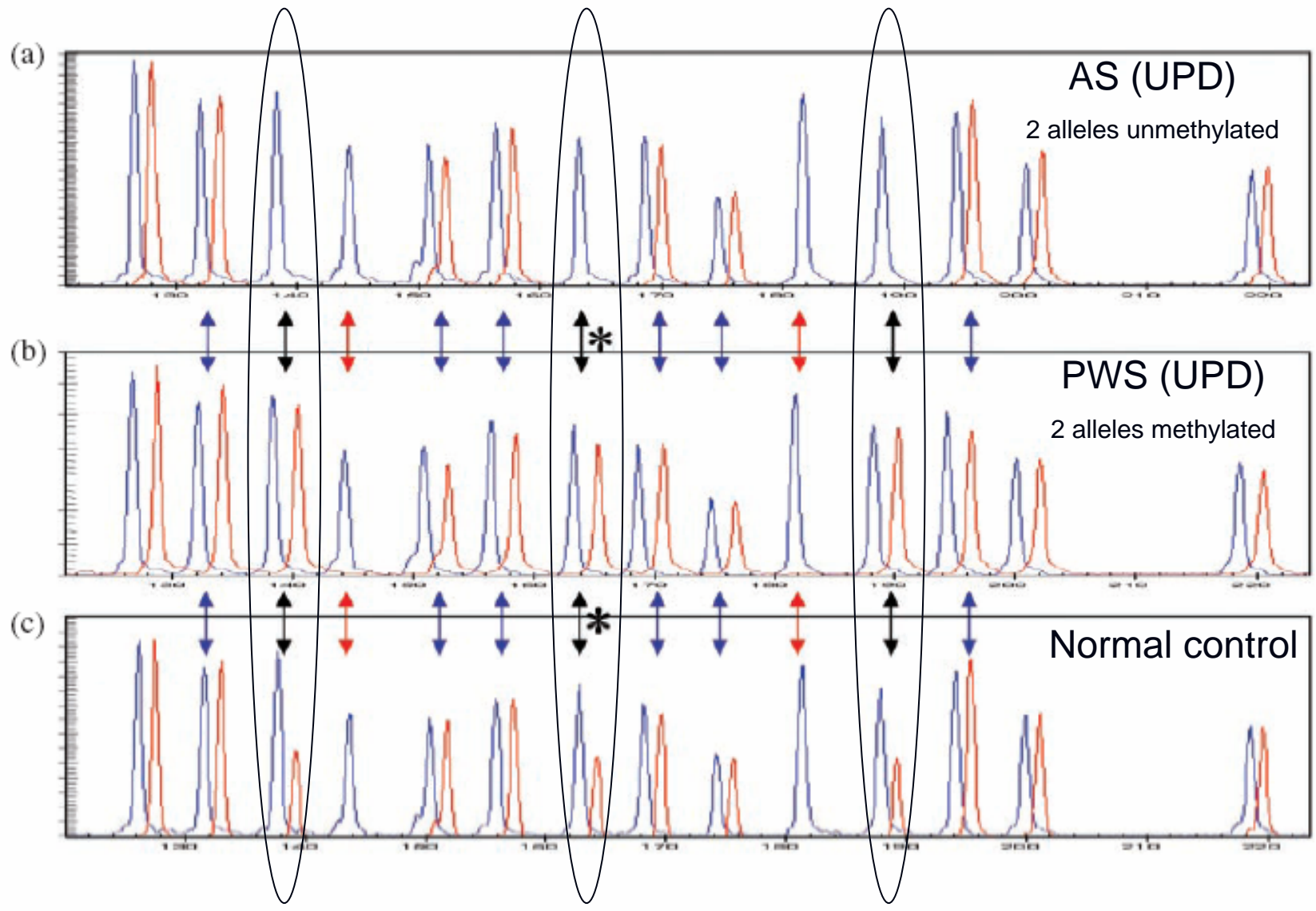
1. Denaturation and Multiplex probe hybridization



2. Simultaneously Ligation and Digestion with methylation sensitive *endonucleases*



3. PCR using one universal primer pair Only undigested (methylated) and ligated probes are exponentially amplified



— Undigested

— Digested

Advantages

- No bisulphite treatment required
- Multiple targets tested simultaneously
- Genomic and epigenomic information
- Relative quantitation

Disadvantages

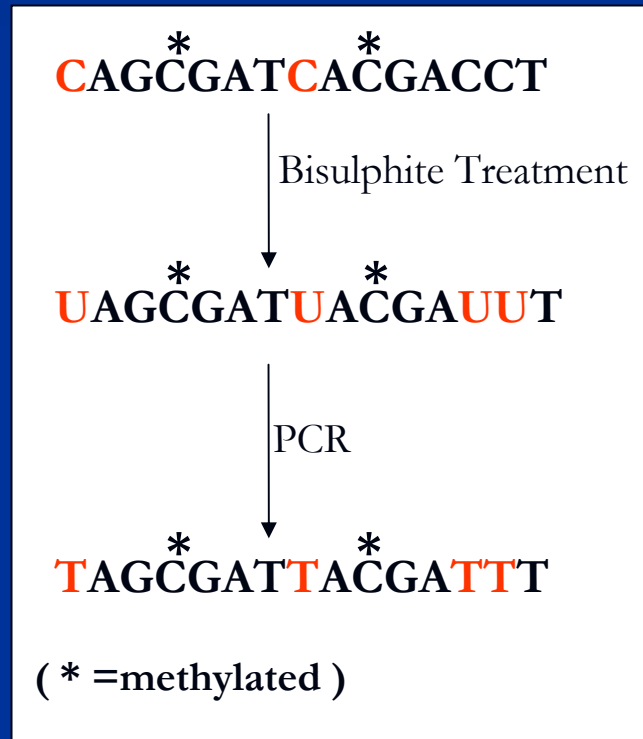
- Possibly not as sensitive as bisulphite based techniques
- Single base variations at HhaI site may results in false positive/negatives

Techniques

- ❖ MS-MLPA
- ❖ Pyrosequencing
- ❖ High resolution melt curve analysis
- ❖ Mass Spectrometry

Bisulphite Treatment

- Bisulphite treatment causes unmethylated Cytosines to convert to Uracil while methylated cytosines remain unchanged.



**SNRPN 1
(Genomic)**

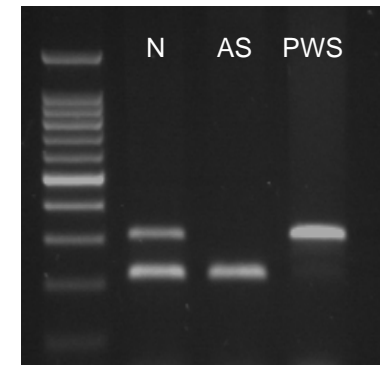
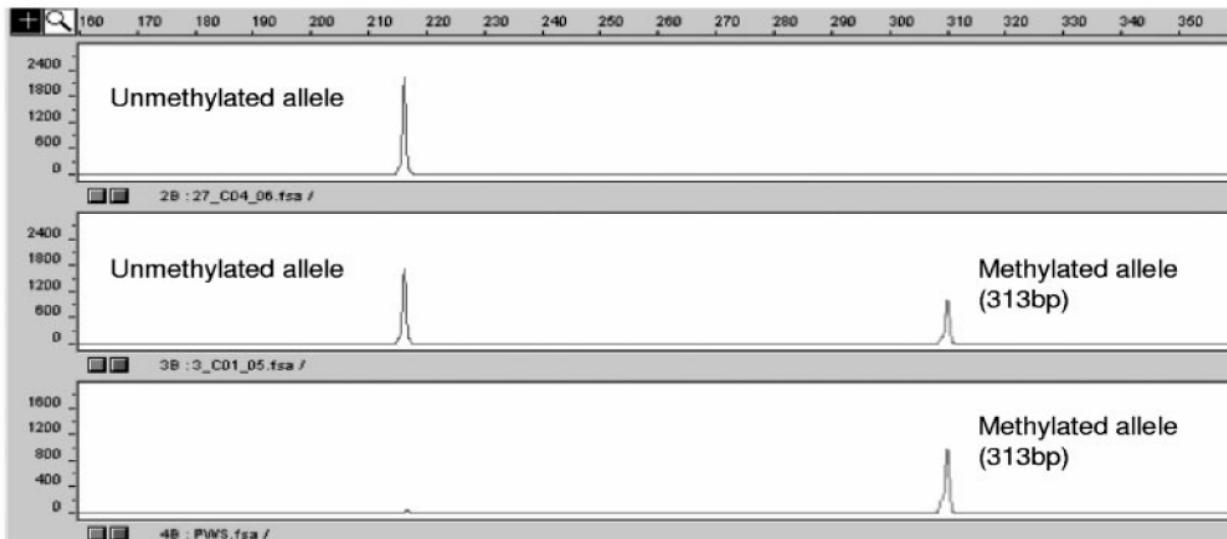
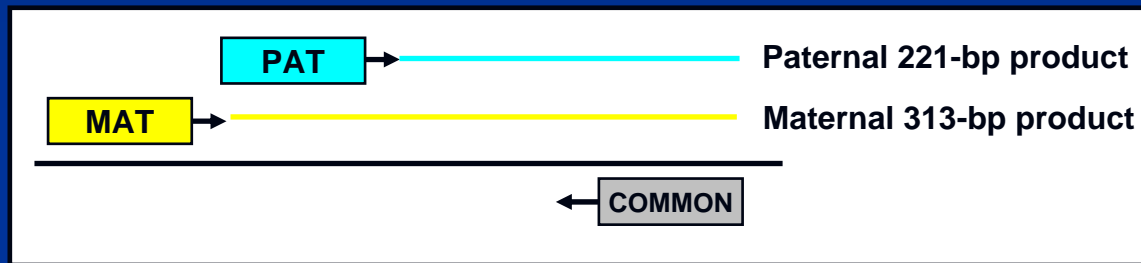
```
TGGGGCC CTAGGGTCCAGTAGC CCCCTCCCCC)AGGTCATT CGGTGAGGGAGGGAGCTGG  
GACCCCTGCACTGCGGCAAACAAGCCGCTGCGCGGCCGCAGAGGCAGGCTGGCGCGCATG  
CTCAGGCGGGGATGTGTGCGAAGCCTGCGCTGCTGCAGCGACTCTGCGCAGAGTGGAGCG  
GCCGCGGAGATGCCTGACGCATCTGTCTGAGGAGCGGTCAGTGACCGCGATGGAGCGGGCAA  
GGTCAGCTGTGCCGGTGGCTTCTCTCAAGAGACAGCCTGGGGAGCGGCCACTTTTATTCATC  
AGATATTCCAAGTTTTTAGGACTTGGAGTACTGAATAAACGGAATTTGGGCCCTAAAATCCT  
TTGTTCTGGAGAACCA
```

**SNRPN 1
(After bisulphite)**

```
TGGGGTTTTAGGGGTTTAGTAGTTTTTTTTTTAGGTTATTTCGTGAGGGAGGGA  
GTTGGGATTTTTGTATTGYGGTAATAAGTAYGTTTGYGYGGTTAGAGGTAGGTT  
GGYGYGTATGTTTAGGYGGGGATGTGTGYGAAGTTTGTYGTTGTTGTAGYGAGTTTG  
GYGTAGAGTGGAGYGGTYYYGGAGATGTTTGAYGTATTTGTTTGAGGAGYGGGTTAG  
TGAYYGYGATGGAGYGGGTAAGGTTAGTTGTGTYGGTGTTTTTTTTTAAGAGATAGTT  
TGGGGAGYGGTTATTTTTATTTATTAGATATTTTAAGTTTTTAGGATTTGGAGTATT  
GAATAAAYGGAATTTGGTTTTTAAAGTTTTTTGTTTTGGAGAATTA
```

MS PCR

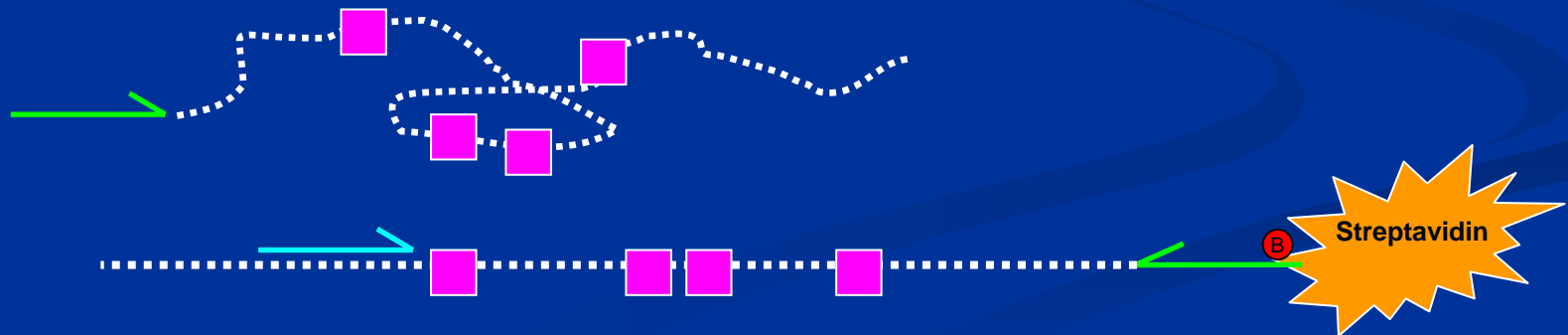
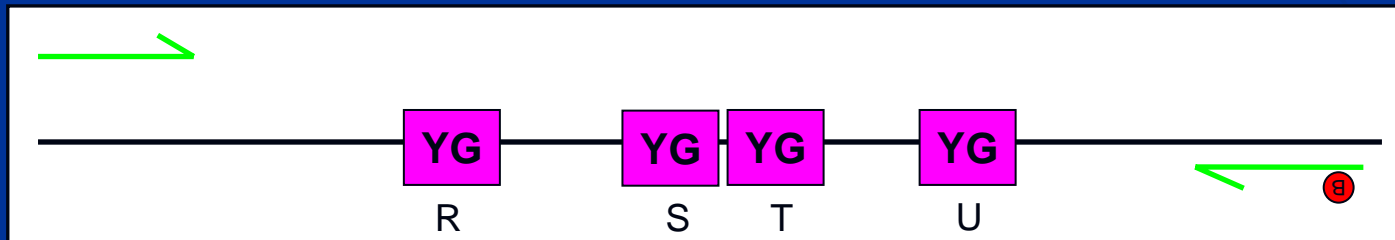
```
GATTTTGTATTGTCGTAATAAGTAYGTTTGTCYCGTYGTAGAGGTAGGTTGGYCYGTATG  
TTTAGGYGGGGATGTGTGYGAAAGTTTGTYTFTTGTGTAGYBAGTTTGYYGTAGAGTGGAGYB  
CTYCTYCGAGATGTTTGAYGTATTTGTTTGAGGAGYCGTTAGTGAYGYGATGGAGYCGCTAA  
GGTTAGTTGTGTTCGTGTTGTTTTTTTAAGAGATAGTTTGGGGAGYCGTTATTTTTTATTIATT  
AGATATTTTAAGTTTTTTAGGATTTGGA GTATT GAATAAAYGGAATTTGGGTTTTAAA GTTTT  
TTGTTTTGGAGAATTA
```



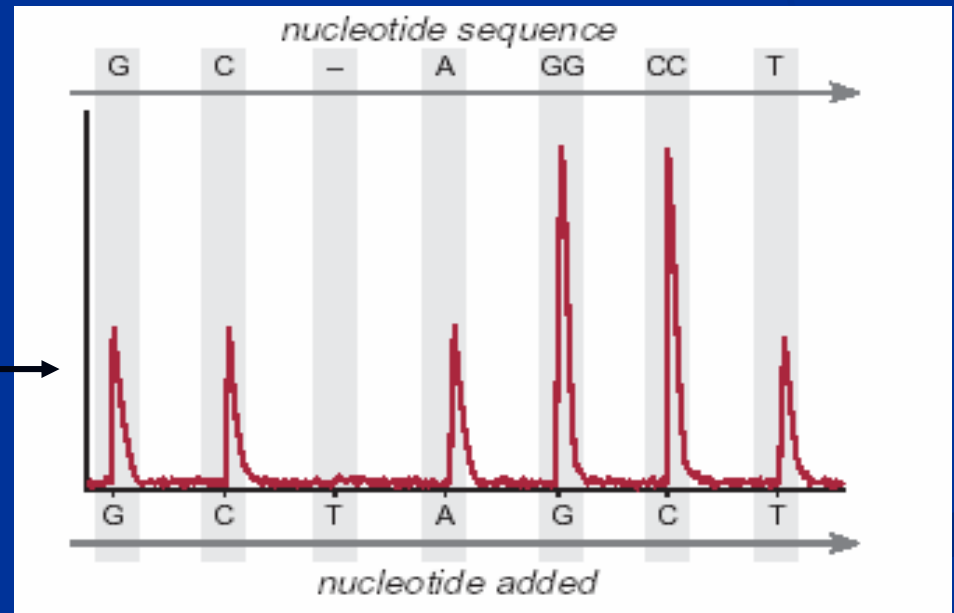
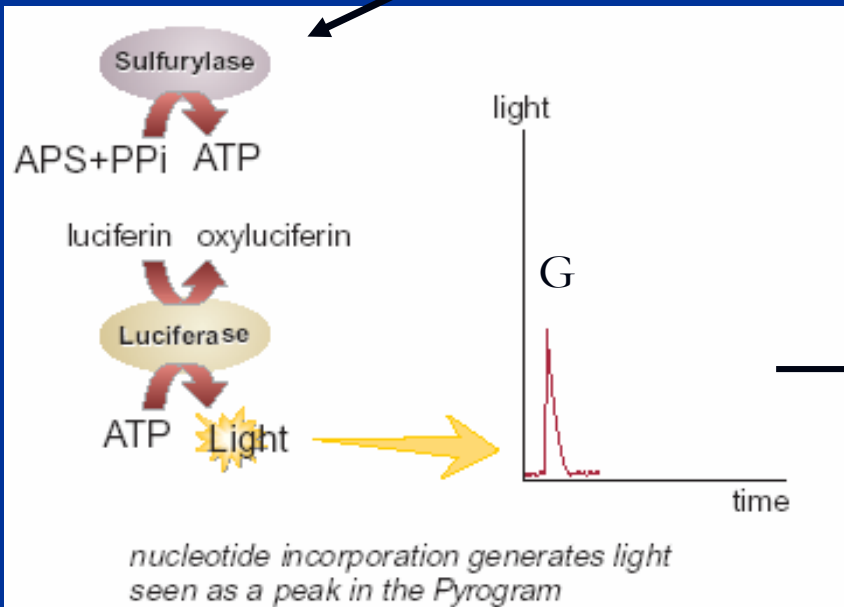
Pyrosequencing

Pyrosequencing assay design

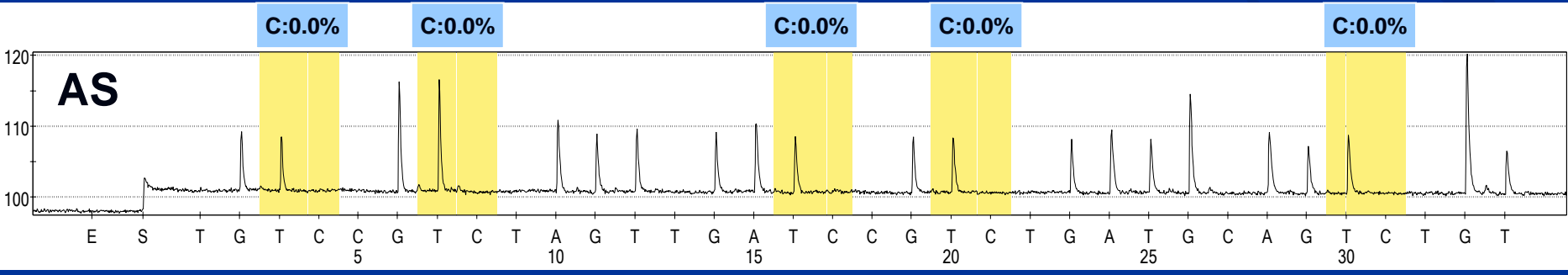
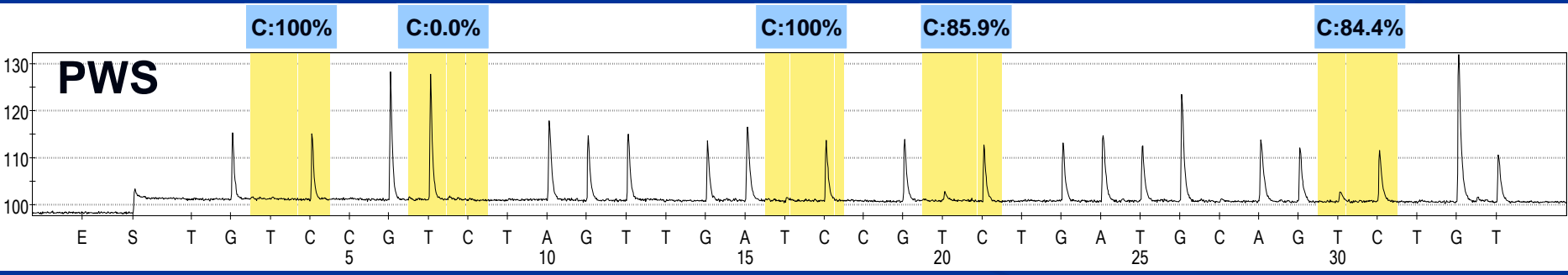
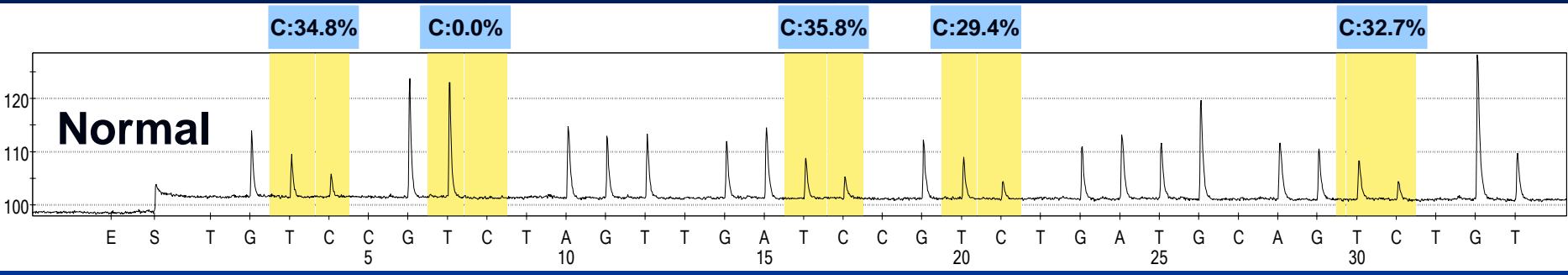
```
TGGGGTTTTAGGGGTTTTAGTAGTTTTTTTTTTTTTTAGGTTATTTYG GTGAGGG AGGGAGTTGG  
GATTTTTGTATT GYG GTA AATAAGTAYG TTTG YGYGG TYGTAGAGGTAGGTTGGYGYGTATG  
TTTAGGYYGGGGATGTGTGYGAA GTTTGT YGTT GTTGTAGYGA GTTTGG YGTA GAGTGGAGY  
GT YGTYGGAGATGTTTGAYGTA TTTGTTTGAGGAGYGGTTAGTGAYE YGATGGAGYGGGTAA  
GGTTAGTTGTGT YGGTG GTTTTTTTAAGAGATAGTTTGGGG AGYGGTTATTTTTTATTATT  
AGATATTTTAAGTTTTT AGGATTTGGAGTATTGAATAAAYGGAATTTGGGTTTTAAA GTTTT
```



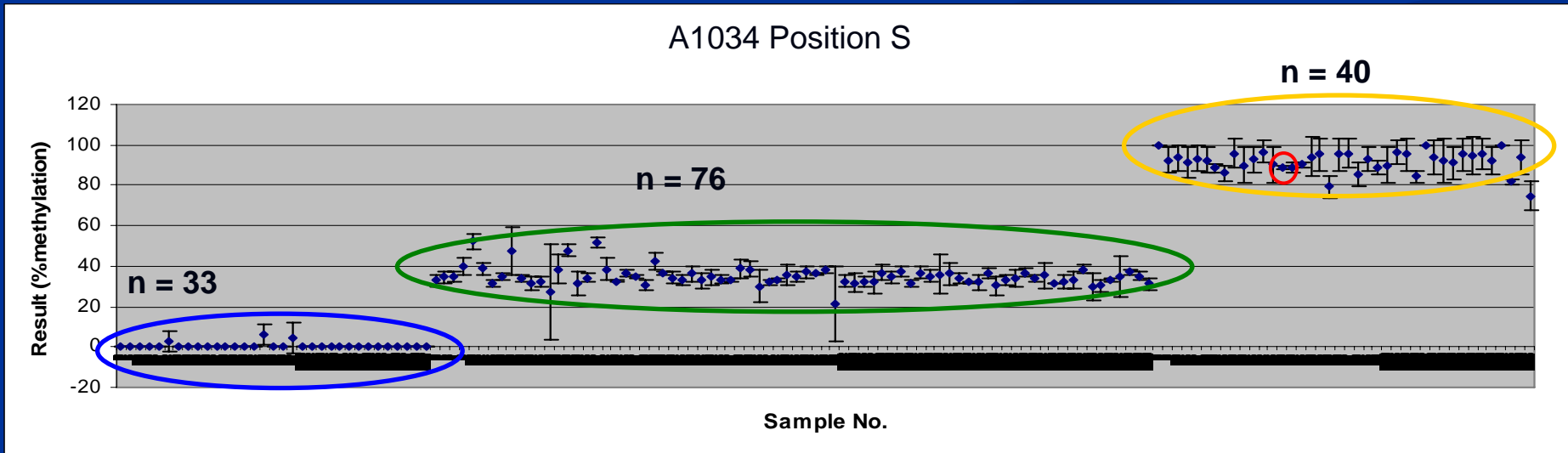
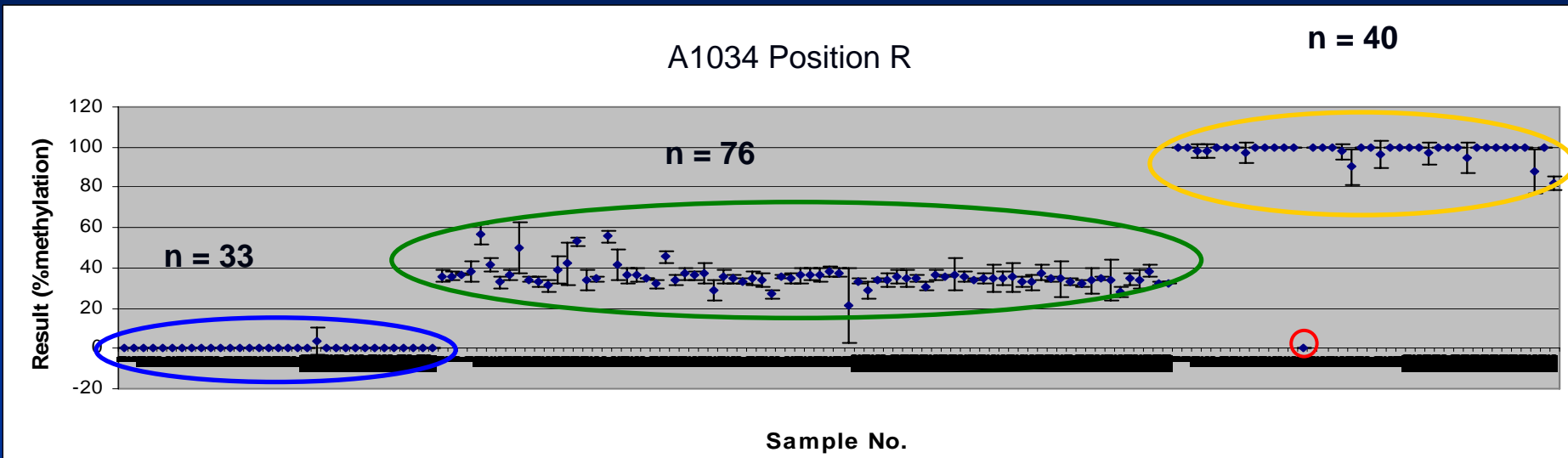
Principle of Pyrosequencing



Pyrograms

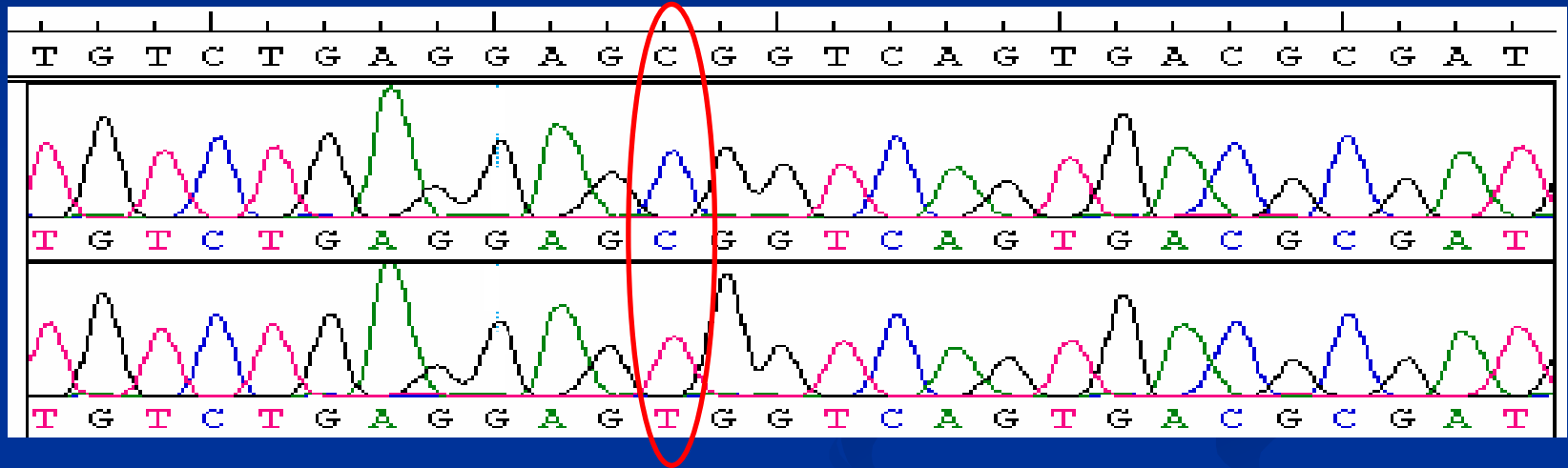


Pyrosequencing Results



Sample 58

Position R



Normal Sequence

Sample 58

PWS Bisulphited sequence

T G T T T G A G G A G C G G T T A G T G A C G C G A T

Sample 58 Bisulphited sequence

T G T T T G A G G A G T G G T T A G T G A C G C G A T

Advantages

- AQ software allows accurate quantification of methylation at multiple CpG sites within amplicon
- PCR primers independent of methylation state
- Confidence scores (passed, checked or failed) alert user to the quality of assay data
- Reference peaks incorporated into the analysis add confidence to data collection – also bisulphite treatment controls are included
- Results are presented in sequence context so sequence variants will be identified
- Assays relatively inexpensive & rapid
- Has potential to detect mosaicism

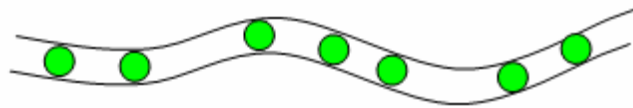
Disadvantages

- Further work is still necessary to establish the cause behind the abnormal methylation e.g. UPD, deletion or an imprinting mutation

High resolution melt curve analysis

High resolution melt curve analysis

Non saturating dsDNA binding dye
e.g. SYBR™ Green



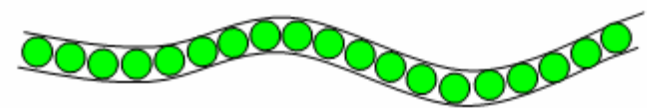
Melting



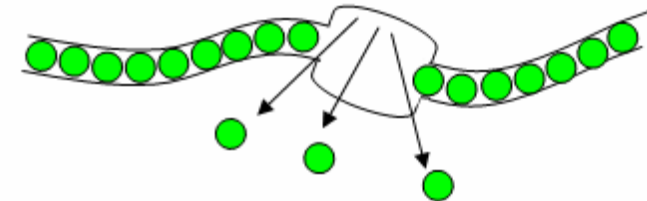
Dye molecules "jump" and redistribute
into molecule

No change in fluorescent signal

Saturating dsDNA binding dye
e.g. LCGreen Plus™



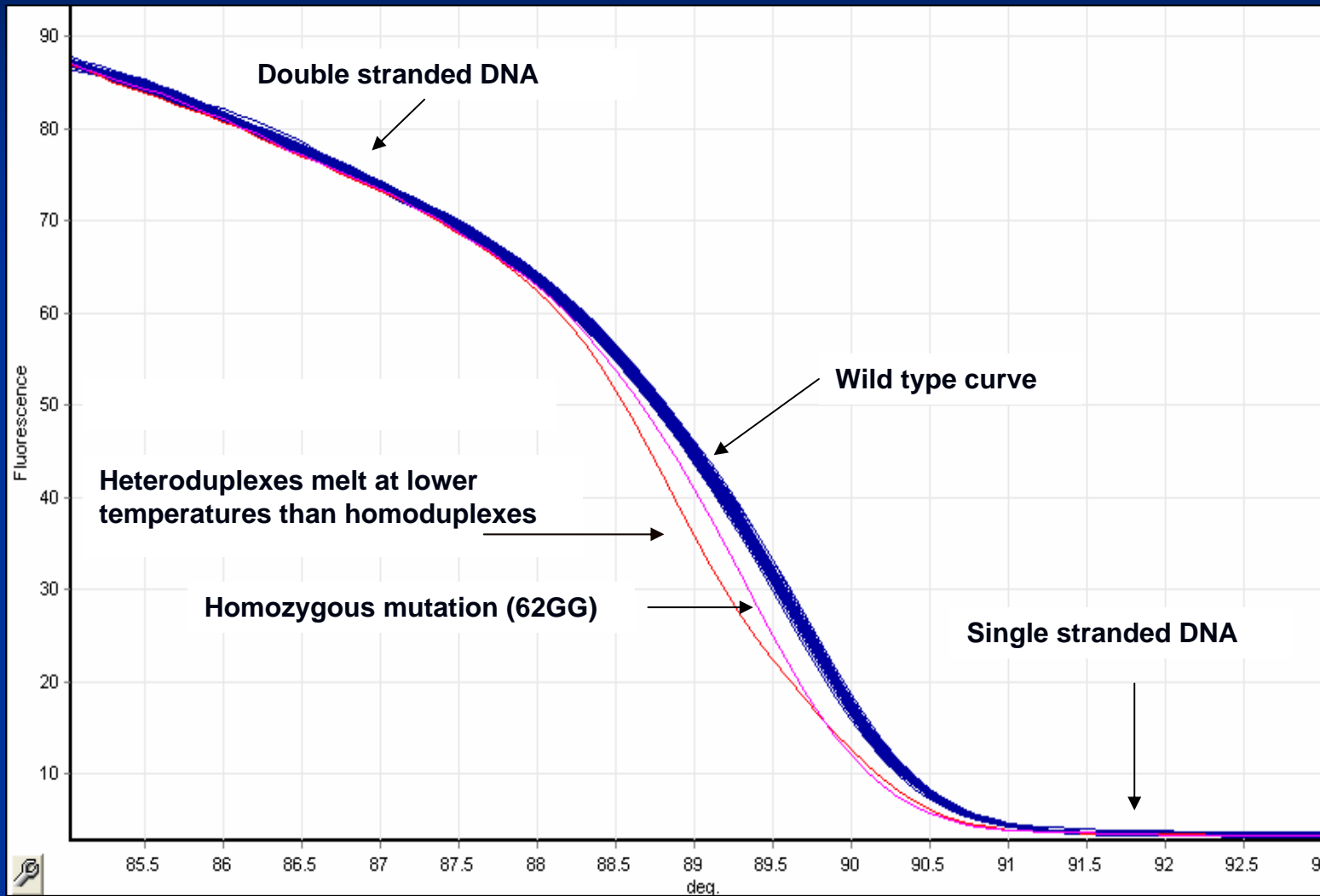
Melting



Dye molecules released

Decrease in fluorescent signal

High resolution melt curve analysis



NORMAL

AGGGAGTTGGGATTTTTGTATTGYGGTAAATAAGTAYGTTTG YGYGGTYGTAGAGGTAGGTTGGYGYGTATG
TTTAGGYGGGGATGTGTGYGAAGTTTGTGTTGTTGTAGYGAGTTTGGYGTAGAGTGGAGYGTGTGTGGAG
ATGTTTGAYGTATTTGTTT GAGGAGYGTAGTGA YGYGATGGAGYGGTAAGGTTAGTTGTGTGGTGGTT
TTTTTTAAGAGATAGTTTGGGG

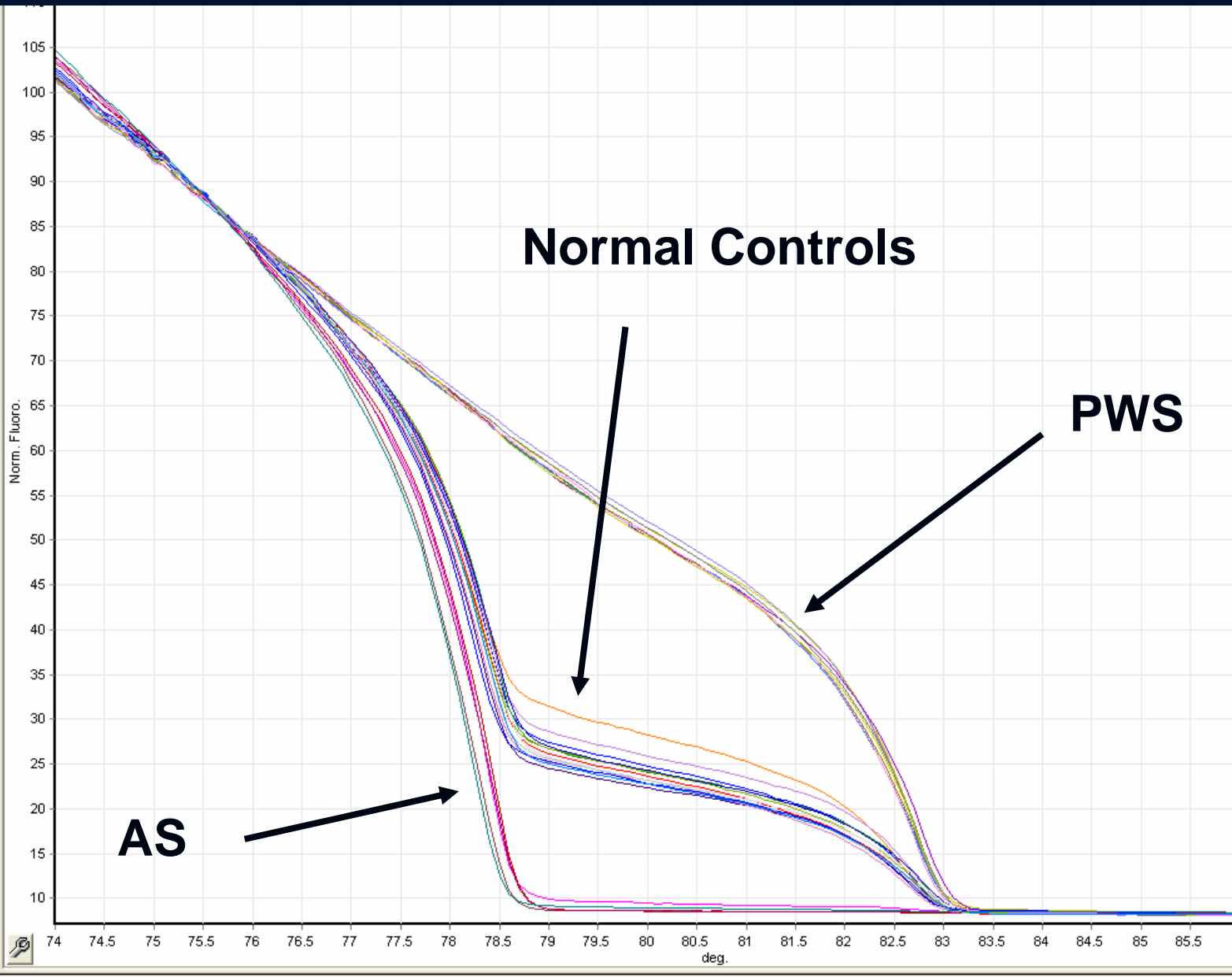
PWS

AGGGAGTTGGGATTTTTGTATTGCGGTAAATAAGTACGTTTGCGCGGTCTAGAGGTAGGTTGGCGCGTATG
TTTAGGCGGGGATGTGTGCGAAGTTTGTGTTGTTGTAGCGAGTTTGGCGTAGAGTGGAGCGGTCTCGGAG
ATGTTTGACGTATTTGTTT GAGGAGCGGTAGTGA CGCGATGGAGCGGGTAAGGTTAGTTGTGTCTCGGTGGTT
TTTTTTAAGAGATAGTTTGGGG

AS

AGGGAGTTGGGATTTTTGTATTGTGGTAAATAAGTATGTTTG TGTGGTTGTAGAGGTAGGTTGGTGTGTATG
TTTAGGTGGGGATGTGTGTGAAGTTTGTGTTGTTGTAGTGAGTTTGGTGTAGAGTGGAGTGTGTGTGGAG
ATGTTTGATGTATTTGTTT GAGGAGTGTAGTGA TGTGATGGAGTGGTAAGGTTAGTTGTGTGGTGGTT
TTTTTTAAGAGATAGTTTGGGG

21 sites can vary



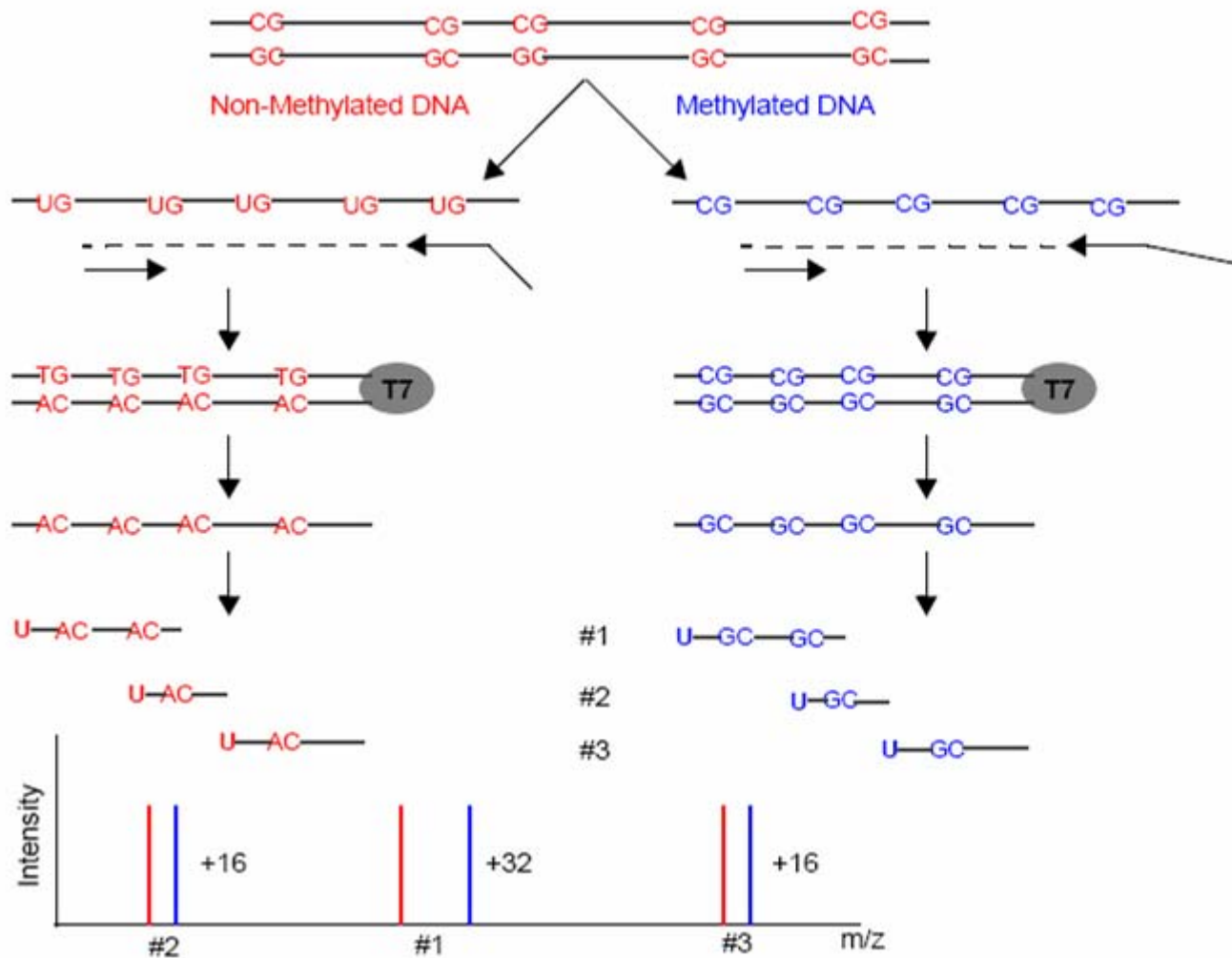
Advantages

- Closed tube method
- No post PCR processing and no separation step
 - improves analysis time
 - reduces contamination risk
- Rapid
- Inexpensive
 - requires the use of only PCR reagents and dsDNA binding dye
- PCR primers independent of methylation state
- Global information about methylation/sequence composition of amplicon

Disadvantages

- Further work is still necessary to establish the cause behind the abnormal methylation e.g. UPD, deletion or an imprinting mutation

Mass Spectrometry



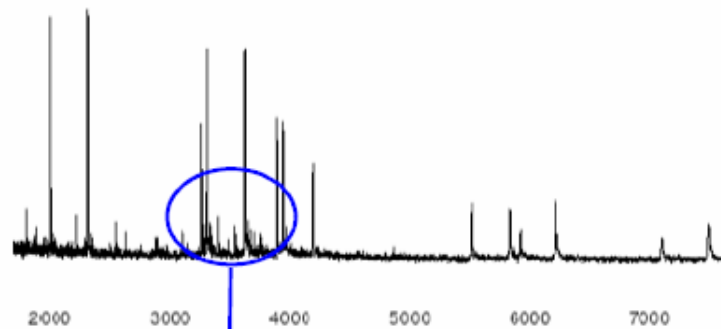
Bisulfite treatment of genomic DNA

PCR and SAP treatment

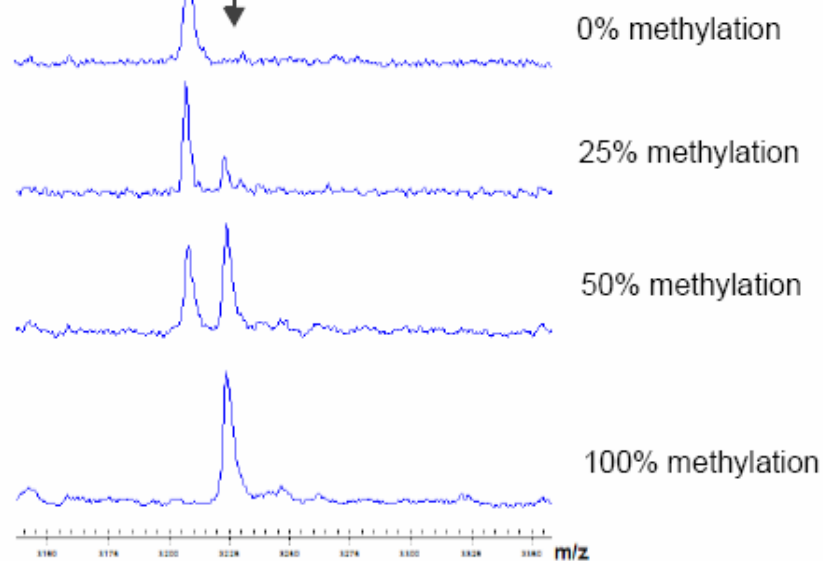
In vitro transcription

Uracil-specific cleavage

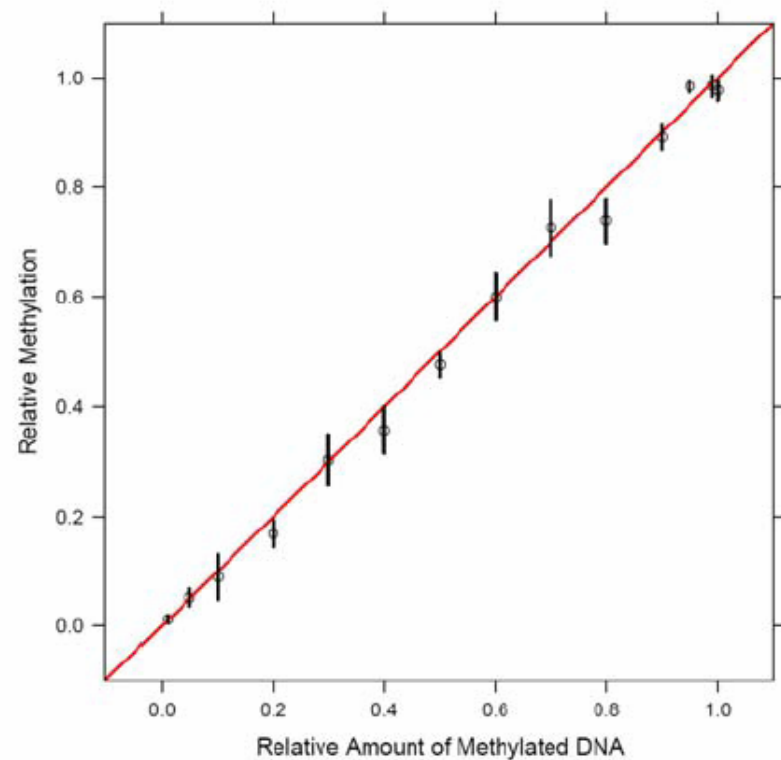
MALDI-TOF MS analysis produces signal pattern pairs indicating non-methylated and methylated DNA



NM (CpG 02) M (CpG 02)



The non-methylated (NM) DNA fragment (5'-AACAAACAAT-3') and the methylated (M) DNA fragment (5'-AACAAACGAT-3') show a mass difference of 16 Da.



Graph showing the relationship between the estimated relative methylation and the relative amount of methylated DNA spiked into the mixture.

Advantages

- PCR primers independent of methylation state
- Quantification of methylation (to 5%) at all CpG sites within amplicons up to 600bp
- Useful for large scale projects to identify 'useful' CpG sites
- High throughput
- High precision

Disadvantages

- Expensive
- Extensive post PCR handling – relies on success of multiple reactions
- Need access to equipment and specialised data analysis software

Quality control issues

❖ **Quality and quantity of DNA**

❖ **Bisulphite treatment**

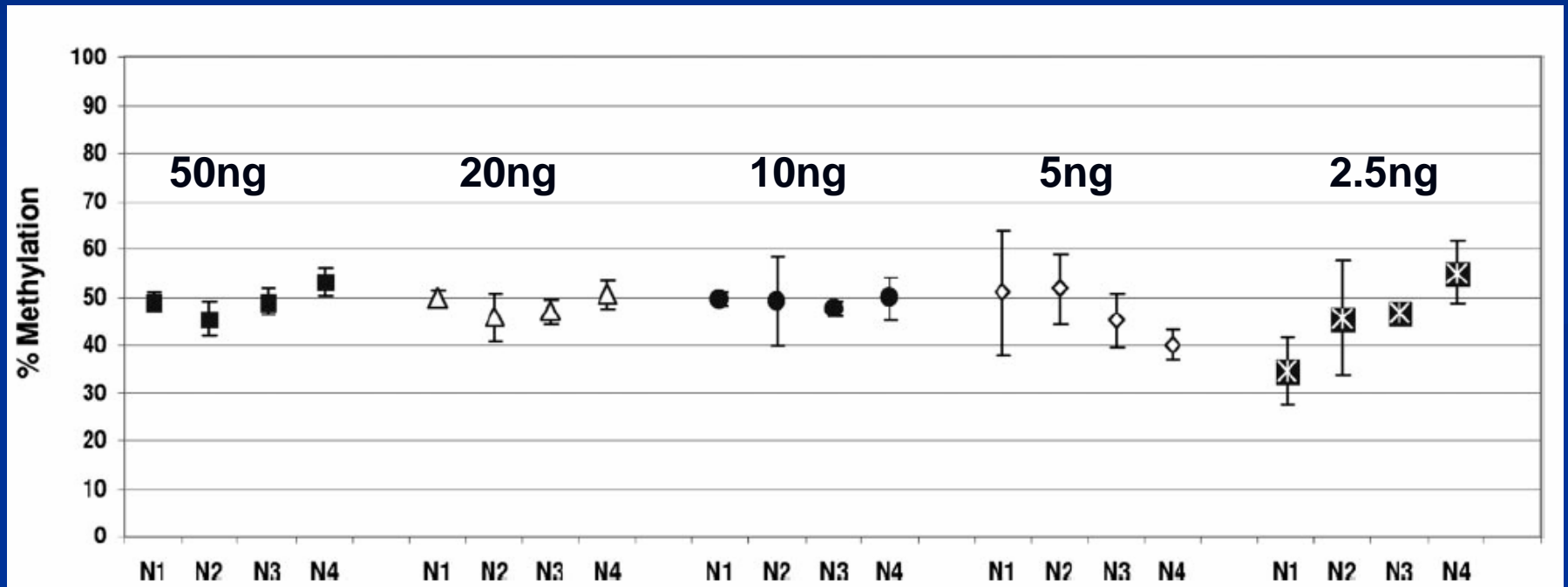
- **quality**

- **batches**

❖ **Preferential amplification**

❖ **Use of standard curves**

Quality and concentration of DNA

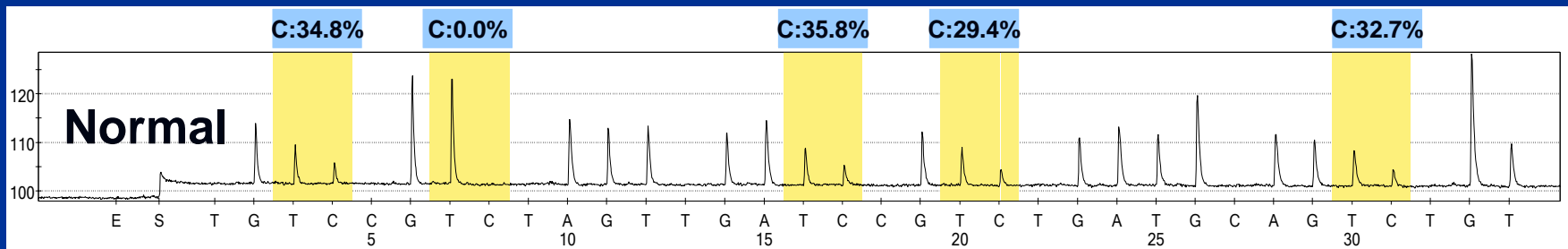


Bisulphite treatment

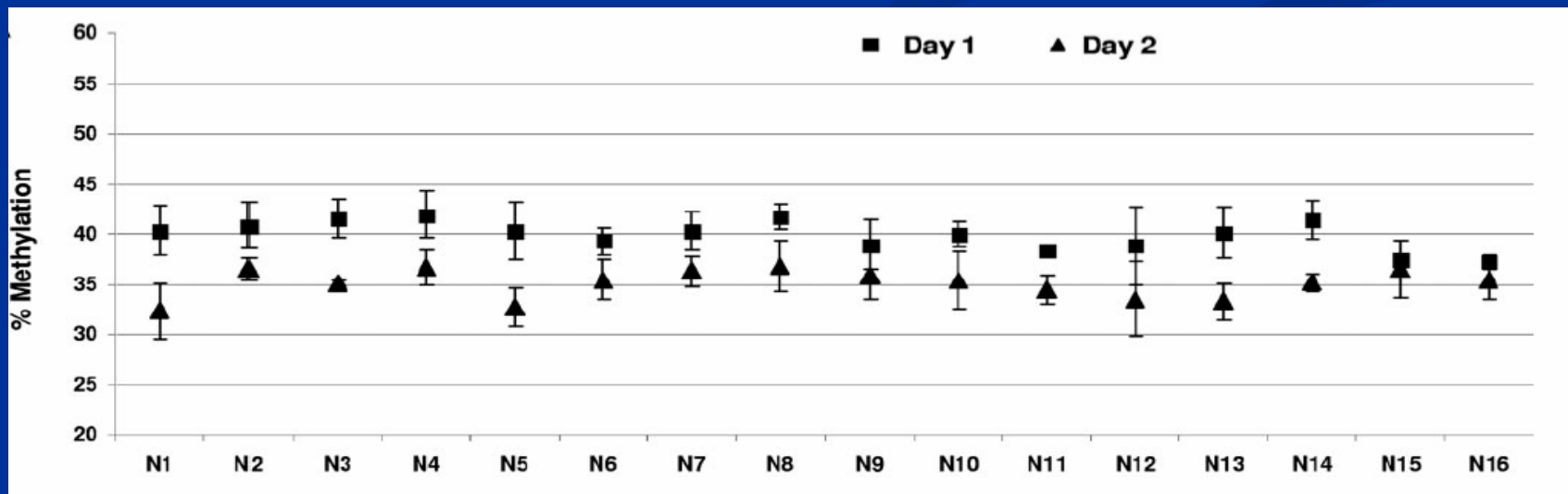
Total conversion

Pyrosequencing has in built control sites to monitor whether bisulphite conversion is complete

Most techniques do not have controls to monitor this



Batches



Preferential amplification and standard curves

