

**Evaluation of the Invader® assay  
platform for molecular analysis of the  
Factor V (G1691A) and Factor II  
(G20210A) mutations.**

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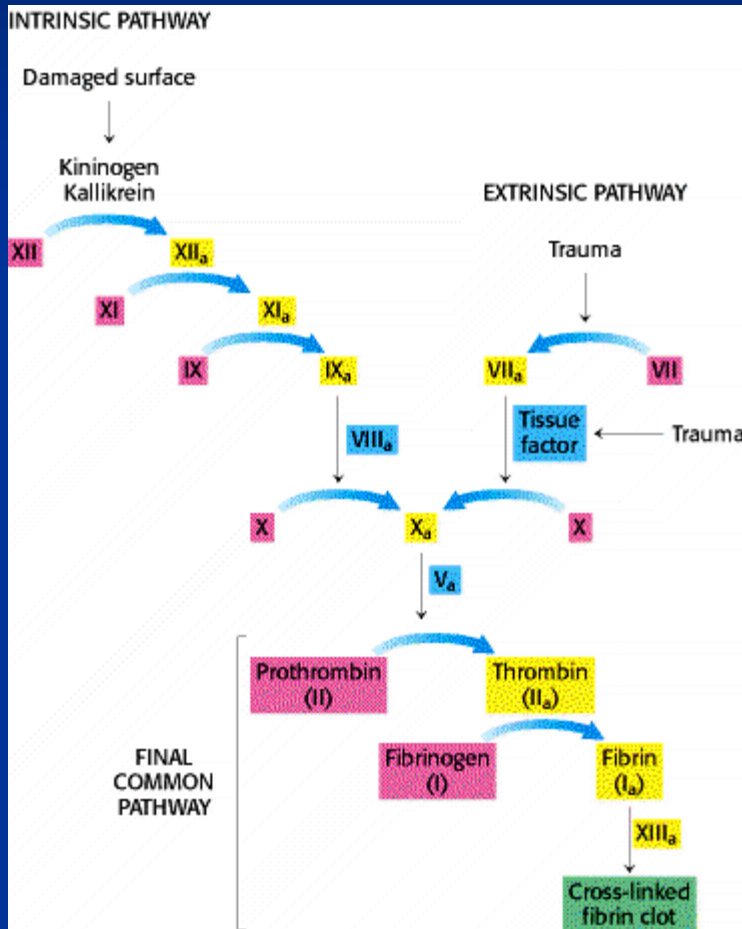
*Wessex Regional Genetics Laboratory*

*and*

*National Genetics Reference Laboratory.*

# Introduction:

## Blood clotting cascade:



- Activated Factor V (FV<sub>a</sub>) is involved in the conversion of prothrombin to thrombin.
- Thrombin then acts on fibrinogen to form fibrin.
- Fibrin forms the main constituent of the blood clot.

# Thrombophilia:

- Pathological activation of the clotting cascade leading to venous thromboembolism (VTE).
- Incidence = 1/ 1000.

## Clinical manifestations:

- Deep vein thrombosis (DVT) of the lower limbs and pulmonary embolism.

## Genetic, acquired and mixed risk factors for thrombosis:

Inherited	acquired	Mixed
Anti-thrombin deficiency	Age	Hyperhomocysteinemia
Protein C deficiency	Immobilisation	Increased Factor VII levels
Factor V Leiden	Surgery	Increased Factor XI levels
Factor II G20210A	Malignancy	Increased factor IX levels
	Pregnancy	Increased Fibrinogen levels
	Oral contraceptives	
	HRT	

## Genetic thrombophilia:

- Individuals with genetic thrombophilia show an increased RISK of recurrent thrombosis with an earlier onset (<45 yrs).

### Factor V (Proaccelerin):

- Factor V Leiden mutation (Arg506Glu) affects an activated protein C (APC) cleavage site and causes APC resistance.
- **Pathology** = lowered turnover of FVa and an increased conversion of prothrombin to thrombin.
  - Heterozygous = 5% of general population and 18% of thrombophilia patients. Increased risk of thrombosis = 2 to 7-fold higher
  - Homozygous mutations carriers have increased risk of 40-80 fold higher.

Cont:

## Factor II (prothrombin):

- The prothrombin mutation G20210A locates to the 3'-untranslated region and causes hyperthrombinemia.
  - It is present in 2% of the population and 7% of individuals with thrombosis.
  - FII (G20210A) is linked to a 2 to 5-fold increase in risk of thrombosis, homozygosity of the mutation is associated with a further increase in risk of thrombosis.

## Co-inheritance of FVL and FII mutations:

- Compound heterozygosity for FVL (G1691A) and the Factor II (G20210A) mutations occurs in 1/1000.
  - This is associated with a 20-fold increase in risk of thrombosis.

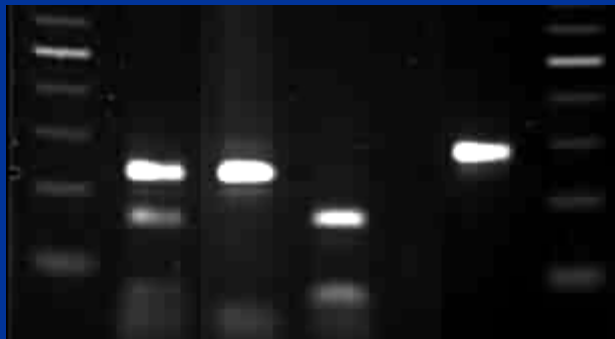
# Methodology:

## The RFLP method (PCR based):

### Factor V Leiden:

Restriction enzyme = Mnl I

(loss of restriction site)



Undigested  
No DNA  
Normal  
Homozygous  
Heterozygous

### Factor II:

Restriction enzyme = Hind III

(Use of mutagenic primer and G > A  
creates restriction site)



Undigested  
No DNA  
Normal  
Heterozygous  
Homozygous

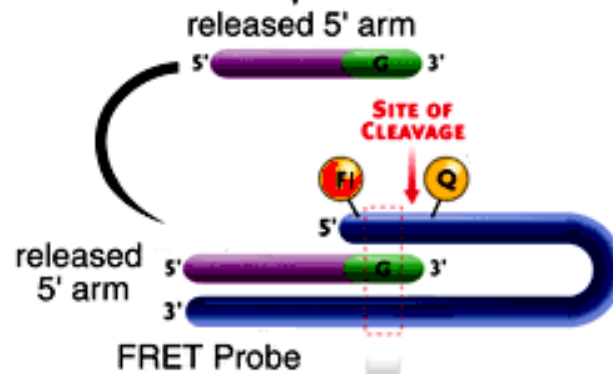
# The Invader<sup>®</sup> assay platform (non-PCR) :

Primary reaction

**Correct DNA structure forms:  
Signal detected**



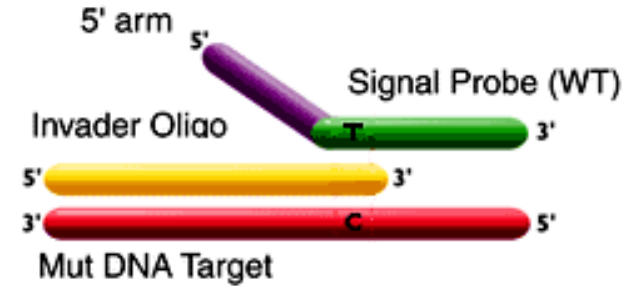
Result: Cleavage



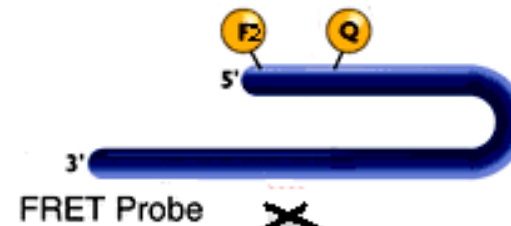
Result: Cleavage  
and signal

Fluorescent  
Signal

**Wrong DNA structure forms:  
No signal detected**



Result: No Cleavage  
No Signal



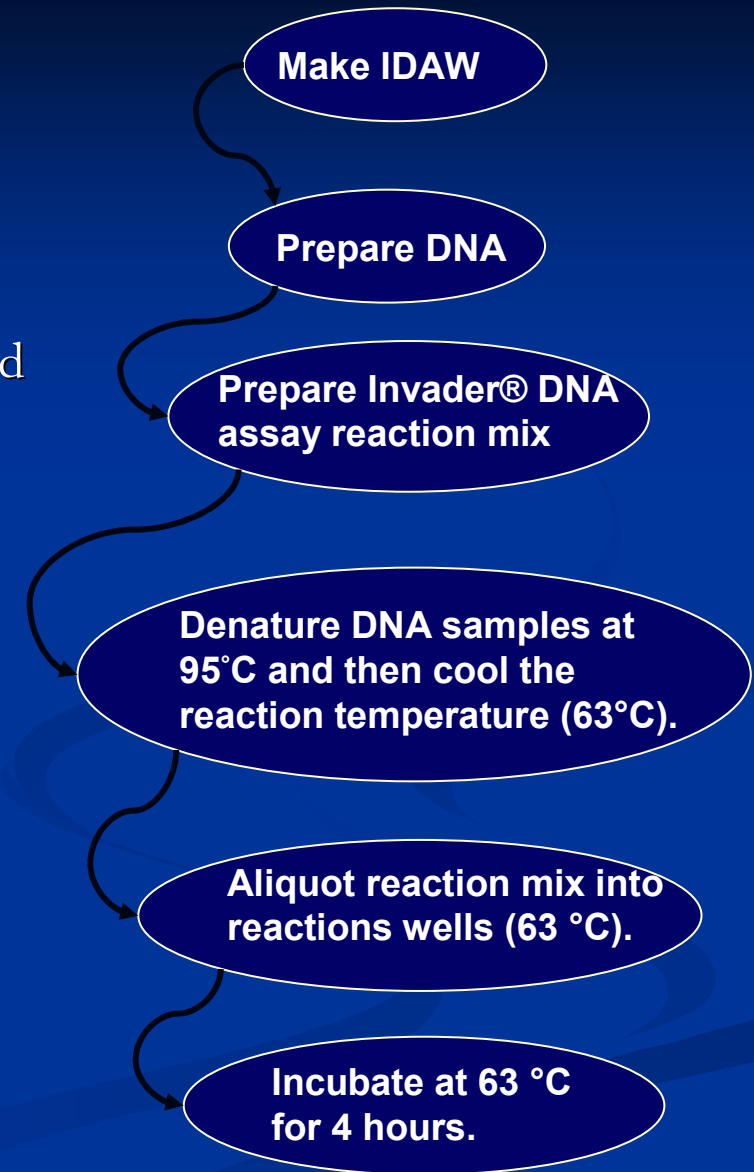
No Fluorescent  
Signal

Secondary reaction

## Invader® assay platform:

(Third Wave technologies)

- >100ng of DNA is needed
- 96 well plate format.
- biplex format F (WT) and R (Mut) signal
- Controls include a Normal, Heterozygote and Homozygote
- Fluorescence analysed using a 96 well plate reader (CytoFluor 96 well plate reader).
- An excel worksheet (IDAW) is used to calculate the net signal/ background or net fold over zero (FOZ).
- Ratio of the WT reaction to the mutant reaction.
  - Heterozygous =  $>0.3$  to  $<3$
  - Homozygous =  $<0.2$
  - Normal =  $>5$
- Total assay time =  $\sim 5$  hours  
(hands on time = 30-45min)



# Aim:

- To evaluate the Third Wave™ Invader® DNA assay for the detection of the FVL (G1691A) and Factor II (G20210A) mutations.
- Test 100 samples where the genotype is known for the FV and FII mutations as determined by the RFLP method.
- Compare and contrast the two methods for use in a diagnostic setting.

# Results:

## IDAW:

Data File Date Stamp:		Raw Data										
<b>F Signal (Mut)</b>												
	1	2	3	4	5	6	7	8	9	10	11	
A	259	211										
B	684	285										
C	909	232										
D	294	217										
E	349	199										
F	318	212										
G	1508	285										
H	352	417										
<b>R Signal (WT)</b>												
	1	2	3	4	5	6	7	8	9	10	11	
A	534	1678										
B	540	227										
C	245	2210										
D	253	911										
E	400	1026										
F	686	1174										
G	1374	1495										
H	1302	847										
<b>Lot Numbers:</b>												
Operator:	Date:	DNA Reaction Buffer 1 (B)						0	FVL (1691) FRET R Cassette (R)		0	FVL (1691) Control 1 (WT)
oly2	30/01/2004	FVL (1691) Invader Oligo (I)						0	FVL (1691) FRET F Cassette (F)		0	FVL (1691) Control 2 (Het)
		FVL (1691) Primary Probes (P)						0	Cleavase X Enzyme 40 ng/ul (E)		0	FVL (1691) Control 3 (Mut)
								0	Control 4 (No Target Blank)		0	
<b>Invader Data Analysis - FVL (G1691A) Biplex Assay</b>										Version 040202		
Sample	Invader Genotype	F Signal	R Signal	F Signal FOZ	R Signal FOZ	Net F Signal FOZ	Net R Signal FOZ	RATIO	Data	Action		
FVL (1691) Control 1	WT	259	534	0.88	2.11	0.04	1.11	27.767	VALID	NONE		
FVL (1691) Control 2	HET	684	540	2.33	2.13	1.33	1.13	0.855	VALID	NONE		
FVL (1691) Control 3	MUT	909	245	3.09	0.97	2.09	0.04	0.019	VALID	NONE		
Control 4		294	253						VALID	NONE		
66(4ul)	EQ1	349	400	1.19	1.58	0.19	0.58	3.106	INVALID	REPEAT SAMPLE		
6(4ul)	WT	318	686	1.08	2.71	0.08	1.71	20.965	VALID	NONE		
9	HET	1508	1374	5.13	5.43	4.13	4.43	1.073	VALID	NONE		
10	WT	352	1302	1.20	5.15	0.20	4.15	21.017	VALID	NONE		
11	WT	211	1678	0.72	6.63	0.04	5.63	140.810	VALID	NONE		
12	-	285	227	0.97	0.90	0.04	0.04	1.000	INVALID	REPEAT SAMPLE		
13	WT	232	2210	0.79	8.74	0.04	7.74	193.379	VALID	NONE		
19	WT	217	911	0.74	3.60	0.04	2.60	65.020	VALID	NONE		
15	WT	199	1026	0.68	4.06	0.04	3.06	76.383	VALID	NONE		
16	WT	212	1174	0.72	4.64	0.04	3.64	91.008	VALID	NONE		
17	WT	285	1495	0.97	5.91	0.04	4.91	122.727	VALID	NONE		
18	WT	417	847	1.42	3.35	0.42	2.35	5.612	VALID	NONE		

## Factor V and Factor II Invader results:

	<b>Factor V</b>	<b>Factor II</b>
<b>Number of samples</b>	110	110
<b>Normal</b>	71	98
<b>Heterozygous</b>	33	9
<b>Homozygous</b>	2	X
<b>% Concordance</b>	100%	100%
<b>% Repeated</b>	5.5%	3.6%
<b>% Fail</b>	3.6%	2.7%

# Discussion:

- The genotypes obtained from the Invader® assay showed 100% concordance to the RFLP method showing that it is suitable for use in diagnostic molecular genetics.

**Repeated samples:** Factor V 5.5% and Factor II 3.6%.

**Possible reasons:**

- Low signal as a result of low DNA concentration meaning the patient sample doesn't exceed the background fluorescence seen in the no target blank control.
  - when repeated with more DNA the correct genotype was obtained.

**Failed samples:** Factor V 3.6% and Factor II 2.7%.

- The samples that could not be genotyped were due to a low DNA concentration.

- 50 /110 samples tested were from Southampton Human genetics unit (total volume 10-15 $\mu$ l).
- All failed samples were from this source and could not be quantified or genotyped due to sample being depleted.

### Comparison of Invader and RFLP:

	<b>RFLP</b>	<b>Invader platform</b>
Total time	7-25 hours	4-5 hours
Hands on time	2-3 hours	<45 mins
Number of steps	8	4
	1: Prepare sample 2: Make master mix 3: Make digest mix 4: Add digest mix to samples 5: Restriction digest 6: Pour gel 7: Load gel 8: Image gel	1: Prepare sample 2: Make master mix 3: Add master mix to plate 4: Read plate
Analysis	Gel (subjectivity)	Excel spreadsheet (IDAW)
Visualisation	EtBr (mutagenic)	FRET (fluorescence)

## Cost per test:

Invader assay platform = £ 8 per result

RFLP = Approx. £ 2-3

## Other applications of the Invader assay platform:

- Cystic fibrosis testing
- Connexin 26
- MTHFR (methylenetetrahydrofolate reductase)
- ApoE
- Rett syndrome
- Prenatal chromosomal analysis

# Conclusion:

- The invader offers cheap rapid detection of SNP's
- Highly reproducible results with 100% concordance to existing methods.
- It is non-PCR based.
- Wide applications to other areas of molecular diagnostics.
- Highly dependant on template concentration as it affects the reaction dynamics and overall signal strength.
- More expensive than the existing RFLP method but is less labour intensive and is more rapid with results in  $\sim 5$  hours.

# Acknowledgements:

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